

ОДБОРУ ЗА НАУЧНОИСТРАЖИВАЧКУ ДЕЛАТНОСТ МЕДИЦИНСКОГ ФАКУЛТЕТА ВОЈНОМЕДИЦИНСКЕ АКАДЕМИЈЕ УНИВЕРЗИТЕТА ОДБРАНЕ У БЕОГРАДУ

На 111. седници Наставно-научног већа одржаној 28.09.2023. године, покренут је поступак за избор др сц. мед. Александра Пантовића, из Клинике за неурологију Војномедицинске академије, у звање **научни сарадник**. На овој седници, Наставно-научно веће је именovalo Комисију за оцену испуњености услова за избор у звање.

На основу приложене документације о научно-истраживачком раду, као и увида у целокупни рад кандидата, а у складу са Законом о науци и истраживањима („Службени гласник РС”, број 49/2019-3) и Правилником о стицању истраживачких и научних звања (159/2020-82), Одбору за научноистраживачку делатност Медицинског факултета Војномедицинске академије Универзитета одбране у Београду подносимо следећи

ИЗВЕШТАЈ

1. БИОГРАФСКИ ПОДАЦИ

Др Александар Пантовић је рођен 27.09.1982. године у Бару. Дипломирао је на Медицинском факултету у Београду 2008. године са просечном оценом 9.17. Након основних студија медицине био је запослен на Медицинском факултету у Београду у лабораторији за молекуларну медицину Института за микробиологију и имунологију. Радио је на пројектима Министарства просвете науке и технолошког развоја Републике Србије: Имуномодулаторно, цитопротективно и цитотоксично дејство наночестица (број пројекта 145073) и Модулација унутарћелијског енергетског баланса – контрола сигналних путева у терапији тумора и неуро-имуно-ендокриних поремећаја (број пројекта 41025). Специјалистички испит из неурологије положио је у децембру 2016. године на Клиници за неурологију Војномедицинске академије. Докторску дисертацију под називом „Улога аденозин-монофосфатом активираних протеинкиназа и mTOR комплекса 1 у *in vitro* цитотоксичном дејству нестероидних анти-инфламаторних лекова на ћелије глиома“ одбранио је у септембру 2020. године. Др Александар Пантовић је запослен на Клиници за неурологију Војномедицинске академије. Члан је Друштва неуролога Србије, Друштва за неуронауке Србије, Међународног удружења за неуроимунологију, Међународног удружења за невољне покрете и Интернационално удружења против главобоља. Ожењен је и отац две ћерке.

2. БИБЛИОГРАФСКИ ПОДАЦИ

2.1 Објављени радови

Поред сваке публикације унесена је позиција на листи часописа и његов импакт фактор из одговарајуће дисциплине са годином у којој је био најповољнији (за период од две године пре публикавања и година публикавања, и то за ону годину у којој је часопис најбоље рангиран, односно ону у којој је имао највећи импакт фактор). Након тога означен је укупан број бодова (ББ) и на крају нормиран број бодова (НББ) и формула која је коришћена за израчунавање према броју коаутора.

Рад у међународном часопису изузетне вредности (M21a):

1. Markovic Z, Harhaji-Trajkovic L, Todorovic-Markovic B, Kepić D, Arsikin K, Jovanović S, **Pantovic A**, Dramićanin M, Trajkovic V. In vitro comparison of the photothermal anticancer activity of graphene nanoparticles and carbon nanotubes. *Biomaterials*. 2011;32(4):1121-9. IF 7.404 (10 бодова) $HBB: 10/(1+0,2*(9-7)) = 7.142$ Biomedical, Multidisciplinary

Рад у врхунском међународном часопису (M21):

2. **Pantovic A**, Bosnjak M, Arsikin K, Kostic M, Mandic M, Ristic B, Tosic J, Suzin-Zivkovic V, Grujicic D, Isakovic A, Trajkovic V, Harhaji-Trajkovic Lj. In vitro antiglioma action of indomethacin is mediated through AMPK/mTOR complex 1 signaling pathway. *Int J Biochem Cell Biol*. 2017;83:84-96. IF 3.905 (8 бодова) $HBB: 8/(1+0,2*(12-7)) = 4$ Molecular biology & Neurosciences
3. **Pantovic A**, Krstic A, Janjetovic K, Kocic J, Harhaji-Trajkovic L, Bugarski D, Trajkovic V. Coordinated time-dependent modulation of AMPK/Akt/mTOR signaling and autophagy controls osteogenic differentiation of human mesenchymal stem cells. *Bone*. 2013;52(1):524-31. IF 4.461 (8 бодова) *Metabolism & Endocrinology*
4. Wasik AM, Grabarek J, **Pantovic A**, Ciešlar-Pobuda A, Asgari HR, Bundgaard-Nielsen C, Rafat M, Dixon IM, Ghavami S, Łos MJ. Reprogramming and carcinogenesis-parallels and distinctions. *Int Rev Cell Mol Biol*. 2014;308:167-203. Review. IF 4.973 (8 бодова) $HBB: 8/(1+0,2*(10-7)) = 5$ Biochemistry & Molecular medicine
5. Harhaji-Trajkovic L, Arsikin K, Kravic-Stevovic T, Petricevic S, Tovilovic G, **Pantovic A**, Zogovic N, Ristic B, Janjetovic K, Bumbasirevic V, Trajkovic VS. Chloroquine-mediated lysosomal dysfunction enhances the anticancer effect of nutrient deprivation. *Pharmaceutical Research*. 2012;29(8):2249-63. IF 4.742 (8 бодова) $HBB: 8/(1+0,2*(11-7)) = 4.44$ Pharmacology & Pharmacy, Multidisciplinary
6. Pantic I, Harhaji-Trajkovic L, **Pantovic A**, Milosevic NT, Trajkovic V. Changes in fractal dimension and lacunarity as early markers of UV-induced apoptosis. *J Theor Biol*. 2012;303:87-92. IF 2.351 (8 бодова) *Biology*
7. Trpkovic A, Todorovic-Markovic B, Kleut D, Misirkic M, Janjetovic K, Vucicevic L, **Pantovic A**, Jovanovic S, Dramicanin M, Markovic Z, Trajkovic V. Oxidative stress-mediated hemolytic activity of solvent exchange-prepared fullerene (C60) nanoparticles. *Nanotechnology*. 2010;21(37):375102. IF 3.652 (8 бодова) $HBB: 8/(1+0,2*(11-7)) = 4.4$ Multidisciplinary

Рад у истакнутом међународном часопису (M22):

8. **Pantović A**, Lepic T, Pasovski V, Krsmanovic Z, Raicevic R. Artery of Percheron infarction associated with COVID-19 in the young adult. *J Neurovirol*. 2021 Dec;27(6):951-953. IF 3.739 (5 бодова) *Neurosciences*
9. Sumarac-Dumanovic M, Jeremic D, **Pantovic A**, Janjetovic K, Stamenkovic-Pejkovic D, Cvijovic G, Stevanovic D, Micic D, Trajkovic V. Therapeutic improvement of glucoregulation in newly diagnosed type 2 diabetes patients is associated with a reduction of IL-17 levels. *Immunobiology*. 2013;218(8):1113-8. IF 3.180 (5 бодова) $HBB: 5/(1+0,2*(9-7)) = 3.57$ Immunology

Рад у међународном часопису (M23):

10. **Pantovic A**, Arsikin K, Kosic M, Ristic B, Trajkovic V, Harhaji-Trajkovic L. Data supporting the inability of indomethacin to induce autophagy in U251 glioma cells. Data Brief. 2017;10;11:225-230. IF 0.736 (3 бода) Multidisciplinary

Рад у националном часопису (M53):

11. **Pantović A**, Barjaktarević D, Kostić S, Dinčić E. Spazam konvergencije: između prave i lažne slabosti okulogira – prikaz slučaja. Sinapsa - publikacija Društva mladih neurologa Srbije, jun 2013. (1 бод) Неврологија
12. **Pantović A**, Jovanovski A, Raičević R, Dinčić E. Siringomijelija u longitudinalnom ekstenzivnom transverzalnom mijelitisu kod bolesnika sa oboljenjem iz spektra neuromijelitis optika. Sinapsa - publikacija Društva mladih neurologa Srbije, decembar 2017. (1 бод) Неврологија

Предавање по позиву на међународном скупу (M32):

13. 5th Symposium of regenerative medicine “Stem cells - a vision beyond borders”, Belgrade novembar 2018. Tema: Stem cell application in neurological disorders (1.5 бодова)

Саопштење са међународног скупа штампано у изводу (M34):

14. MDS- ES Winter School for Young Neurologists, Belgrade 2014. Dystonic or pyramidal sign in a patient with clinical presentation of Parkinson disease (video prikaz) (0.5 бодова)
15. Adriatic neurology forum, Опатија 2015. Optical coherence tomography in the assessment of structural with functional and cognitive changes in multiple sclerosis (0.5 бодова)
16. Adriatic neurology forum, Београд 2016. Siringomyelic presentation of Longitudinal Extensive Transverse Myelitis as first manifestation of Sistemik Lupus Eritematosus and NMO spectrum disorder – a case report (0.5 бодова)
17. International Society of Neuroimmunology congress, Јерусалим 2016. Gene polymorphisms as modifiers of response to interferon beta 1b in Serbian Multiple Sclerosis population (candidate gene study) (0.5 бодова)
18. International Society of Neuroimmunology congress, Јерусалим 2016. Angiotensin-converting enzyme and angiotensin receptors gene polymorphisms as risk factors for multiple sclerosis (0.5 бодова)

Саопштење са скупа националног значаја штампано у изводу (M64):

19. Конгрес неуролога и неуронаука Србије Београд, 2013. Спазам конвергенције – приказ случаја (0.2 бода)

20. Конгрес неуролога и неуронаука Србије Београд, 2013. Повезаност СНП у генима за ренин-ангиотензин систем са ризиком за настанак мултипле склерозе (0.2 бода)
21. Конгрес неуролога и неуронаука Србије Београд, 2013. Indomethacin exerts in vitro antiglioma effect through modulation of AMPK/mTOR signaling pathway (0.2 бода)
22. Конгрес неурофизиолога Србије 2015. Retinal nerve fiber layer thickness and multifocal visual evoked potentials in optic neuritis associated with multiple sclerosis – A case study (0.2 бода)
23. Конгрес неуролога и неуронаука Србије Нови сад, 2015. Оптичка кохерентна томографија у процени структурних са функционалним и когнитивним променама код пацијента са мултиплом склерозом (0.2 бода)
24. Школа за младе неурологе – Палић 2015. Мултипла склероза на путу до дијагнозе неуролупуса (0.2 бода) (0.2 бода)
25. Школа за младе неурологе – Палић 2015. Неуролупус/Неуромијелитис оптика спектар – диференцијална дијагноза (0.2 бода)
26. Конгрес неуролога и неуронаука Србије Нови сад, 2015. Мултифокални визуелни евоцирани потенцијали и структурне промене на мрежњачи код болесника са лонгитудиналним екстензивним трансферзалним мијелитисом и IgG аквапоринским антителима – НМО, приказ случаја (0.2 бода)
27. Семинар практичног лекара, Игало, 2019. Тумерфактивна мултипла склероза (0.2 бода)
28. Симпозијум Covid-19 и неурологија Врдник, 2021. Биталамички инфаркт повезан са COVID-19 приказ случаја младог одрасог пацијента (0.2 бода)

Одбрањена докторска дисертација (М70):

29. Uloga adenosin-monofosfatom aktivirane protein-kinaze i mTOR kompleksa 1 u in vitro citotoksičnom dejstvu nesteroidnih anti-inflamatornih lekova na ćelije glioma. Медицински факултет, Универзитет у Београду, септембар 2020. (6 бодова)

Табела 1. Преглед публикација др сц. мед. Александра Пантовића од почетка научног и клиничког рада по категоријама, вредности резултата (укупно и нормирано) и импакт фактор.

КАТЕГОРИЈА НАУЧНЕ ПУБЛИКАЦИЈЕ	М	БРОЈ РАДОВА	УКУПАН БРОЈ ПОЕНА	НОРМИРАНИ БРОЈ ПОЕНА	ИМПАКТ ФАКТОР
Рад у међународном часопису	M21	7	58	40.98	31.934
	M22	2	10	8.57	6.919
	M23	1	3	3	0.736
Predavanje po pozivu sa međunarodnog skupa štampano u izvodu	M32	1	1.5	1,5	/
Saopštenje sa međunarodnog skupa štampano u izvodu	M34	5	2.5	2.5	/
Рад у научном часопису	M53	2	2	2	/
Саопштење са скупа националног значаја штампано у изводу	M64	10	2	2	/
Одбрањена докторска дисертација	M71	1	6	6	/
УКУПНО		29	85	66.55	39.583

3. АНАЛИЗА РАДОВА

(радови штампани у целини, публиковани од почетка каријере)

Објављени радови др сц. мед. Александра Пантовића у истраживачком делу дају објашњења за молекуларне механизме цитотоксичних ефеката нестероидних анти-инфламаторних лекова на ћелије глиома. Посебно се истражује улога сигналних путева који контролишу процесе апоптозе и аутофагије у цитотоксичном деловању ових лекова на ћелије тумора централног нервног система. Истраживања која се односе на активацију процеса аутофагије у контроли диференцијације мезенхимских матичних ћелија су по први пут објављена у раду др Пантовића а ова публикација је цитирана преко 200 пута (референца 3). Након отпочињања специјализације из неурологије и након тога др Пантовић је објавио неколико клиничких случајева од којих посебан значај има приказ акутног исемијског можданог удара код младог пацијента у сливу Перчеронове артерије истовремено са оболевањем од Covid-19.

Из радова под називом „In vitro antiglioma action of indomethacin is mediated through AMPK/mTOR complex 1 signaling pathway“ (референца 2) и „Data supporting the inability of indomethacin to induce autophagy in U251 glioma cells“ (референца 10) коршћен је највећи број резултата за докторску дисертацију кандидата. У поменутих радовима и дисертацији др Александар Пантовић износи податке о примарним туморима централног нервног система глиомима са посебним освртом на најчешћи малигни тумор централног нервног система глиобластом. У уводном делу наведени су подаци о доказима за антитуморско деловање нестероидних анти-инфламаторних лекова (НСАИЛ). Кандидат у уводном делу цитира и

основне неурофармаколошке карактеристика нестероидног анти-инфламаторног лека индометацина, који је у истраживању показао најизраженији цитотоксични ефекат и једини од испитиваних лекова утицао на активацију AMPK молекула. Описана је улога сигналног пута AMPK/mTOR и модел mTORC1- зависне инхибиције АКТ механизмима повратне спреге. Такође описан је молекуларни механизам апоптозе, програмиране ћелијске смрти типа I и аутофагије, катаболичког процеса који омогућује ћелијску деградацију сопствених протеина и органела у аутофаголизозомима.

Имајући у виду улогу AMPK у инхибицији mTOR и индукцији апоптозе и аутофагије, претпостављено је да НСАИЛ своје антиглиомско деловање остварују модулацијом AMPK/mTORC1 сигналног пута. Да би ова претпоставка била испитана постављени су следећи циљеви овог истраживања: (1) испитати утицај НСАИЛ на вијабилитет ћелија хумане ћелијске линије глиома U251 и примарних туморских ћелија изолованих из узорака ткива пацијената оболелих од глиобластома (2) испитати тип и механизме ћелијске смрти индуковане НСАИЛ у ћелијама глиома (3) испитати утицај НСАИЛ на активацију AMPK/mTOR сигналног пута у ћелијама глиома (4) испитати улогу AMPK/mTOR сигналног пута у индукцији ћелијске смрти индуковане НСАИЛ у ћелијама глиома.

У делу о материјалу и методама, др Пантовић описује експерименталне методе и услове под којима су експерименти изведени:

(а) Гајење хумане ћелијске линије глиобластома U251 као и изолацију примарних туморских ћелија изолованих из узорака ткива пацијената оболелих од глиома градуса IV. На Клиници за неурохирургију Клиничког центра Србије у Београду постављена је дијагноза основног обољења и узети су узорци ткива за изолацију примарне културе. Истраживање је спроведено у складу са Хелсиншком Декларацијом и одобрено од стране Етичког комитета Клиничког центра Србије и Етичког комитета Медицинског факултета Универзитета у Београду. Сви оболели су дали пристанак у писаној форми за учешће у истраживању и анализу узорака у дијагностичке и научне сврхе. Поглавље материјал и методе је подељено у следеће целине:

(б) Одређивање вијабилитета ћелија мерењем активности ћелијског ензима лактат дехидрогеназе, мерењем активности митохондријалних дехидрогеназа и кристал виолет тестом.

(е) Испитивање улоге AMPK/mTORC1 сигналног пута у цитотоксичној активности третираних ћелија мерењем вијабилности ћелија код којих је експресија AMPK инхибирана РНК интерференцијом, као и након третмана ћелија активатором mTORC1 леуцином. Такође, у циљу доказивања улоге AMPK/mTORC1 сигналног пута испитивано је да ли фармаколошки активатори AMPK метформин и AICAR попут индометацина остварују антиглиомске ефекте модулацијом овог сигналног пута.

(ф) Утврђивање типа и механизма ћелијске смрти. За ову сврху коришћена је проточна цитофлуориметрија за анализу апоптотске фрагментације ДНК, транслокације фосфатидилсерина, активације каспаза, продукције супероксида, промене потенцијала митохондријалне мембране, ацидификације ћелијске цитоплазме. Real-time RT-PCR метода је коришћена за анализу експресије инхибитора циклин зависне киназе p21. Имуноблот метода за испитивање експресије или активације сигналних молекула који учествују у регулацији енергетске хомеостазе, ћелијске пролиферације, апоптозе и аутофагије (AMPK α , ACC, Akt, Raptor, mTOR, S6K, PRAS40, каспазу-3, PARP, LC3, beclin-1).

(г) Утврђивање механизма ативације AMPK. Садржај AMP и ATP у ћелијама линије U251 је анализиран градијентном течном хроматографијом високих перформанси (HPLC) и биолуминисцентним есејем.

(х) Статистичке методе коришћене за обраду добијених података.

У резултатима кандидат јасно и документовано приказује добијене експерименталне податке о антиглиомском ефекту НСАИЛ на ћелије глиома U251 линије и примарне културе ћелија глиома градуса IV (глиобластома).

Први резултати експеримената су показали да индометацин снажније од других COX инхибитора, диклофенака, напроксена и кетопрофена, смањује вијабилитет U251 ћелија

хуманог глиобластома. Такође у истраживању кандидата показано је да једино индометацин од коришћених нестероидних анти-инфламаторних лекова статистички значајано активира AMPK. Третман индометацином који је надаље коришћен у истраживању довео је до морфолошких промена на ћелијама U251 линије које су карактеристичне за апоптозу. У ћелијама које су третиране индометацином показане су промене у степену продукције реактивних кисеоничних врста, деполаризацији митохондрија, транслокацији фосфатидилсерина на површину ћелијске мембране, ДНК фрагментацији, активацији каспаза и PARP. Цитотоксичном ефекту индометацина претходило је повећање експресије тумор супресорског протеина p21 и застој ћелијског циклуса у G2M фази. Приказни резултати директно доводе у везу улогу сигналног пута AMPK/mTORC1 са антиглиомским деловањем индометацина. Наиме, показано је да индометацин повећава фосфорилацију AMPK и нисходних молекула које ензим фосфорилише као што су Raptor и ацетил-CoA карбоксилаза (ACC). Активацију AMPK прати смањење фосфорилације mTOR и молекула чију активност mTORC1 регулише, као што су рибозомална p70S6 киназа (S6K) и PRAS40 (Ser183). Генетска инхибиција експресије AMPK молекула РНК интерференцијом, као и третман ћелија активатором mTOR1 леуцином доведе до делимичног поништавања цитотоксичних ефеката изазваних индометацином. Са друге стране, третирање U251 ћелија инхибитором mTOR-а рапамицином смањује број живих ћелија у култури. Осим тога, фармаколошки активатори AMPK метформин и AICAR попут индометацина испољавају антиглиомски ефекат инхибицијом mTORC1. Истражујући механизме којима индометацин активира AMPK измерено је смањење ћелијског ATP и пораст AMP/ATP односа у третираним ћелијама док повезаност фосфорилације AMPK са инхибицијом COX и повећањем интраћелијског нивоа калцијума није нађена.

Резултати показују да је цитотоксични ефекат индометацина на ћелије примарне културе глиобластома такође посредован активацијом AMPK/Raptor/ACC и инхибицијом mTORC1/S6K сигналних молекула. Коначно, истраживана је и способност индометацина да индукује аутофагију у ћелијској линији глиобластома U251. Међутим, није детектовано повећање маркера аутофагије као нити повећање цитоплазматских везикула са киселим садржајем, експресија beclin-1 и конверзија лаког ланца 3-I (LC3-I) у аутофагозомну форму LC3-II у присуству лизозомалних инхибитора. Показано је да генетска и фармаколошка инхибиција аутофагије не утичу на цитотоксични ефекат индометацина, што је потврдило да је антиглиомски ефекат лека независан од аутофагије.

У дискусији резултата кандидат је на систематичан начин упоредио резултате приказане у овој публикацији са резултатима других истраживача. На основу тога, кандидат је изнео своје закључке и хипотезе, који објашњавају резултате ове публикације у складу са резултатима већине других аутора који су имали исто истраживачко интересовање.

У поглављу закључци су сумирани сви резултати добијени у овом истраживању који у потпуности одговарају на циљеве постављене на почетку.

У најцитиранијој публикацији на којој кандидат партиципира као коаутор (**референца 1**) поређена је фототермална антиканцерска активност графенских наночестица и угљеничних наночестица побуђених спектром који је близак инфрацрвеној светлости. Упркос нижем капацитету апсорбовања графенских наночестица оне су генерисале више топлоте од угљеничних наночестица растворених под одговарајућим условима. Механизми фототермалног убијања канцерских ћелија посредованог графеном су укључивали оксидативни стрес и деполаризацију митохондријалне мембране што је доводило до мешовите апоптотске и некротичне ћелијске смрти.

У наредном истраживању (**референца 3**) аутори су процењивали улогу протеин киназе активираних од AMP (AMPK), молекула Akt, mTOR и аутофагију као и интеракцију наведених молекула у диференцијацији мезенхимских матичних ћелија. Добијени подаци указују да AMPK контролише диференцијацију хумане мезенхимске матичне ћелије кроз рану аутофагију посредовану инхибицијом mTOR-а и касну активацију Akt/mTOR сигналне осе.

У ревијалном раду у којем је др Пантовић учествовао као део међународне групе истраживача (**референца 4**) дата су разматрања о употреби четири транскрипциона фактора, Oct3/4, Sox2, c-Myc и Klf4 –тзв. "Јаманака фактори" у конверзији диференцираних ћелија, назад

у плурипотентни/ембрионални стадијум. Овако “репрограмирани” ћелије отворају велико поље могућности примене укључујући и неурорегенерацију али и етичку дилему медицинске употребе због потенцијала за малигу трансформацију.

Истраживана је способност хлорокина, лизозомског инхибитора аутофагије, како би се побољшао ефекат против малигних ћелија које су изглађиване у *in vitro* условима (референца 5). Хлорокин је брзо убио малигне ћелије механизмом независним од аутофагије. Нађено је да су основни механизми цитотоксичности акумулација лизозома изазвана хлорокином, оксидативни стрес, митохондријална деполаризација, активација каспазе и мешана апоптотска/некротична ћелијска смрт ћелије. Хлорокин је инхибирао и раст меланома мишева са ограниченим уносом калорија сличним механизмом. Закључено је да комбиновани третман са хлорокином и калоријском рестрикцијом може бити користан у терапији канцера.

Циљ наредног истраживања (референца 6) у којем је др Пантовић учествовао као коаутор био је да се примени фрактална анализа за процену ултраструктурних промена у раним фазама апоптозе. Апоптоза је индукована у U251 ћелијској линији хуманог глиома излагањем ултравиолетном (УВ) спектру светлости. Ћелије су визуелизоване оптичком фазно-контрастном микроскопијом и фотографисане пре УВ третмана, непосредно након третмана, као и у интервалима од 30 мин током периода посматрања од 5 сати. За сваку од 32 анализираних ћелије, ћелијска и нуклеарна фрактална димензија, као и нуклеарна лакуарност, одређене су у свакој временској тачки. Наши подаци показују да ћелијска ултраструктурна сложеност одређена фракталном димензијом и лакуарношћу значајно опада након УВ зрачења, при чему је нуклеарна лакуарност посебно осетљив параметар у откривању ране апоптозе. Фрактална анализа била у стању да открије ћелијске апоптотске промене пре конвенционалне проточне цитометријске анализе екстернализације фосфатидилсерина, фрагментације ДНК и пермеабилитетске мембране. Ови резултати указују да би фрактална анализа могла бити моћна и приступачна метода за неинвазивну рану идентификацију апоптозе у ћелијским културама.

У наредном истраживању испитиване су хемолитичке особине наночестица фулерена (C60) припремљених изменом растварача тетрахидрофурана (pC60THF) или са механохемијски потпомогнутим формирањем комплекса са макроцикличним олигосахаридом гама циклодекстрином (pC60CDKS) или кополимером етилен винил ацетат–етилен винил версататом (pC60EVA–EVV) (референца 7). Спектрофотометријска анализа ослобађања хемоглобина открила је да је само pC60THF, али не и pC60CDKS или pC60EVA–EVV, био у стању да изазове лизу еритроцита на начин који зависи од дозе и времена. Микроскопски је утврђено да хемолизи посредованој pC60THF претходи скупљање еритроцита и повећање храпавости површине ћелије. Проточна цитометријска анализа је потврдила смањење величине еритроцита и показала значајно смањење повећања производње реактивних врста кисеоника у црвеним крвним зрнцима изложеним pC60THF. Хемолитичку активност изазвану pC60THF ефикасно су смањили антиоксиданси Н-ацетилцистеин и бутиловани хидроксианизол, као и серумски албумин, најзаступљенији протеин у крвној плазми човека. Ови подаци указују да pC60THF може изазвати хемолизу која се може спречити у серумском албумину кроз оштећење мембране еритроцита посредовано оксидативним стресом.

Следећа публикација садржи истраживање ефеката лечења нарушене глукорегулације на нивое проинфламаторних Th17 цитокина, интерлеукина IL-17 и IL-23, Th1, интерферона (INF)- γ и IL-12 у серуму код новодијагностикованих пацијената са дијабетесом типа 2 (референца 9). Нивои цитокина у серуму, гликозилираног хемоглобина (HbA1C), индекс процене модела хомеостазе као мера инсулинске резистенције (HOMA-IR), и индекс телесне масе (BMI) су одређени пре и после 12 недеља терапије која се састојала од стандардних модификација начина живота и метформина (1000 mg два пута дневно). Нивои Th17 и Th1 цитокина пре третмана нису били у корелацији са узрастом, BMI или HOMA-IR. Пацијенти са лошом глукорегулацијом (HbA1C >7%), у поређењу са онима са добром глукорегулацијом (HbA1C $\leq$ 7%), имали су виши ниво Th17 и Th1 цитокина у серуму, али само разлике у IL-17 и INF- γ достигле су статистичку значајност ($p=0,003$ и $p=0,012$). Смањење вредности HbA1C (са 8,6 на 5,9%, $p=0,000$) примећено код пацијената са лошом регулацијом глукозе било је повезано

са значајним смањењем концентрације IL-17 (са 21,2 на 12,9 pg/ml, $p=0,020$), али не и INF- γ (50,6 наспрам 52,3, $p=0,349$). Ови подаци указују на то да терапијско побољшање глукорегулације може допринети смањењу нивоа IL-17 код новодијагностикованих пацијената са дијабетесом типа 2.

Клинички публицистички допринос кандидата је поткрепљен са три публикације у којима су представљени случајеви пацијената са лонгитуиналним екстензивним трансферзалним мијелитисом (референца 11), функционалном слабошћу мишића покретача очних јабучица (референца 12) и публикација у којој је претстављен случај пацијента млађе животне доби са можданим ударом у сливу Перчеронове артерија током оболевања од COVID-19 (референца 10).

4. НАУЧНО-ИСТРАЖИВАЧКИ РАД

4.1. Учесће у реализацији научних пројеката и ангажовање у руковођењу научним радом

Др сц. мед. Александар Пантовић је учествовао у реализацији 2 научноистраживачка пројекта:

1. Имуномодулаторно, цитопротективно и цитотоксично дејство наночестица (број пројекта 145073)
2. Модулација унутарћелијског енергетског баланса – контрола сигналних путева у терапији тумора и неуро-имуно-ендокриних поремећаја (број пројекта 41025)

5. КВАЛИТАТИВНИ ПОКАЗАТЕЉИ НАУЧНОГ УСПЕХА

5.1. Организација научног рада

До сада др Александар Пантовић није руководио пројектима нити пројектним задацима.

5.2. Ангажованост у образовању и формирању научних кадрова

Др сц. мед. Александар Пантовић води практичну наставу као сарадник у настави од марта месеца 2023. г.

5.3. Рецензија радова публикованих у научним часописима и предлога за пројекте

Др сц. мед. Алексанадр Пантовић није имао искуства као рецензент.

5.4. Међународна сарадња

- Др сц. мед. Александар Пантовић је више пута учествовао на домаћим и међународним конгресима као аутор радова
- IHead Academy: Precision medicine approach for migraine management
- COST Action Proposal OC-2023-1-26779, поднета апликација

5.5. Чланства и активност у научним друштвима

Др сц. мед. Александар Пантовић је члан следећих научних друштава:

- Društvo neurologa Srbije (DNS)
- Nacionalno udruženje za glavobolje Srbije (NUGS)
- International Society of Neuroimmunology (ISNI)
- Movement disorder society (MDS)
- European Federation of Neurological Societies (EFNS)
- Federation of European Neuroscience Societies (FENS)
- International Headache Society (IHS)

5.6. Оригиналност научног рада, степен самосталности у научноистраживачком раду и улога у реализацији радова

- Током научноистраживачког рада кандидат је исказао познавање научноистраживачке методологије. Учествовао је у свим фазама научноистраживачког процеса, од дизајнирања истраживања па све до публикаовања рада. У неколико радова заузима прво или друго место у списку коаутора, а такође у неколико радова је аутор за кореспонденцију.
- Од почетка научног и клиничког рада, др Пантовић је објавио 29 публицистичких јединица, од тога 10 радова штампаних у целини у међународним часописима ($7 \times M_{21} + 2 \times M_{22} + 1 \times M_{23}$). Збирни импакт фактор радова које је др Пантовић до сада публикаовао је **39,583** (као први аутор збирни импакт фактор је **12,841**). Просечан број коаутора у радовима др Пантовића публикаованим у целиности износи осам. На неколико радова број аутора је већи од осам, што је условљено мултидисциплинарним истраживачким приступом.
- Др Пантовић је од почетка научног и клиничког рада објавио 12 радова *in extenso*, при чему је на 6 био први аутор. На основу захтева Правилника о избору у звање збир од потребних 10 бодова у категоријама $M_{10}+M_{20}+M_{31}+M_{32}+M_{33}+M_{41}+M_{42}$ износи 72.5 (нормирано 54.04), односно, од потребних 6 бодова у категоријама $M_{11}+M_{12}+M_{21}+M_{22}+M_{23}$, збир износи 71 (нормирано 52.55).
- Пуубликације у којима је Др Пантовић први аутор или коаутор до сада су цитиране преко 900 пута (хетероцитати): Scopus 901; Web of Science 850

1. ПЕТ НАЈЗНАЧАЈНИХ НАУЧНИХ ОСТВАРЕЊА

Према мишљењу Комисије међу најважнијим научним остварењима др сц. мед. Александра Пантовића истичу се следећи радови:

1. **Pantovic A**, Bosnjak M, Arsikin K, Kostic M, Mandic M, Ristic B, Tosic J, Suzin-Zivkovic V, Grujicic D, Isakovic A, Trajkovic V, Harhaji-Trajkovic Lj. In vitro antiglioma action of indomethacin is mediated through AMPK/mTOR complex 1 signaling pathway. Int J Biochem Cell Biol. 2017;83:84-96. IF 3.905 (8 бодова) НББ: $8/(1+0,2*(12-7)) = 4$ Molecular biology & Neurosciences
2. **Pantovic A**, Krstic A, Janjetovic K, Kocic J, Harhaji-Trajkovic L, Bugarski D, Trajkovic V. Coordinated time-dependent modulation of AMPK/Akt/mTOR signaling and autophagy controls osteogenic differentiation of human mesenchymal stem cells. Bone. 2013;52(1):524-31. IF 4.461 (8 бодова) Metabolism & Endocrinology
3. Wasik AM, Grabarek J, **Pantovic A**, Ciešlar-Pobuda A, Asgari HR, Bundgaard-Nielsen C, Rafat M, Dixon IM, Ghavami S, Łos MJ. Reprogramming and carcinogenesis-parallels and

distinctions. *Int Rev Cell Mol Biol*. 2014;308:167-203. Review. IF 4.973 (8 бодова) НББ: $8/(1+0,2*(10-7)) = 5$ Biochemistry & Molecular medicine

4. Markovic Z, Harhaji-Trajkovic L, Todorovic-Markovic B, Kepić D, Arsikin K, Jovanović S, **Pantovic A**, Dramićanin M, Trajkovic V. In vitro comparison of the photothermal anticancer activity of graphene nanoparticles and carbon nanotubes. *Biomaterials*. 2011;32(4):1121-9. IF 7.404 (10 бодова) НББ: $10/(1+0,2*(9-7)) = 7.142$ Biomedical, Multidisciplinary
5. **Pantović A**, Lepic T, Pasovski V, Krsmanovic Z, Raicevic R. Artery of Percheron infarction associated with COVID-19 in the young adult. *J Neurovirol*. 2021 Dec;27(6):951-953. IF 3.739 (5 бодова) Neuroscience

6.1. Утицајност

- Др Пантовић је до сада цитиран 901 пута (хетероцитати): Scopus 901; Web of Science 850

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Markovic Z, Harhaji-Trajkovic L, Todorovic-Markovic B, Kepić D, Arsikin K, Jovanović S, **Pantovic A**, Dramićanin M, Trajkovic V. In vitro comparison of the photothermal anticancer activity of graphene nanoparticles and carbon nanotubes. *Biomaterials*. 2011;32(4):1121-9. IF 7.404 (10 бодова)

Хетероцитата 403:

Amaral, S.I., Costa-Almeida, R., Goncalves, I.C., Magalhaes, F.D., Pinto, A.M., 2022. Carbon nanomaterials for phototherapy of cancer and microbial infections. *CARBON* 190, 194–244. <https://doi.org/10.1016/j.carbon.2021.12.084>

Seifi, T., Kamali, A.R., 2022. The influence of mechanochemical treatment in hexane on dispersibility and floatability of graphite flakes with enhanced water evaporation performance. *COLLOIDS AND SURFACES A-PHYSICOCHEMICAL AND ENGINEERING ASPECTS* 638. <https://doi.org/10.1016/j.colsurfa.2022.128326>

Rodriguez-Montelongo, S.A., Gonzalez-Maldonado, M.A., Silva-Ramirez, A.S., del Campo, C., Ruiz, F., Gonzalez-Ortega, O., n.d. Graphene oxide and reduced graphene oxide nanocomposites grafted with hollow gold nanoshells as photothermal agents. *JOURNAL OF COMPOSITE MATERIALS*. <https://doi.org/10.1177/00219983221073982>

Gosecka, M., Gosecki, M., Urbaniak, M., n.d. Composite Dynamic Hydrogels Constructed on Boronic Ester Cross-Links with NIR-Enhanced Diffusivity. *BIOMACROMOLECULES*. <https://doi.org/10.1021/acs.biomac.1c01359>

Gonzalez-Colsa, J., Serrera, G., Saiz, J.M., Ortiz, D., Gonzalez, F., Bresme, F., Moreno, F., Albella, P., 2022. Gold nanodoughnut as an outstanding nanoheater for photothermal applications. *OPTICS EXPRESS* 30, 125–137. <https://doi.org/10.1364/OE.446637>

Lin, L.L., Li, H.Y., Su, S.S., Wen, X.Y., Yan, R., Tao, C.H., 2022. Study on the structure and properties of Fe₃O₄@HMPDA@HA magnetic hollow mesoporous submicron drug-carrying system. *MICROPOROUS AND MESOPOROUS MATERIALS* 330. <https://doi.org/10.1016/j.micromeso.2021.111582>

- Su, X.Y., Li, L.T., Cui, D.D., Fang, W., Shi, Y.J., 2022. Enhanced Photothermal and Photoacoustic Performance of Graphene Oxide in NIR-II Biowindow by Chemical Reduction. *PHOTONICS* 9. <https://doi.org/10.3390/photonics9010002>
- Kearns, O., Camisasca, A., Giordani, S., 2022. Hyaluronic Acid-Conjugated Carbon Nanomaterials for Enhanced Tumour Targeting Ability. *MOLECULES* 27. <https://doi.org/10.3390/molecules27010048>
- Zhang, W., Li, B., Duan, W.Y., Yao, X., Lu, X., Li, S.L., Tian, Y.P., Li, D.D., 2022. Confined in situ polymerization in a nanoscale porphyrinic metal-organic framework for fluorescence imaging-guided synergistic phototherapy. *INORGANIC CHEMISTRY FRONTIERS* 9, 670–677. <https://doi.org/10.1039/d1qi01384j>
- Kumar, N., Fazal, S., Miyako, E., Matsumura, K., Rajan, R., 2021. Avengers against cancer: A new era of nano-biomaterial-based therapeutics. *MATERIALS TODAY* 51, 317–349. <https://doi.org/10.1016/j.mattod.2021.09.020>
- Liang, P., Mao, L.S., Dong, Y.L., Zhao, Z.W., Sun, Q., Mazhar, M., Ma, Y.N., Yang, S.J., Ren, W., 2021. Design and Application of Near-Infrared Nanomaterial-Liposome Hybrid Nanocarriers for Cancer Photothermal Therapy. *PHARMACEUTICS* 13. <https://doi.org/10.3390/pharmaceutics13122070>
- Fatima, F., Singh, H.R., Jha, S.K., 2021. Assessment of antioxidant and cytotoxicity activities against A-549 lung cancer cell line by synthesized reduced graphene oxide nanoparticles mediated by Camellia sinensis. *BIOTECH* 11. <https://doi.org/10.1007/s13205-021-03015-z>
- Ma, H.S., Yu, Q.Q., Qu, Y., Zhu, Y.F., Wu, C.T., 2021. Manganese silicate nanospheres-incorporated hydrogels:starvation therapy and tissue regeneration. *BIOACTIVE MATERIALS* 6, 4558–4567. <https://doi.org/10.1016/j.bioactmat.2021.04.042>
- Milosavljevic, V., Mitrevska, K., Adam, V., 2022. Benefits of oxidation and size reduction of graphene/graphene oxide nanoparticles in biosensing application: Classification of graphene/ graphene oxide nanoparticles. *SENSORS AND ACTUATORS B-CHEMICAL* 353. <https://doi.org/10.1016/j.snb.2021.131122>
- Chattaraj, K.G., Paul, S., 2022. Appraising the potency of small molecule inhibitors and their graphene surface-mediated organizational attributes on uric acid-melamine clusters. *PHYSICAL CHEMISTRY CHEMICAL PHYSICS* 24, 1029–1047. <https://doi.org/10.1039/d1cp03695e>
- Wu, X.Y., Manickam, S., Wu, T., Pang, C.H., 2021. Insights into the Role of Graphene/Graphene-hybrid Nanocomposites in Antiviral Therapy. *CHEMBIOENG REVIEWS* 8, 549–563. <https://doi.org/10.1002/cben.202100018>
- Hadidi, N., Moghadam, N.S., Pazuki, G., Parvin, P., Shahi, F., 2021. In Vitro Evaluation of DSPE-PEG (5000) Amine SWCNT Toxicity and Efficacy as a Novel Nanovector Candidate in Photothermal Therapy by Response Surface Methodology (RSM). *CELLS* 10. <https://doi.org/10.3390/cells10112874>
- Wang, J.L., Sui, L.H., Huang, J., Miao, L., Nie, Y.B., Wang, K.S., Yang, Z.C., Huang, Q., Gong, X., Nan, Y.Y., Ai, K.L., 2021. MoS₂-based nanocomposites for cancer diagnosis and therapy. *BIOACTIVE MATERIALS* 6, 4209–4242. <https://doi.org/10.1016/j.bioactmat.2021.04.021>
- Wang, H., Zheng, Y., Sun, Q., Zhang, Z., Zhao, M.N., Peng, C., Shi, S.J., 2021. Ginsenosides emerging as both bifunctional drugs and nanocarriers for enhanced antitumor therapies. *JOURNAL OF NANOBIO TECHNOLOGY* 19. <https://doi.org/10.1186/s12951-021-01062-5>
- Chen, A.F., Wang, Q.K., Li, M.K., Peng, Z.Y., Lai, J.D., Zhang, J.J., Xu, J.B., Huang, H.X., Lei, C.H., 2021. Combined Approach of Compression Molding and Magnetic Attraction to Micropatterning of Magnetic Polydimethylsiloxane Composite Surfaces with Excellent Anti-Icing/Deicing Performance. *ACS APPLIED MATERIALS & INTERFACES* 13, 48153–48162. <https://doi.org/10.1021/acsami.1c15428>

- Parthasarathy, R., Kumar, S.P., Rao, H.C.Y., Chelliah, J., 2021. Synthesis of beta-Glucan Nanoparticles from Red Algae-Derived beta-Glucan for Potential Biomedical Applications. *APPLIED BIOCHEMISTRY AND BIOTECHNOLOGY* 193, 3983–3995. <https://doi.org/10.1007/s12010-021-03674-x>
- Jain, N.K., Chathoth, B.M., Bhaskar, V.S., Meena, H., Prasad, R., Srivastava, R., 2021. Nanoengineered photoactive theranostic agents for cancer. *NANOPHOTONICS* 10, 2973–2997. <https://doi.org/10.1515/nanoph-2021-0205>
- Mohan, H., Yoo, S., Thimmarayan, S., Oh, H.S., Kim, G., Seralathan, K.K., Shin, T., 2021. Nickel decorated manganese oxynitride over graphene nanosheets as highly efficient visible light driven photocatalysts for acetylsalicylic acid degradation. *ENVIRONMENTAL POLLUTION* 289. <https://doi.org/10.1016/j.envpol.2021.117864>
- Granja, A., Pinheiro, M., Sousa, C.T., Reis, S., 2021. Gold nanostructures as mediators of hyperthermia therapies in breast cancer. *BIOCHEMICAL PHARMACOLOGY* 190. <https://doi.org/10.1016/j.bcp.2021.114639>
- Hu, P.S., Chou, H.J., Chen, C.A., Wu, P.Y., Hsiao, K.H., Kuo, Y.M., 2021. Devising Hyperthermia Dose of NIR-Irradiated Cs_{0.33}WO₃ Nanoparticles for HepG2 Hepatic Cancer Cells. *NANOSCALE RESEARCH LETTERS* 16. <https://doi.org/10.1186/s11671-021-03565-4>
- Gao, F.C., Sun, Z.W., Zhao, L., Chen, F., Stenzel, M., Wang, F.L., Li, H., Zhang, L., Jiang, Y.Y., 2021. Bioactive engineered photothermal nanomaterials: from theoretical understanding to cutting-edge application strategies in anti-cancer therapy. *MATERIALS CHEMISTRY FRONTIERS* 5, 5257–5297. <https://doi.org/10.1039/d1qm00402f>
- Zhang, W.J., Shan, S.Y., Fan, J.Y., Yuan, F., Lawson, T., Kong, L.D., Hu, R.D., Liu, Y., 2021. A novel Vancomycin-Functionalized-Magnetic Graphene Composite for Use as a Near-Infrared-Induced Synergistic Chemo-Photothermal Antibacterial. *MACROMOLECULAR BIOSCIENCE* 21. <https://doi.org/10.1002/mabi.202100082>
- Prekodravac, J.R., Kepic, D.P., Colmenares, J.C., Giannakoudakis, D.A., Jovanovic, S.P., 2021. A comprehensive review on selected graphene synthesis methods: from electrochemical exfoliation through rapid thermal annealing towards biomass pyrolysis. *JOURNAL OF MATERIALS CHEMISTRY C* 9, 6722–6748. <https://doi.org/10.1039/d1tc01316e>
- Picchio, M.L., Bergueiro, J., Wedepohl, S., Minari, R.J., Igarzabal, C.I.A., Gugliotta, L.M., Cuggino, J.C., Calderon, M., 2021. Exploiting cyanine dye J-aggregates/monomer equilibrium in hydrophobic protein pockets for efficient multi-step phototherapy: an innovative concept for smart nanotheranostics. *NANOSCALE* 13, 8909–8921. <https://doi.org/10.1039/d0nr09058a>
- Chen, K., Fang, W.J., Zhang, Q.Y., Jiang, X.Y., Chen, Y., Xu, W.J., Shen, Q.M., Sun, P.F., Huang, W., 2021. Tunable NIR Absorption Property of a Dithiolenic Nickel Complex: A Promising NIR-II Absorption Material for Photothermal Therapy. *ACS APPLIED BIO MATERIALS* 4, 4406–4412. <https://doi.org/10.1021/acsabm.1c00168>
- Li, K., Lu, M., Xia, X.H., Huang, Y.Y., 2021. Recent advances in photothermal and RNA interfering synergistic therapy. *CHINESE CHEMICAL LETTERS* 32, 1010–1016. <https://doi.org/10.1016/j.cclet.2020.09.010>
- Al-Hakkani, M.F., Gouda, G.A., Hassan, S.H.A., Nagiub, A.M., 2021. Echinacea purpurea Mediated Hematite Nanoparticles (alpha-HNPs) Biofabrication, Characterization, Physicochemical Properties, and its In-vitro Biocompatibility Evaluation. *SURFACES AND INTERFACES* 24. <https://doi.org/10.1016/j.surfin.2021.101113>
- Ma, W.Q., Yang, H.L., Hu, Y.F., Chen, L., 2021. Fabrication of PEGylated porphyrin/reduced graphene oxide/doxorubicin nanoplatfrom for tumour combination therapy. *POLYMER INTERNATIONAL* 70, 1413–1420. <https://doi.org/10.1002/pi.6216>

- Gupta, N., Malviya, R., 2021. Understanding and advancement in gold nanoparticle targeted photothermal therapy of cancer. *BIOCHIMICA ET BIOPHYSICA ACTA-REVIEWS ON CANCER* 1875. <https://doi.org/10.1016/j.bbcan.2021.188532>
- Shariare, M.H., Masum, A.A., Alshehri, S., Alanazi, F.K., Uddin, J., Kazi, M., 2021. Preparation and Optimization of PEGylated Nano Graphene Oxide-Based Delivery System for Drugs with Different Molecular Structures Using Design of Experiment (DoE). *MOLECULES* 26. <https://doi.org/10.3390/molecules26051457>
- Smima, C.S., Lalitha, P., Sharma, S.C., Nagabhushana, H., 2021. Screening of anti-cancer activity of reduced graphene oxide biogenically synthesized against human breast cancer MCF-7 cell lines. *APPLIED NANOSCIENCE* 11, 1093–1105. <https://doi.org/10.1007/s13204-021-01696-9>
- Gao, R.F., van der Mei, H.C., Ren, Y.J., Chen, H., Chen, G.J., Busscher, H.J., Peterson, B.W., 2021. Thermo-resistance of ESKAPE-panel pathogens, eradication and growth prevention of an infectious biofilm by photothermal, polydopamine-nanoparticles in vitro. *NANOMEDICINE-NANOTECHNOLOGY BIOLOGY AND MEDICINE* 32. <https://doi.org/10.1016/j.nano.2020.102324>
- Ramal-Sanchez, M., Fontana, A., Valbonetti, L., Ordinelli, A., Bernabo, N., Barboni, B., 2021. Graphene and Reproduction: A Love-Hate Relationship. *NANOMATERIALS* 11. <https://doi.org/10.3390/nano11020547>
- Bastiancich, C., Da Silva, A., Esteve, M.A., 2021. Photothermal Therapy for the Treatment of Glioblastoma: Potential and Preclinical Challenges. *FRONTIERS IN ONCOLOGY* 10. <https://doi.org/10.3389/fonc.2020.610356>
- Gloriozov, I.P., Dem'yanov, P.I., Zhulyaev, N.S., Nechaev, M.S., Oprunenko, Y.F., Gam, F., Saillard, J.Y., Kuznetsov, A.E., 2021. DFT Investigation of the eta(6) reversible arrow eta(6)-Inter-ring Haptotropic Rearrangement of the Group 8 Metals Complexes [(graphene)MCp](+) (M = Fe, Ru, Os). *JOURNAL OF PHYSICAL CHEMISTRY A* 125, 366–375. <https://doi.org/10.1021/acs.jpca.0c08251>
- Nanda, S.S., Kaushal, S., Shin, Y., Yun, K., An, S.S.A., Hembram, K., Papaefthymiou, G.C., Yi, D.K., 2021. Cancer Cell Detection on the Surface of Top-Gated Monolayer Graphene via Raman Spectroscopy. *ACS APPLIED BIO MATERIALS* 4, 1493–1498. <https://doi.org/10.1021/acsabm.0c01377>
- Shende, P., Saini, P., 2021. Drug-free Nanotherapies for Cancer Treatment. *CURRENT NANOSCIENCE* 17, 463–474. <https://doi.org/10.2174/1573413716999201111202009>
- Yan, J.X., Wang, C.L., Jiang, X.M., Wei, Y.Q., Wang, Q., Cui, K.L., Xu, X., Wang, F., Zhang, L., 2021. Application of phototherapeutic-based nanoparticles in colorectal cancer. *INTERNATIONAL JOURNAL OF BIOLOGICAL SCIENCES* 17, 1361–1381. <https://doi.org/10.7150/ijbs.58773>
- Chen, X.J., Zou, J.F., Zhang, K., Zhu, J.J., Zhang, Y., Zhu, Z.H., Zheng, H.Y., Li, F.Z., Piao, J.G., 2021. Photothermal/matrix metalloproteinase-2 dual-responsive gelatin nanoparticles for breast cancer treatment. *ACTA PHARMACEUTICA SINICA B* 11, 271–282. <https://doi.org/10.1016/j.apsb.2020.08.009>
- Mohan, H., Ramalingam, V., Lim, J.M., Lee, S.W., Kim, J., Lee, J.H., Park, Y.J., Seralathan, K.K., Oh, B.T., 2020. E-waste based graphene oxide/V2O5/Pt ternary composite: Enhanced visible light driven photocatalyst for anti-microbial and anti-cancer activity. *COLLOIDS AND SURFACES A-PHYSICOCHEMICAL AND ENGINEERING ASPECTS* 607. <https://doi.org/10.1016/j.colsurfa.2020.125469>
- Mukherjee, S., Bytesnikova, Z., Ashrafi, A.M., Adam, V., Richtera, L., 2020. Graphene Oxide as a Nanocarrier for Biochemical Molecules: Current Understanding and Trends. *PROCESSES* 8. <https://doi.org/10.3390/pr8121636>
- Li, Y.L., Zheng, L., Xiao, L., Wang, L.D., Cui, J.Y., Sha, D.Y., Liu, C.S., 2020. Eco-friendly development of an ultrasmall IONP-loaded nanoplatfor for bimodal imaging-guided cancer theranostics. *BIOMATERIALS SCIENCE* 8, 6375–6386. <https://doi.org/10.1039/d0bm00867b>

- Chen, Y.L., Zhang, M.L., Lin, Z.W., Shi, X.Y., 2020. Fast near-infrared light responsive shape memory composites: Polydopamine nanospheres hybrid polynorbornene. *POLYMER* 206. <https://doi.org/10.1016/j.polymer.2020.122898>
- Shang, T.Y., Yu, X.Y., Han, S.S., Yang, B., 2020. Nanomedicine-based tumor photothermal therapy synergized immunotherapy. *BIOMATERIALS SCIENCE* 8, 5241–5259. <https://doi.org/10.1039/d0bm01158d>
- Farokhnezhad, M., Esmailzadeh, M., Nourian, M., Jalaeikhoo, H., Rajaeinejad, M., Irvani, S., Majidzadeh-A, K., 2020. Silica-gold nanoshell@graphene: a novel class of plasmonic nanoagents for photothermal cancer therapy. *JOURNAL OF PHYSICS D-APPLIED PHYSICS* 53. <https://doi.org/10.1088/1361-6463/ab95bf>
- Zuchowska, A., Dabrowski, B., Jastrzebska, E., Mazurkiewicz-Pawlicka, M., Malolepszy, A., Stobinski, L., Trzaskowski, M., Brzozka, Z., 2020. Cytotoxic properties of graphene derivatives depending on origin and type of cell line. *JOURNAL OF MATERIALS RESEARCH* 35, 2385–2395. <https://doi.org/10.1557/jmr.2020.201>
- Karki, N., Tiwari, H., Tewari, C., Rana, A., Pandey, N., Basak, S., Sahoo, N.G., 2020. Functionalized graphene oxide as a vehicle for targeted drug delivery and bioimaging applications. *JOURNAL OF MATERIALS CHEMISTRY B* 8, 8116–8148. <https://doi.org/10.1039/d0tb01149e>
- Dai, X.F., Yu, L.H., Zhao, X.J., Ostrikov, K., 2020. Nanomaterials for oncotherapies targeting the hallmarks of cancer. *NANOTECHNOLOGY* 31. <https://doi.org/10.1088/1361-6528/ab99f1>
- Lu, P., Yazdi, A.Z., Han, X.X., Al Husaini, K., Haime, J., Waye, N., Chen, P., 2020. Mechanistic Insights into the Cytotoxicity of Graphene Oxide Derivatives in Mammalian Cells. *CHEMICAL RESEARCH IN TOXICOLOGY* 33, 2247–2260. <https://doi.org/10.1021/acs.chemrestox.9b00391>
- Liu, L., Zhang, M.M., Zhang, Q., Jiang, W., 2020. Graphene nanosheets damage the lysosomal and mitochondrial membranes and induce the apoptosis of RBL-2H3 cells. *SCIENCE OF THE TOTAL ENVIRONMENT* 734. <https://doi.org/10.1016/j.scitotenv.2020.139229>
- Bonilla-Represa, V., Abalos-Labruzzi, C., Herrera-Martinez, M., Guerrero-Perez, M.O., 2020. Nanomaterials in Dentistry: State of the Art and Future Challenges. *NANOMATERIALS* 10. <https://doi.org/10.3390/nano10091770>
- Mathew, T., Sree, R.A., Aishwarya, S., Kounaina, K., Patil, A.G., Satapathy, P., Hudeda, S.P., More, S.S., Muthucheliyan, K., Kumar, T.N., Raghu, A. V, Reddy, K.R., Zameer, F., 2020. Graphene-based functional nanomaterials for biomedical and bioanalysis applications. *FLATCHEM* 23. <https://doi.org/10.1016/j.flatc.2020.100184>
- Xie, Z.J., Lu, R.T., Zhu, Y., Peng, M.H., Fan, T.J., Ren, P.G., Wang, B., Kang, L., Liu, X.Y., Li, S., Cui, H.X., 2020. Liquid-phase exfoliation of black sesame to create a nanoplatfor for in vitro photoluminescence and photothermal therapy. *NANOMEDICINE* 15, 2041–2052. <https://doi.org/10.2217/nnm-2020-0151>
- Smita, S.S., Das, A., Barui, A., 2020. Surface Functionalization of Green-synthesized Reduced Graphene Oxide with PPIX Enhances Photosensitization of Cancer Cells. *PHOTOCHEMISTRY AND PHOTOBIOLOGY* 96, 1283–1293. <https://doi.org/10.1111/php.13316>
- Gurbuz, B., Ayan, S., Bozlar, M., Ustundag, C.B., 2020. Carbonaceous nanomaterials for phototherapy: a review. *EMERGENT MATERIALS* 3, 479–502. <https://doi.org/10.1007/s42247-020-00118-w>
- de Paula, R.F.O., Rosa, I.A., Gafanhao, I.F.M., Fachi, J.L., Melero, A.M.G., Roque, A.O., Boldrini, V.O., Ferreira, L.A.B., Irazusta, S.P., Ceragioli, H.J., de Oliveira, E.C., 2020. Reduced graphene oxide, but not carbon nanotubes, slows murine melanoma after thermal ablation using LED light in B16F10 lineage cells. *NANOMEDICINE-NANOTECHNOLOGY BIOLOGY AND MEDICINE* 28. <https://doi.org/10.1016/j.nano.2020.102231>

- Afshar, E.G., Zarrabi, A., Dehshahri, A., Ashrafizadeh, M., Dehghannoudeh, G., Behnam, B., Mandegary, A., Pardakhty, A., Mohammadinejad, R., Tavakol, S., 2020. Graphene as a promising multifunctional nanoplatform for glioblastoma theranostic applications. *FLATCHEM* 22. <https://doi.org/10.1016/j.flatc.2020.100173>
- Auge, A., Camerel, F., Benoist, A., Zhao, Y., 2020. Near-infrared light-responsive UCST-nanogels using an efficient nickel-bis(dithiolene) photothermal crosslinker. *POLYMER CHEMISTRY* 11, 3863–3875. <https://doi.org/10.1039/d0py00567c>
- El Magdoub, H.M., Kenawy, S.H., Khalil, A.M., 2020. Betanin: a promising molecule for biomedical applications. *BIOINTERFACE RESEARCH IN APPLIED CHEMISTRY* 10, 5392–5399. <https://doi.org/10.33263/BRIAC103.392399>
- Martelli, C., King, A., Simon, T., Giamas, G., 2020. Graphene-Induced Transdifferentiation of Cancer Stem Cells as a Therapeutic Strategy against Glioblastoma. *ACS BIOMATERIALS SCIENCE & ENGINEERING* 6, 3258–3269. <https://doi.org/10.1021/acsbiomaterials.0c00197>
- Rajashekara, S., Shrivastava, A., Sumhitha, S., Kumari, S., 2020. Biomedical Applications of Biogenic Zinc Oxide Nanoparticles Manufactured from Leaf Extracts of *Calotropis gigantea* (L.) Dryand. *BIONANOSCIENCE* 10, 654–671. <https://doi.org/10.1007/s12668-020-00746-w>
- Shen, X., Li, T.T., Xie, X.X., Feng, Y., Chen, Z.Y., Yang, H., Wu, C.H., Deng, S.Q., Liu, Y.Y., 2020. PLGA-Based Drug Delivery Systems for Remotely Triggered Cancer Therapeutic and Diagnostic Applications. *FRONTIERS IN BIOENGINEERING AND BIOTECHNOLOGY* 8. <https://doi.org/10.3389/fbioe.2020.00381>
- Shafique, S., Karimov, K.S., Abid, M., Ahmed, M.M., Akhmedov, K.M., Aziz-ur-Rehman, 2020. Carbon nanotubes, orange dye, and graphene powder based multifunctional temperature, pressure, and displacement sensors. *JOURNAL OF MATERIALS SCIENCE-MATERIALS IN ELECTRONICS* 31, 8893–8899. <https://doi.org/10.1007/s10854-020-03424-5>
- Waikar, M.R., Sonker, R.K., Gupta, S., Chakravarti, S.K., Sonkawade, R.G., 2020. Post-gamma -irradiation effects on structural, optical and morphological properties of chemical vapour deposited MWCNTs. *MATERIALS SCIENCE IN SEMICONDUCTOR PROCESSING* 110. <https://doi.org/10.1016/j.mssp.2020.104975>
- You, Y.W., Cao, F.F., Zhao, Y.J., Deng, Q.Q., Sang, Y.J., Li, Y., Dong, K., Ren, J.S., Qu, X.G., 2020. Near-Infrared Light Dual-Promoted Heterogeneous Copper Nanocatalyst for Highly Efficient Bioorthogonal Chemistry in Vivo. *ACS NANO* 14, 4178–4187. <https://doi.org/10.1021/acsnano.9b08949>
- Zhao, J., Li, Q., Miao, B.L., Pi, H.M., Yang, P., 2020. Controlling Long-Distance Photoactuation with Protein Additives. *SMALL* 16. <https://doi.org/10.1002/smll.202000043>
- Wang, Y.F., Zhang, H.T., Xie, J., Liu, Y.X., Wang, S.L., Zhao, Q.F., 2020. Three dimensional mesoporous carbon nanospheres as carriers for chemo-photothermal therapy compared with two dimensional graphene oxide nanosheets. *COLLOIDS AND SURFACES A-PHYSICOCHEMICAL AND ENGINEERING ASPECTS* 590. <https://doi.org/10.1016/j.colsurfa.2020.124498>
- Mousavi, S.M., Low, F.W., Hashemi, S.A., Samsudin, N.A., Shakeri, M., Yusoff, Y., Rahsepar, M., Lai, C.W., Babapoor, A., Soroshnia, S., Goh, S.M., Tiong, S.K., Amin, N., 2020. Development of hydrophobic reduced graphene oxide as a new efficient approach for photochemotherapy. *RSC ADVANCES* 10, 12851–12863. <https://doi.org/10.1039/d0ra00186d>
- Pahang, F., Parvin, P., Bavali, A., 2020. Fluorescence quenching effects of carbon nano-structures (Graphene Oxide and Nano Diamond) coupled with Methylene Blue. *SPECTROCHIMICA ACTA PART A-MOLECULAR AND BIOMOLECULAR SPECTROSCOPY* 229. <https://doi.org/10.1016/j.saa.2019.117888>

- Prabhakar, O., Arun, K., 2020. Recent progresses and challenges in graphene based nano materials for advanced therapeutical applications: a comprehensive review. *MATERIALS TODAY COMMUNICATIONS* 22. <https://doi.org/10.1016/j.mtcomm.2019.100823>
- Baldea, I., Olteanu, D., Filip, G.A., Pogacean, F., Coros, M., Suciu, M., Tripon, S.C., Cenariu, M., Magerusan, L., Stefan-van Staden, R.I., Pruneanu, S., 2020. Cytotoxicity mechanisms of nitrogen-doped graphene obtained by electrochemical exfoliation of graphite rods, on human endothelial and colon cancer cells. *CARBON* 158, 267–281. <https://doi.org/10.1016/j.carbon.2019.12.011>
- Ahamed, M., Akhtar, M.J., Khan, M.A.M., 2020. Investigation of Cytotoxicity, Apoptosis, and Oxidative Stress Response of Fe₃O₄-RGO Nanocomposites in Human Liver HepG2 cells. *MATERIALS* 13. <https://doi.org/10.3390/ma13030660>
- Guday, G., Nickl, P., Adeli, M., Haag, R., 2020. Reversible Photothermal Homogenization of Fluorous Biphasic Systems with Perfluoroalkylated Nanographene. *ACS APPLIED NANO MATERIALS* 3, 1139–1146. <https://doi.org/10.1021/acsanm.9b01611>
- Farani, M.R., Khadiv-Parsi, P., Riazi, G.H., Ardestani, M.S., Rad, H.S., 2020. PEGylation of graphene/iron oxide nanocomposite: assessment of release of doxorubicin, magnetically targeted drug delivery and photothermal therapy. *APPLIED NANOSCIENCE* 10, 1205–1217. <https://doi.org/10.1007/s13204-020-01255-8>
- Chani, M.T.S., Karimov, K.S., Asiri, A.M., 2020. Carbon Nanotubes and Graphene Powder Based Multifunctional Pressure, Displacement and Gradient of Temperature Sensors. *SEMICONDUCTORS* 54, 85–90. <https://doi.org/10.1134/S1063782620010170>
- Chani, M.T.S., Karimov, K.S., Asiri, A.M., 2019. Carbon Nanotubes and Graphene Powder based Multifunctional Pressure, Displacement and Gradient of Temperature Sensors. *SEMICONDUCTORS* 53, 1622–+. <https://doi.org/10.1134/S106378261916019X>
- Kumar, J., Karthik, P., Neppolian, B., Thiruvadigal, D.J., 2019. Functionalization of zigzag graphene nanoribbon with DNA nucleobases-A DFT study. *APPLIED SURFACE SCIENCE* 496. <https://doi.org/10.1016/j.apsusc.2019.143667>
- Farokhnezhad, M., Esmaeilzadeh, M., 2019. Optical and Photothermal Properties of Graphene Coated Au-Ag Hollow Nanoshells: A Modeling for Efficient Photothermal Therapy. *JOURNAL OF PHYSICAL CHEMISTRY C* 123, 28907–28918. <https://doi.org/10.1021/acs.jpcc.9b08648>
- Jiang, M.J., Zhang, Y., Wu, G., Chen, S.C., Chen, L., Wang, Y.Z., 2019. NIR light manipulated “paper art” for customizing devices with sophisticated structure from DA-epoxy/graphene composites. *COMPOSITES PART B-ENGINEERING* 177. <https://doi.org/10.1016/j.compositesb.2019.107369>
- Shen, S., Chen, Y., Qi, S.Y., Zhang, X., Qiao, C., Wu, L., 2019. Erythrocyte Membrane Coated Fe₃O₄ Nanoparticles for Near Infrared Light Responsive Drug Delivery. *CHEMISTRY LETTERS* 48, 1414–1416. <https://doi.org/10.1246/cl.190538>
- Wei, W.F., Zhang, X.Y., Zhang, S., Wei, G., Su, Z.Q., 2019. Biomedical and bioactive engineered nanomaterials for targeted tumor photothermal therapy: A review. *MATERIALS SCIENCE & ENGINEERING C-MATERIALS FOR BIOLOGICAL APPLICATIONS* 104. <https://doi.org/10.1016/j.msec.2019.109891>
- He, Z., Li, J.Y., Chen, S.H., Zhou, R.H., 2019. Surface Inhomogeneity of Graphene Oxide Influences Dissociation of A beta(16-21) Peptide Assembly. *JOURNAL OF PHYSICAL CHEMISTRY B* 123, 9098–9103. <https://doi.org/10.1021/acs.jpcc.9b07359>
- Ha, Y.M., Kim, Y.O., Kim, Y.N., Kim, J., Lee, J.S., Cho, J.W., Endo, M., Muramatsu, H., Kim, Y.A., Jung, Y.C., 2019. Rapidly self-heating shape memory polyurethane nanocomposite with boron-doped single-walled carbon nanotubes using near-infrared laser. *COMPOSITES PART B-ENGINEERING* 175. <https://doi.org/10.1016/j.compositesb.2019.107065>

- Farokhnezhad, M., Esmailzadeh, M., 2019. Graphene coated gold nanoparticles: an emerging class of nanoagents for photothermal therapy applications. *PHYSICAL CHEMISTRY CHEMICAL PHYSICS* 21, 18352–18362. <https://doi.org/10.1039/c9cp03126j>
- Alsaedi, I.I.J., Taqi, Z.J., Hussien, A.M.A., Sulaiman, G.M., Jabir, M.S., 2019. Graphene nanoparticles induces apoptosis in MCF-7 cells through mitochondrial damage and NF-KB pathway. *MATERIALS RESEARCH EXPRESS* 6. <https://doi.org/10.1088/2053-1591/ab33af>
- Yu, Y., Yan, L.H., Si, J.H., Xu, Y.M., Hou, X., 2019. Femtosecond laser assisted synthesis of gold nanorod and graphene hybrids and its photothermal property in the near-infrared region. *JOURNAL OF PHYSICS AND CHEMISTRY OF SOLIDS* 132, 116–120. <https://doi.org/10.1016/j.jpcs.2019.04.020>
- AL-Jawad, S.M.H., Taha, A.A., Redha, A.M., 2019. Studying the structural, morphological, and optical properties of CuS:Ni nanostructure prepared by a hydrothermal method for biological activity. *JOURNAL OF SOL-GEL SCIENCE AND TECHNOLOGY* 91, 310–323. <https://doi.org/10.1007/s10971-019-05023-1>
- Girma, W.M., Dehvari, K., Ling, Y.C., Chang, J.Y., 2019. Albumin-functionalized CuFeS₂/photosensitizer nanohybrid for single-laser-induced folate receptor-targeted photothermal and photodynamic therapy. *MATERIALS SCIENCE & ENGINEERING C-MATERIALS FOR BIOLOGICAL APPLICATIONS* 101, 179–189. <https://doi.org/10.1016/j.msec.2019.03.074>
- Kitko, K.E., Zhang, Q., 2019. Graphene-Based Nanomaterials: From Production to Integration With Modern Tools in Neuroscience. *FRONTIERS IN SYSTEMS NEUROSCIENCE* 13. <https://doi.org/10.3389/fnsys.2019.00026>
- Gloriozov, I.P., Nechaev, M.S., Zaitsev, K. V., Oprunenko, Y.F., Gam, F., Saillard, J.Y., 2019. DFT study of inter- ring haptotropic rearrangement in CpRu⁺ complexes of polycyclic aromatic ligands. *JOURNAL OF ORGANOMETALLIC CHEMISTRY* 889, 9–14. <https://doi.org/10.1016/j.jorganchem.2019.03.016>
- Yao, J., Wang, H., Chen, M., Yang, M., 2019. Recent advances in graphene-based nanomaterials: properties, toxicity and applications in chemistry, biology and medicine. *MICROCHIMICA ACTA* 186. <https://doi.org/10.1007/s00604-019-3458-x>
- Qi, J., Zhang, W., Cao, R., 2019. A New Strategy for Solar-to-Hydrogen Energy Conversion: Photothermal-Promoted Electrocatalytic Water Splitting. *CHEMELECTROCHEM* 6, 2762–2765. <https://doi.org/10.1002/celec.201900530>
- Xu, C., Ma, B., Peng, J.L., Gao, L., Xu, Y.H., Huan, Z.G., Chang, J., 2019. Tricalcium silicate/graphene oxide bone cement with photothermal properties for tumor ablation. *JOURNAL OF MATERIALS CHEMISTRY B* 7, 2808–2818. <https://doi.org/10.1039/c9tb00246d>
- Gai, L.X., Wang, W.Q., Wu, X., Su, X.J., Yang, F.C., 2019. NIR absorbing reduced graphene oxide for photothermal radiotherapy for treatment of esophageal cancer. *JOURNAL OF PHOTOCHEMISTRY AND PHOTOBIOLOGY B-BIOLOGY* 194, 188–193. <https://doi.org/10.1016/j.jphotobiol.2019.03.014>
- Liu, Y.J., Bhattarai, P., Dai, Z.F., Chen, X.Y., 2019. Photothermal therapy and photoacoustic imaging via nanotheranostics in fighting cancer. *CHEMICAL SOCIETY REVIEWS* 48, 2053–2108. <https://doi.org/10.1039/c8cs00618k>
- Benos, L., Spyrou, L.A., Sarris, I.E., 2019. Development of a new theoretical model for blood-CNTs effective thermal conductivity pertaining to hyperthermia therapy of glioblastoma multiform. *COMPUTER METHODS AND PROGRAMS IN BIOMEDICINE* 172, 79–85. <https://doi.org/10.1016/j.cmpb.2019.02.008>
- Sattar, T., 2019. Current Review on Synthesis, Composites and Multifunctional Properties of Graphene. *TOPICS IN CURRENT CHEMISTRY* 377. <https://doi.org/10.1007/s41061-019-0235-6>

- Wang, J., Zhang, Y., Jin, N., Mao, C.B., Yang, M.Y., 2019. Protein-Induced Gold Nanoparticle Assembly for Improving the Photothermal Effect in Cancer Therapy. *ACS APPLIED MATERIALS & INTERFACES* 11, 11136–11143. <https://doi.org/10.1021/acsami.8b21488>
- Sun, W., Zhang, X.D., Jia, H.R., Zhu, Y.X., Guo, Y.X., Gao, G., Li, Y.H., Wu, F.G., 2019. Water-Dispersible Candle Soot-Derived Carbon Nano-Onion Clusters for Imaging-Guided Photothermal Cancer Therapy. *SMALL* 15. <https://doi.org/10.1002/sml.201804575>
- Jiang, B.P., Zhou, B., Lin, Z.X., Liang, H., Shen, X.C., 2019. Recent Advances in Carbon Nanomaterials for Cancer Phototherapy. *CHEMISTRY-A EUROPEAN JOURNAL* 25, 3993–4004. <https://doi.org/10.1002/chem.201804383>
- Shi, X.Q., Wang, Y., Sun, H.Y., Chen, Y.J., Zhang, X.Z., Xu, J.K., Zhai, G.X., 2019. Heparin-reduced graphene oxide nanocomposites for curcumin delivery: in vitro, in vivo and molecular dynamics simulation study. *BIOMATERIALS SCIENCE* 7, 1011–1027. <https://doi.org/10.1039/c8bm00907d>
- Xiang, H.J., Chen, Y., 2019. Energy-Converting Nanomedicine. *SMALL* 15. <https://doi.org/10.1002/sml.201805339>
- Li, C.L., Li, L.F., Li, J.C., Wu, X., Qi, L., Li, W.B., 2019. Fabrication and characterisation of viscose fibre with photoinduced heat-generating properties. *CELLULOSE* 26, 1631–1640. <https://doi.org/10.1007/s10570-018-2207-3>
- Bian, K.X., Zhang, X.W., Yang, M.X., Luo, L.Y., Li, L., He, Y.C., Cong, C., Li, X.L., Zhu, R.Y., Gao, D.W., 2019. Dual-template cascade synthesis of highly multi-branched Au nanoshells with ultrastrong NIR absorption and efficient photothermal therapeutic intervention. *JOURNAL OF MATERIALS CHEMISTRY B* 7, 598–610. <https://doi.org/10.1039/c8tb02753f>
- Maziukiewicz, D., Grzeskowiak, B.F., Coy, E., Jurga, S., Mrowczynski, R., 2019. NDs@PDA@ICG Conjugates for Photothermal Therapy of Glioblastoma Multiforme. *BIOMIMETICS* 4. <https://doi.org/10.3390/biomimetics4010003>
- Mousavi, S.M., Hashemi, S.A., Ghasemi, Y., Amani, A.M., Babapoor, A., Arjmand, O., 2019. Applications of graphene oxide in case of nanomedicines and nanocarriers for biomolecules: review study. *DRUG METABOLISM REVIEWS* 51, 12–41. <https://doi.org/10.1080/03602532.2018.1522328>
- Mousavi, S.M., Soroshnia, S., Hashemi, S.A., Babapoor, A., Ghasemi, Y., Savardashtaki, A., Amani, A.M., 2019. Graphene nano-ribbon based high potential and efficiency for DNA, cancer therapy and drug delivery applications. *DRUG METABOLISM REVIEWS* 51, 91–104. <https://doi.org/10.1080/03602532.2019.1582661>
- Gooneh-Farahani, S., Naimi-Jamal, M.R., Naghib, S.M., 2019. Stimuli-responsive graphene-incorporated multifunctional chitosan for drug delivery applications: a review. *EXPERT OPINION ON DRUG DELIVERY* 16, 79–99. <https://doi.org/10.1080/17425247.2019.1556257>
- Gupta, N., Rai, D.B., Jangid, A.K., Kulhari, H., 2019. A Review of Theranostics Applications and Toxicities of Carbon Nanomaterials. *CURRENT DRUG METABOLISM* 20, 506–532. <https://doi.org/10.2174/1389200219666180925094515>
- Hlapisi, N., Motaung, T.E., Lingano, L.Z., Oluwafemi, O.S., Songea, S.P., 2019. Encapsulation of Gold Nanorods with Porphyrins for the Potential Treatment of Cancer and Bacterial Diseases: A Critical Review. *BIOMATERIALS SCIENCE* 7, 1011–1027. <https://doi.org/10.1039/c8bm00907d>
- Gallagher, K.A., Doughty, A., Layton, E., Zukerman, S., Zhou, B.Q., Chen, W.R., 2019. The characterization of graphene-reconstituted low-density lipoproteins and small molecule inhibitor nanoparticles in preparation for use in laser immunotherapy. *BIOPHOTONICS AND IMMUNE RESPONSES XIV*. <https://doi.org/10.1117/12.2506714>

- Farani, M.R., Parsi, P.K., Riazi, G., Ardestani, M.S., Rad, H.S., 2019. Extending the application of a magnetic PEG three-part drug release device on a graphene substrate for the removal of Gram-positive and Gram-negative bacteria and cancerous and pathologic cells. *DRUG DESIGN DEVELOPMENT AND THERAPY* 13, 1581–1591. <https://doi.org/10.2147/DDDT.S181090>
- Goushbolagh, N.A., Keshavarz, M., Zare, M.H., Bahreyni-Toosi, M.H., Kargar, M., Farhood, B., 2019. Photosensitizer effects of MWCNTs-COOH particles on CT26 fibroblastic cells exposed to laser irradiation. *ARTIFICIAL CELLS NANOMEDICINE AND BIOTECHNOLOGY* 47, 1326–1334. <https://doi.org/10.1080/21691401.2019.1593997>
- Suhrland, C., Truman, J.P., Obeid, L.M., Sitharaman, B., 2019. Oxidized graphene nanoparticles as a delivery system for the pro-apoptotic sphingolipid C-6 ceramide. *JOURNAL OF BIOMEDICAL MATERIALS RESEARCH PART A* 107, 25–37. <https://doi.org/10.1002/jbm.a.36474>
- Chen, J.Q., Ning, C.Y., Zhou, Z.N., Yu, P., Zhu, Y., Tan, G.X., Mao, C.B., 2019. Nanomaterials as photothermal therapeutic agents. *PROGRESS IN MATERIALS SCIENCE* 99, 1–26. <https://doi.org/10.1016/j.pmatsci.2018.07.005>
- Li, L.P., 2018. Cisplatin-Loaded Polymeric Micelles with Aggregation-Induced Emission Feature for Cellular Imaging and Chemotherapy. *CHEMISTRYSELECT* 3, 13682–13691. <https://doi.org/10.1002/slct.201802542>
- Szczepaniak, J., Strojny, B., Sawosz Chwalibog, E., Jaworski, S., Jagiello, J., Winkowska, M., Szmidt, M., Wierzbicki, M., Sosnowska, M., Balaban, J., Winnicka, A., Lipinska, L., Witkowska Pilaszewicz, O., Grodzik, M., 2018. Effects of Reduced Graphene Oxides on Apoptosis and Cell Cycle of Glioblastoma Multiforme. *INTERNATIONAL JOURNAL OF MOLECULAR SCIENCES* 19. <https://doi.org/10.3390/ijms19123939>
- Pierini, F., Nakielski, P., Urbanek, O., Pawłowska, S., Lanzi, M., De Sio, L., Kowalewski, T.A., 2018. Polymer-Based Nanomaterials for Photothermal Therapy: From Light-Responsive to Multifunctional Nanoplatfoms for Synergistically Combined Technologies. *BIOMACROMOLECULES* 19, 4147–4167. <https://doi.org/10.1021/acs.biomac.8b01138>
- Fernandes, A.L., Nascimento, J.P., Santos, A.P., Furtado, C.A., Romano, L.A., da Rosa, C.E., Monserrat, J.M., Ventura-Lima, J., 2018. Assessment of the effects of graphene exposure in Danio rerio: A molecular, biochemical and histological approach to investigating mechanisms of toxicity. *CHEMOSPHERE* 210, 458–466. <https://doi.org/10.1016/j.chemosphere.2018.06.183>
- Jiang, G., Chen, L., Zhang, S.D., Huang, H.X., 2018. Superhydrophobic SiC/CNTs Coatings with Photothermal Deicing and Passive Anti-Icing Properties. *ACS APPLIED MATERIALS & INTERFACES* 10, 36505–36511. <https://doi.org/10.1021/acsami.8b11201>
- Liao, Y.Y., Wang, W.Y., Huang, X.M., Sun, Y.Y., Tian, S., Cai, P., 2018. Reduced graphene oxide triggered epithelial-mesenchymal transition in A549 cells. *SCIENTIFIC REPORTS* 8. <https://doi.org/10.1038/s41598-018-33414-x>
- Zhang, D.Y., Zheng, Y., Zhang, H., Sun, J.H., Tan, C.P., He, L., Zhang, W., Ji, L.N., Mao, Z.W., 2018. Delivery of Phosphorescent Anticancer Iridium(III) Complexes by Polydopamine Nanoparticles for Targeted Combined Photothermal-Chemotherapy and Thermal/Photoacoustic/Lifetime Imaging. *ADVANCED SCIENCE* 5. <https://doi.org/10.1002/advs.201800581>
- He, S., Li, H., Chen, H.L., 2018. Preparation of light-sensitive polymer/graphene composite via molecular recognition by beta-cyclodextrin. *JOURNAL OF MATERIALS SCIENCE* 53, 14337–14349. <https://doi.org/10.1007/s10853-018-2639-z>
- Cicuendez, M., Flores, J., Oliveira, H., Portoles, M.T., Vallet-Regi, M., Vila, M., Duarte, I.F., 2018. Metabolomic response of osteosarcoma cells to nanographene oxide-mediated hyperthermia. *MATERIALS SCIENCE & ENGINEERING C-MATERIALS FOR BIOLOGICAL APPLICATIONS* 91, 340–348. <https://doi.org/10.1016/j.msec.2018.05.057>

- Javadian, S., Khosravian, M., 2018. Revealing Factors Governing Self-Assembly Morphology of Fatty Acid on Graphene Synthesized by Surfactant-Assisted LPE: A Joint MD, SAPT(DFT), and Experimental Study. *JOURNAL OF PHYSICAL CHEMISTRY C* 122, 21387–21400. <https://doi.org/10.1021/acs.jpcc.8b05057>
- Moorthy, M.S., Hoang, G., Subramanian, B., Bui, N.Q., Panchanathan, M., Mondal, S., Tuong, V.P.T., Kim, H., Oh, J., 2018. Prussian blue decorated mesoporous silica hybrid nanocarriers for photoacoustic imaging-guided synergistic chemo-photothermal combination therapy. *JOURNAL OF MATERIALS CHEMISTRY B* 6, 5220–5233. <https://doi.org/10.1039/c8tb01214h>
- Tu, Z.X., Guday, G., Adeli, M., Haag, R., 2018. Multivalent Interactions between 2D Nanomaterials and Biointerfaces. *ADVANCED MATERIALS* 30. <https://doi.org/10.1002/adma.201706709>
- Siregar, S., Oktamuliani, S., Saijo, Y., 2018. A Theoretical Model of Laser Heating Carbon Nanotubes. *NANOMATERIALS* 8. <https://doi.org/10.3390/nano8080580>
- Wu, Q.W., Wei, G., Xu, Z.B., Han, J., Xi, J.Q., Fan, L., Gao, L.Z., 2018. Mechanistic Insight into the Light-Irradiated Carbon Capsules as an Antibacterial Agent. *ACS APPLIED MATERIALS & INTERFACES* 10, 25026–25036. <https://doi.org/10.1021/acsami.8b04932>
- Santos, C.I.M., Goncalves, G., Cicuendez, M., Mariz, I., Silva, V.S., Oliveira, H., Campos, F., Vieira, S.I., Marques, P., Macoas, E.M.S., Neves, M., Martinho, J.M.G., 2018. Biocompatible hybrids based on nanographene oxide covalently linked to glycolporphyrins: Synthesis, characterization and biological evaluation. *CARBON* 135, 202–214. <https://doi.org/10.1016/j.carbon.2018.04.040>
- Jovanovic, S., Holclajtner-Antunovic, I., Uskokovic-Markovic, S., Bajuk-Bogdanovic, D., Pavlovic, V., Tomic, D., Milenkovic, M., Markovic, B.T., 2018. Modification of graphene oxide surfaces with 12-molybdophosphoric acid: Structural and antibacterial study. *MATERIALS CHEMISTRY AND PHYSICS* 213, 157–167. <https://doi.org/10.1016/j.matchemphys.2018.04.011>
- Cai, Y., Si, W.L., Huang, W., Chen, P., Shao, J.J., Dong, X.C., 2018. Organic Dye Based Nanoparticles for Cancer Phototheranostics. *SMALL* 14. <https://doi.org/10.1002/smll.201704247>
- Cendrowski, K., Jedrzejczak-Silicka, M., 2018. Carbon nanotubes with controlled length - preparation, characterization and their cytocompatibility effects. *POLISH JOURNAL OF CHEMICAL TECHNOLOGY* 20, 71–79. <https://doi.org/10.2478/pjct-2018-0025>
- Priyadarsini, S., Mohanty, S., Mukherjee, S., Basu, S., Mishra, M., 2018. Graphene and graphene oxide as nanomaterials for medicine and biology application. *JOURNAL OF NANOSTRUCTURE IN CHEMISTRY* 8, 123–137. <https://doi.org/10.1007/s40097-018-0265-6>
- Polo, E., Poupard, M.F.N., Guerrini, L., Taboada, P., Pelaz, B., Alvarez-Puebla, R.A., del Pino, P., 2018. Colloidal bioplasmonics. *NANO TODAY* 20, 58–73. <https://doi.org/10.1016/j.nantod.2018.04.003>
- Li, R.Y., Shi, Y., Shi, L., Alsaedi, M., Wang, P., 2018. Harvesting Water from Air: Using Anhydrous Salt with Sunlight. *ENVIRONMENTAL SCIENCE & TECHNOLOGY* 52, 5398–5406. <https://doi.org/10.1021/acs.est.7b06373>
- Bramini, M., Alberini, G., Colombo, E., Chiacchiaretta, M., DiFrancesco, M.L., Maya-Vetencourt, J.F., Maragliano, L., Benfenati, F., Cesca, F., 2018. Interfacing Graphene-Based Materials With Neural Cells. *FRONTIERS IN SYSTEMS NEUROSCIENCE* 12. <https://doi.org/10.3389/fnysys.2018.00012>
- Mondal, H.S., Hossain, M.M., Rahaman, M.E., Amin, S.M.B., Hossain, M.B., Hasan Mahasin, M.M., Mondal, P.K., 2018. Optoelectronics Based Dynamic Advancement of Graphene: Characteristics and Applications. *CRYSTALS* 8. <https://doi.org/10.3390/cryst8040171>
- Zhu, D.M., Xie, W., Xiao, Y.S., Suo, M., Zan, M.H., Liao, Q.Q., Hu, X.J., Chen, L.B., Chen, B., Wu, W.T., Ji, L.W., Huang, H.M., Guo, S.S., Zhao, X.Z., Liu, Q.Y., Liu, W., 2018. Erythrocyte membrane-coated gold

- nanocages for targeted photothermal and chemical cancer therapy. *NANOTECHNOLOGY* 29. <https://doi.org/10.1088/1361-6528/aa9ca1>
- Gupta, J., Prakash, A., Jaiswal, M.K., Agarrwal, A., Bahadur, D., 2018. Superparamagnetic iron oxide-reduced graphene oxide nanohybrid-a vehicle for targeted drug delivery and hyperthermia treatment of cancer. *JOURNAL OF MAGNETISM AND MAGNETIC MATERIALS* 448, 332–338. <https://doi.org/10.1016/j.jmmm.2017.05.084>
- Pandey, R.K., Prajapati, V.K., 2018. Molecular and immunological toxic effects of nanoparticles. *INTERNATIONAL JOURNAL OF BIOLOGICAL MACROMOLECULES* 107, 1278–1293. <https://doi.org/10.1016/j.ijbiomac.2017.09.110>
- Du, Y.X., Li, D., Liu, L.B., Gai, G.J., 2018. Recent Achievements of Self-Healing Graphene/Polymer Composites. *POLYMERS* 10. <https://doi.org/10.3390/polym10020114>
- Deb, A., Vimala, R., 2018. Natural and synthetic polymer for graphene oxide mediated anticancer drug delivery- A comparative study. *INTERNATIONAL JOURNAL OF BIOLOGICAL MACROMOLECULES* 107, 2320–2333. <https://doi.org/10.1016/j.ijbiomac.2017.10.119>
- Peng, X., Wang, R., Wang, T.J., Yang, W.N., Wang, H., Gu, W., Ye, L., 2018. Carbon Dots/Prussian Blue Satellite/Core Nanocomposites for Optical Imaging and Photothermal Therapy. *ACS APPLIED MATERIALS & INTERFACES* 10, 1084–1092. <https://doi.org/10.1021/acsami.7b14972>
- Gazestani, A.M., Khoei, S., Khoei, S., Minaei, S.E., Motevalian, M., 2018. In vivo evaluation of the combination effect of near-infrared laser and 5-fluorouracil-loaded PLGA-coated magnetite nanographene oxide. *ARTIFICIAL CELLS NANOMEDICINE AND BIOTECHNOLOGY* 46, 25–33. <https://doi.org/10.1080/21691401.2018.1450265>
- Xu, Y.H., Shan, Y.L., Cong, H.L., Shen, Y.Q., Yu, B., 2018. Advanced Carbon-based Nanoplatfroms Combining Drug Delivery and Thermal Therapy for Cancer Treatment. *CURRENT PHARMACEUTICAL DESIGN* 24, 4060–4076. <https://doi.org/10.2174/1381612825666181120160959>
- Liu, Z., Zhang, J., Tian, Y.H., Zhang, L., Han, X., Wang, Q.C., Cheng, W., 2018. Targeted delivery of reduced graphene oxide nanosheets using multifunctional ultrasound nanobubbles for visualization and enhanced photothermal therapy. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 13, 7859–7872. <https://doi.org/10.2147/IJN.S181268>
- Lim, D.J., Park, H., 2018. Near-infrared light for on-demand drug delivery. *JOURNAL OF BIOMATERIALS SCIENCE-POLYMER EDITION* 29, 750–761. <https://doi.org/10.1080/09205063.2017.1398994>
- Madni, A., Noreen, S., Maqbool, I., Rehman, F., Batool, A., Kashif, P.M., Rehman, M., Tahir, N., Khan, M.I., 2018. Graphene-based nanocomposites: synthesis and their theranostic applications. *JOURNAL OF DRUG TARGETING* 26, 858–883. <https://doi.org/10.1080/1061186X.2018.1437920>
- Huang, Q.L., Li, M.Y., Wang, L.L., Yuan, H.H., Wang, M., Wu, Y.P., Li, T., 2018. Synthesis of novel cyclodextrin-modified reduced graphene oxide composites by a simple hydrothermal method. *RSC ADVANCES* 8, 37623–37630. <https://doi.org/10.1039/c8ra07807f>
- Chen, Y.C., Chiu, W.T., Chang, C., Wu, P.C., Tu, T.Y., Lin, H.P., Chang, H.C., 2018. Chemo-photothermal effects of doxorubicin/silica-carbon hollow spheres on liver cancer. *RSC ADVANCES* 8, 36775–36784. <https://doi.org/10.1039/c8ra08538b>
- Nassireslami, E., Ajdarzade, M., 2018. Gold Coated Superparamagnetic Iron Oxide Nanoparticles as Effective Nanoparticles to Eradicate Breast Cancer Cells via Photothermal Therapy. *ADVANCED PHARMACEUTICAL BULLETIN* 8, 201–209. <https://doi.org/10.15171/apb.2018.024>

- Tiwari, P., Agarwal, S., Srivastava, S., Jain, S., 2018. The combined effect of thermal and chemotherapy on HeLa cells using magnetically actuated smart textured fibrous system. *JOURNAL OF BIOMEDICAL MATERIALS RESEARCH PART B-APPLIED BIOMATERIALS* 106, 40–51. <https://doi.org/10.1002/jbm.b.33812>
- Kim, H., Nguyen, V.P., Manivasagan, P., Jung, M.J., Kim, S.W., Oh, J., Kang, H.W., 2017. Doxorubicin-fucoidan-gold nanoparticles composite for dual-chemo-photothermal treatment on eye tumors. *ONCOTARGET* 8, 113719–113733. <https://doi.org/10.18632/oncotarget.23092>
- Khan, M.S., Pandey, S., Bhaisare, M.L., Gedda, G., Talib, A., Wu, H.F., 2017. Graphene oxide@gold nanorods for chemo-photothermal treatment and controlled release of doxorubicin in mice Tumor. *COLLOIDS AND SURFACES B-BIOINTERFACES* 160, 543–552. <https://doi.org/10.1016/j.colsurfb.2017.09.001>
- Pang, Y.L., Mai, Z.H., Wang, B., Wang, L., Wu, L.P., Wang, X.P., Chen, T.S., 2017. Artesunate-modified nano-graphene oxide for chemo-photothermal cancer therapy. *ONCOTARGET* 8, 93800–93812. <https://doi.org/10.18632/oncotarget.21191>
- Song, Y., Shi, Q.R., Zhu, C.Z., Luo, Y.A., Lu, Q., Li, H., Ye, R.F., Du, D., Lin, Y.H., 2017. Mitochondrial-targeted multifunctional mesoporous Au@Pt nanoparticles for dual-mode photodynamic and photothermal therapy of cancers. *NANOSCALE* 9, 15813–15824. <https://doi.org/10.1039/c7nr04881e>
- Khan, M.S., Talib, A., Pandey, S., Bhaisare, M.L., Gedda, G., Wu, H.F., 2017. Folic Acid navigated Silver Selenide nanoparticles for photo-thermal ablation of cancer cells. *COLLOIDS AND SURFACES B-BIOINTERFACES* 159, 564–570. <https://doi.org/10.1016/j.colsurfb.2017.07.070>
- Shaheen, F., Aziz, M.H., Fakhar-e-Alam, M., Atif, M., Fatima, M., Ahmad, R., Hanif, A., Anwar, S., Zafar, F., Abbas, G., Ali, S.M., Ahmed, M., 2017. An In Vitro Study of the Photodynamic Effectiveness of GO-Ag Nanocomposites against Human Breast Cancer Cells. *NANOMATERIALS* 7. <https://doi.org/10.3390/nano7110401>
- Duverger, E., Picaud, F., Stauffer, L., Sonnet, P., 2017. Simulations of a Graphene Nanoflake as a Nanovector To Improve ZnPc Phototherapy Toxicity: From Vacuum to Cell Membrane. *ACS APPLIED MATERIALS & INTERFACES* 9, 37554–37562. <https://doi.org/10.1021/acsami.7b09054>
- Zhang, J., Li, J.C., Wang, X.H., Kawazoe, N., Chen, G.P., 2017. Targeting ligand-functionalized photothermal scaffolds for cancer cell capture and in situ ablation. *BIOMATERIALS SCIENCE* 5, 2276–2284. <https://doi.org/10.1039/c7bm00639j>
- Wang, K.W., Chen, Q.Q., Xue, W., Li, S., Liu, Z.H., 2017. Combined Chemo-photothermal Antitumor Therapy Using Molybdenum Disulfide Modified with Hyperbranched Polyglycidyl. *ACS BIOMATERIALS SCIENCE & ENGINEERING* 3, 2325–2335. <https://doi.org/10.1021/acsbiomaterials.7b00499>
- Silambarasan, D., Surya, V.J., Iyakutti, K., Asokan, K., Vasu, V., Kawazoe, Y., 2017. Gamma (gamma)-ray irradiated multi-walled carbon nanotubes (MWCNTs) for hydrogen storage. *APPLIED SURFACE SCIENCE* 418, 49–55. <https://doi.org/10.1016/j.apsusc.2017.02.262>
- Sun, L., Gong, P.W., Liu, X.C., Pang, M.H., Tian, M., Chen, J.Y., Du, J.Y., Liu, Z., 2017. Fluorinated carbon fiber as a novel nanocarrier for cancer chemo-photothermal therapy. *JOURNAL OF MATERIALS CHEMISTRY B* 5, 6128–6137. <https://doi.org/10.1039/c7tb01155e>
- Zhang, Y.M., Hong, H., Sun, B.Y., Carter, K., Qin, Y.R., Wei, W., Wang, D.P., Jeon, M., Geng, J.M., Nickles, R.J., Chen, G.Y., Prasad, P.N., Kim, C., Xia, J., Cai, W.B., Lovell, J.F., 2017. Surfactant-stripped naphthalocyanines for multimodal tumor theranostics with upconversion guidance cream. *NANOSCALE* 9, 3391–3398. <https://doi.org/10.1039/c6nr09321c>

- Zhu, R., Gao, F., Piao, J.G., Yang, L.H., 2017. Skin-safe photothermal therapy enabled by responsive release of acid-activated membrane-disruptive polymer from polydopamine nanoparticle upon very low laser irradiation. *BIOMATERIALS SCIENCE* 5, 1596–1602. <https://doi.org/10.1039/c7bm00256d>
- Li, A.H., Wang, Y., Chen, T., Zhao, W., Zhang, A.T., Feng, S.Y., Liu, J.Q., 2017. NIR-laser switched ICG/DOX loaded thermo-responsive polymeric capsule for chemo-photothermal targeted therapy. *EUROPEAN POLYMER JOURNAL* 92, 51–60. <https://doi.org/10.1016/j.eurpolymj.2017.04.019>
- Chowdhury, A., Tan, B., Venkatakrishnan, K., 2017. Fibroblast-Cytophilic and HeLa-Cytotoxic Dual Function Carbon Nanoribbon Network Platform. *ACS APPLIED MATERIALS & INTERFACES* 9, 19662–19676. <https://doi.org/10.1021/acsami.7b04819>
- Rubio-Pereda, P., Vilhena, J.G., Takeuchi, N., Serena, P.A., Perez, R., 2017. Albumin (BSA) adsorption onto graphite stepped surfaces. *JOURNAL OF CHEMICAL PHYSICS* 146. <https://doi.org/10.1063/1.4984037>
- Augustine, S., Singh, J., Srivastava, M., Sharma, M., Das, A., Malhotra, B.D., 2017. Recent advances in carbon based nanosystems for cancer theranostics. *BIOMATERIALS SCIENCE* 5, 901–952. <https://doi.org/10.1039/c7bm00008a>
- Zuchowska, A., Chudy, M., Dybko, A., Brzozka, Z., 2017. Graphene as a new material in anticancer therapy-in vitro studies. *SENSORS AND ACTUATORS B-CHEMICAL* 243, 152–165. <https://doi.org/10.1016/j.snb.2016.11.105>
- Chen, Y.J., Wu, Y.K., Sun, B.B., Liu, S.J., Liu, H.Y., 2017. Two-Dimensional Nanomaterials for Cancer Nanotheranostics. *SMALL* 13. <https://doi.org/10.1002/smll.201603446>
- Zhang, Y.X., Zhang, H., Wang, Y.K., Wu, H.X., Zeng, B., Zhang, Y.J., Tian, Q.W., Yang, S.P., 2017. Hydrophilic graphene oxide/bismuth selenide nanocomposites for CT imaging, photoacoustic imaging, and photothermal therapy. *JOURNAL OF MATERIALS CHEMISTRY B* 5, 1846–1855. <https://doi.org/10.1039/c6tb02137a>
- Ahmad, K., Wan, C., Al-Eshaikh, M.A., Kadachi, A.N., 2017. Reduction in thermal conductivity of BiSbTe lump. *APPLIED PHYSICS A-MATERIALS SCIENCE & PROCESSING* 123. <https://doi.org/10.1007/s00339-016-0738-8>
- de Melo-Diogo, D., Pais-Silva, C., Costa, E.C., Louro, R.O., Correia, I.J., 2017. D-alpha-tocopheryl polyethylene glycol 1000 succinate functionalized nanographene oxide for cancer therapy. *NANOMEDICINE* 12, 443–456. <https://doi.org/10.2217/nnm-2016-0384>
- Yoon, H.J., Lee, H.S., Lim, J.Y., Park, J.H., 2017. Liposomal Indocyanine Green for Enhanced Photothermal Therapy. *ACS APPLIED MATERIALS & INTERFACES* 9, 5683–5691. <https://doi.org/10.1021/acsami.6b16801>
- Vimala, K., Shanthi, K., Sundarraj, S., Kannan, S., 2017. Synergistic effect of chemo-photothermal for breast cancer therapy using folic acid (FA) modified zinc oxide nanosheet. *JOURNAL OF COLLOID AND INTERFACE SCIENCE* 488, 92–108. <https://doi.org/10.1016/j.jcis.2016.10.067>
- Ban, Q.F., Bai, T., Duan, X., Kong, J., 2017. Noninvasive photothermal cancer therapy nanoplatfroms via integrating nanomaterials and functional polymers. *BIOMATERIALS SCIENCE* 5, 190–210. <https://doi.org/10.1039/c6bm00600k>
- Jaleel, J.A., Sruthi, S., Pramod, K., 2017. Reinforcing nanomedicine using graphene family nanomaterials. *JOURNAL OF CONTROLLED RELEASE* 255, 218–230. <https://doi.org/10.1016/j.jconrel.2017.04.041>
- Teodorescu, F., Queniat, G., Foulon, C., Lecoœur, M., Barras, A., Boulahneche, S., Medjram, M.S., Hubert, T., Abderrahmani, A., Boukherroub, R., Szunerits, S., 2017. Transdermal skin patch based on reduced graphene

oxide: A new approach for photothermal triggered permeation of ondansetron across porcine skin. *JOURNAL OF CONTROLLED RELEASE* 245, 137–146. <https://doi.org/10.1016/j.jconrel.2016.11.029>

Parida, S., Maiti, C., Rajesh, Y., Dey, K.K., Pal, I., Parekh, A., Patra, R., Dibakar, D., Dutta, P.K., Mandal, M., 2017. Gold nanorod embedded reduction responsive block copolymer micelle-triggered drug delivery combined with photothermal ablation for targeted cancer therapy. *BIOCHIMICA ET BIOPHYSICA ACTA-GENERAL SUBJECTS* 1861, 3039–3052. <https://doi.org/10.1016/j.bbagen.2016.10.004>

Li, A., Li, X., Yu, X.J., Li, W., Zhao, R.Y., An, X., Cui, D.X., Chen, X.Y., Li, W.W., 2017. Synergistic thermoradiotherapy based on PEGylated Cu₃BiS₃ ternary semiconductor nanorods with strong absorption in the second near-infrared window. *BIOMATERIALS* 112, 164–175. <https://doi.org/10.1016/j.biomaterials.2016.10.024>

Zhang, B.M., Wang, Y., Liu, J.Y., Zhai, G.X., 2017. Recent Developments of Phototherapy Based on Graphene Family Nanomaterials. *CURRENT MEDICINAL CHEMISTRY* 24, 268–291. <https://doi.org/10.2174/0929867323666161019141817>

Sobhani, Z., Behnam, M.A., Emami, F., Dehghanian, A., Jamhiri, I., 2017. Photothermal therapy of melanoma tumor using multiwalled carbon nanotubes. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 12, 4509–4517. <https://doi.org/10.2147/IJN.S134661>

Rahmanian, N., Eskandani, M., Barar, J., Omid, Y., 2017. Recent trends in targeted therapy of cancer using graphene oxide-modified multifunctional nanomedicines. *JOURNAL OF DRUG TARGETING* 25, 202–215. <https://doi.org/10.1080/1061186X.2016.1238475>

Gong, C., Lu, C.C., Li, B.Q., Shan, M., Wu, G.L., 2017. Dopamine-modified poly(amino acid): an efficient near-infrared photothermal therapeutic agent for cancer therapy. *JOURNAL OF MATERIALS SCIENCE* 52, 955–967. <https://doi.org/10.1007/s10853-016-0391-9>

Chen, Y.L., Chiang, J.Y., Chou, H.T., Fu, C.Y., Chen, Y.C., Lee, C.Y., Tai, N.H., Chang, H.Y., 2017. Toxicity analysis of poly(sodium-4-styrenesulfonate) coated graphene on HMEC-1 cells under dynamic conditions mimicking blood flow. *RSC ADVANCES* 7, 51910–51918. <https://doi.org/10.1039/c7ra08286j>

Liu, C.C., Zhao, J.J., Zhang, R., Li, H., Chen, B., Zhang, L.L., Yang, H., 2017. Multifunctionalization of graphene and graphene oxide for controlled release and targeted delivery of anticancer drugs. *AMERICAN JOURNAL OF TRANSLATIONAL RESEARCH* 9, 5197–5219.

Ou, L.L., Lin, S.Q., Song, B., Liu, J., Lai, R.F., Shao, L.Q., 2017. The mechanisms of graphene-based materials-induced programmed cell death: a review of apoptosis, autophagy, and programmed necrosis. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 12, 6633–6646. <https://doi.org/10.2147/IJN.S140526>

Raghav, S., Painuli, R., Kumar, D., 2017. Multifunctional Nanomaterials for Multifaceted Applications in Biomedical Arena. *INTERNATIONAL JOURNAL OF PHARMACOLOGY* 13, 890–906. <https://doi.org/10.3923/ijp.2017.890.906>

Pattanaik, S., Swain, K., Lin, Z.Q., 2016. Graphene and graphene-based nanocomposites: biomedical applications and biosafety. *JOURNAL OF MATERIALS CHEMISTRY B* 4, 7813–7831. <https://doi.org/10.1039/c6tb02086k>

Cho, Y.C., Pak, P.J., Joo, Y.H., Lee, H.S., Chung, N., 2016. In vitro and in vivo comparison of the immunotoxicity of single- and multi-layered graphene oxides with or without pluronic F-127. *SCIENTIFIC REPORTS* 6. <https://doi.org/10.1038/srep38884>

Dhar, P., Paul, A., Narasimhan, A., Das, S.K., 2016. Analytical prediction of sub-surface thermal history in translucent tissue phantoms during plasmonic photo-thermotherapy (PPTT). *JOURNAL OF THERMAL BIOLOGY* 62, 143–149. <https://doi.org/10.1016/j.jtherbio.2016.06.023>

- Wu, Q., Chu, M.Q., Shao, Y.X., Wo, F.J., Shi, D.L., 2016. Reduced graphene oxide conjugated with CuInS₂/ZnS nanocrystals with low toxicity for enhanced photothermal and photodynamic cancer therapies. *CARBON* 108, 21–37. <https://doi.org/10.1016/j.carbon.2016.06.070>
- Chen, Y.W., Su, Y.L., Hu, S.H., Chen, S.Y., 2016. Functionalized graphene nanocomposites for enhancing photothermal therapy in tumor treatment. *ADVANCED DRUG DELIVERY REVIEWS* 105, 190–204. <https://doi.org/10.1016/j.addr.2016.05.022>
- Tian, M.W., Du, M.Z., Qu, L.J., Zhang, K., Li, H.L., Zhu, S.F., Liu, D.D., 2016. Conductive reduced graphene oxide/MnO₂ carbonized cotton fabrics with enhanced electro-chemical, -heating, and -mechanical properties. *JOURNAL OF POWER SOURCES* 326, 428–437. <https://doi.org/10.1016/j.jpowsour.2016.07.028>
- Zheng, M., Li, Y., Liu, S., Wang, W.Q., Xie, Z.G., Jing, X.B., 2016. One-Pot To Synthesize Multifunctional Carbon Dots for Near Infrared Fluorescence Imaging and Photothermal Cancer Therapy. *ACS APPLIED MATERIALS & INTERFACES* 8, 23533–23541. <https://doi.org/10.1021/acsami.6b07453>
- Ryu, T.K., Baek, S.W., Kang, R.H., Choi, S.W., 2016. Selective Photothermal Tumor Therapy Using Nanodiamond-Based Nanoclusters with Folic Acid. *ADVANCED FUNCTIONAL MATERIALS* 26, 6428–6436. <https://doi.org/10.1002/adfm.201601207>
- He, C.F., Wang, S.H., Yu, Y.J., Shen, H.Y., Zhao, Y., Gao, H.L., Wang, H., Li, L.L., Liu, H.Y., 2016. Advances in biodegradable nanomaterials for photothermal therapy of cancer. *CANCER BIOLOGY & MEDICINE* 13, 299–312. <https://doi.org/10.20892/j.issn.2095-3941.2016.0052>
- Kim, H., Kim, J., Lee, M., Choi, H.C., Kim, W.J., 2016. Stimuli-Regulated Enzymatically Degradable Smart Graphene-Oxide-Polymer Nanocarrier Facilitating Photothermal Gene Delivery. *ADVANCED HEALTHCARE MATERIALS* 5, 1918–1930. <https://doi.org/10.1002/adhm.201600246>
- Su, S.H., Wang, J.L., Vargas, E., Wei, J.H., Martinez-Zaguilan, R., Sennoune, S.R., Pantoya, M.L., Wang, S.R., Chaudhuri, J., Qiu, J.L., 2016. Porphyrin Immobilized Nanographene Oxide for Enhanced and Targeted Photothermal Therapy of Brain Cancer. *ACS BIOMATERIALS SCIENCE & ENGINEERING* 2, 1357–1366. <https://doi.org/10.1021/acsbiomaterials.6b00290>
- Dong, H.X., Jin, M., Liu, Z.M., Xiong, H.L., Qiu, X.J., Zhang, W., Guo, Z.Y., 2016. In vitro and in vivo brain-targeting chemo-photothermal therapy using graphene oxide conjugated with transferrin for Gliomas. *LASERS IN MEDICAL SCIENCE* 31, 1123–1131. <https://doi.org/10.1007/s10103-016-1955-2>
- Liu, T.M., Conde, J., Lipinski, T., Bednarkiewicz, A., Huang, C.C., 2016. Revisiting the classification of NIR-absorbing/emitting nanomaterials for in vivo bioapplications. *NPG ASIA MATERIALS* 8. <https://doi.org/10.1038/am.2016.106>
- Azhdarzadeh, M., Atyabi, F., Saei, A.A., Varnamkhashi, B.S., Omid, Y., Fateh, M., Ghavami, M., Shانهsazzadeh, S., Dinarvand, R., 2016. Theranostic MUC-1 aptamer targeted gold coated superparamagnetic iron oxide nanoparticles for magnetic resonance imaging and photothermal therapy of colon cancer. *COLLOIDS AND SURFACES B-BIOINTERFACES* 143, 224–232. <https://doi.org/10.1016/j.colsurfb.2016.02.058>
- Dudek, I., Skoda, M., Jarosz, A., Szukiewicz, D., 2016. The Molecular Influence of Graphene and Graphene Oxide on the Immune System Under In Vitro and In Vivo Conditions. *ARCHIVUM IMMUNOLOGIAE ET THERAPIAE EXPERIMENTALIS* 64, 195–215. <https://doi.org/10.1007/s00005-015-0369-3>
- Georgakilas, V., Tiwari, J.N., Kemp, K.C., Perman, J.A., Bourlinos, A.B., Kim, K.S., Zboril, R., 2016. Noncovalent Functionalization of Graphene and Graphene Oxide for Energy Materials, Biosensing, Catalytic, and Biomedical Applications. *CHEMICAL REVIEWS* 116, 5464–5519. <https://doi.org/10.1021/acs.chemrev.5b00620>

- Liu, H.Y., Li, T., Liu, Y.H., Qin, G.Q., Wang, X.P., Chen, T.S., 2016. Glucose-Reduced Graphene Oxide with Excellent Biocompatibility and Photothermal Efficiency as well as Drug Loading. *NANOSCALE RESEARCH LETTERS* 11. <https://doi.org/10.1186/s11671-016-1423-8>
- Mahmoudifard, M., Soleimani, M., Hatamie, S., Zamanlui, S., Ranjbarvan, P., Vossoughi, M., Hosseinzadeh, S., 2016. The different fate of satellite cells on conductive composite electrospun nanofibers with graphene and graphene oxide nanosheets. *BIOMEDICAL MATERIALS* 11. <https://doi.org/10.1088/1748-6041/11/2/025006>
- Cherukula, K., Lekshmi, K.M., Uthaman, S., Cho, K., Cho, C.S., Park, I.K., 2016. Multifunctional Inorganic Nanoparticles: Recent Progress in Thermal Therapy and Imaging. *NANOMATERIALS* 6. <https://doi.org/10.3390/nano6040076>
- Wang, L., Wen, W., Li, S., Fu, Z.Q., Tong, Y.J., Wang, J., Han, D., Zhang, A.P., 2016. Photocatalytic Killing Effect of Fe₃O₄-TiO₂ Nanoparticles on Hepatoma Carcinoma Cells for Targeting Photodynamic Therapy. *CHINESE JOURNAL OF INORGANIC CHEMISTRY* 32, 393–404.
- Zhou, F., Wang, H.J., Chang, J., 2016. Progress in the Field of Constructing Near-Infrared Light-Responsive Drug Delivery Platforms. *JOURNAL OF NANOSCIENCE AND NANOTECHNOLOGY* 16, 2111–2125. <https://doi.org/10.1166/jnn.2016.10945>
- Liu, H.Z., Niu, D., Jiang, W.T., Zhao, T.T., Lei, B.A., Yin, L., Shi, Y.S., Chen, B.D., Lu, B.H., 2016. Illumination-oriented and thickness-dependent photomechanical bilayer actuators realized by graphene-nanoplatelets. *SENSORS AND ACTUATORS A-PHYSICAL* 239, 45–53. <https://doi.org/10.1016/j.sna.2016.01.018>
- Ma, H.S., Jiang, C.A., Zhai, D., Luo, Y.X., Chen, Y., Lv, F., Yi, Z.F., Deng, Y., Wang, J.W., Chang, J., Wu, C.T., 2016. A Bifunctional Biomaterial with Photothermal Effect for Tumor Therapy and Bone Regeneration. *ADVANCED FUNCTIONAL MATERIALS* 26, 1197–1208. <https://doi.org/10.1002/adfm.201504142>
- Vilhena, J.G., Rubio-Pereda, P., Velloso, P., Serena, P.A., Perez, R., 2016. Albumin (BSA) Adsorption over Graphene in Aqueous Environment: Influence of Orientation, Adsorption Protocol, and Solvent Treatment. *LANGMUIR* 32, 1742–1755. <https://doi.org/10.1021/acs.langmuir.5b03170>
- Piao, J.G., Liu, D., Hu, K., Wang, L.M., Gao, F., Long, Y.J., Yang, L.H., 2016. Cooperative Nanoparticle System for Photothermal Tumor Treatment without Skin Damage. *ACS APPLIED MATERIALS & INTERFACES* 8, 2847–2856. <https://doi.org/10.1021/acsami.5b11664>
- Wu, C.H., Zhu, A.N., Li, D., Wang, L.H., Yang, H., Zeng, H.J., Liu, Y.Y., 2016. Photosensitizer-assembled PEGylated graphene-copper sulfide nanohybrids as a synergistic near-infrared phototherapeutic agent. *EXPERT OPINION ON DRUG DELIVERY* 13, 155–165. <https://doi.org/10.1517/17425247.2016.1118049>
- Paul, A., Narasimhan, A., Das, S.K., Sengupta, S., Pradeep, T., 2016. Subsurface thermal behaviour of tissue mimics embedded with large blood vessels during plasmonic photo-thermal therapy. *INTERNATIONAL JOURNAL OF HYPERTHERMIA* 32, 765–777. <https://doi.org/10.1080/02656736.2016.1196831>
- Yasoda, K.Y., Bobba, K.N., Nedungadi, D., Dutta, D., Kumar, M.S., Kothurkar, N., Mishra, N., Bhuniya, S., 2016. GSH-responsive biotinylated poly(vinyl alcohol)-grafted GO as a nanocarrier for targeted delivery of camptothecin. *RSC ADVANCES* 6, 62385–62389. <https://doi.org/10.1039/c6ra09706e>
- Cao, F., Yao, Q., Yang, T.S., Zhang, Z., Han, Y., Feng, J.C., Wang, X.H., 2016. Marriage of antibody-drug conjugate with gold nanorods to achieve multi-modal ablation of breast cancer cells and enhanced photoacoustic performance. *RSC ADVANCES* 6, 46594–46606. <https://doi.org/10.1039/c6ra01557c>
- Trusovas, R., Raciukaitis, G., Niaura, G., Barkauskas, J., Valusis, G., Pauliukaite, R., 2016. Recent Advances in Laser Utilization in the Chemical Modification of Graphene Oxide and Its Applications. *ADVANCED OPTICAL MATERIALS* 4, 37–65. <https://doi.org/10.1002/adom.201500469>

- Cheng, J.S., Wan, W.H., Zhu, W.J., 2016. One-Pot Solvothermal Synthesis of TiO₂ Nanobelt/Graphene Composites for Selective Renal Cancer Cells Destruction. CHINESE JOURNAL OF CHEMISTRY 34, 53–58. <https://doi.org/10.1002/cjoc.201500339>
- Jin, Z., Cho, S., Ko, S.Y., Park, J.O., Park, S., IEEE, 2016. Controlled Drug Releasing of Doxorubicin Loaded Magnetic Nanoliposome using NIR Irradiation. 2016 13TH INTERNATIONAL CONFERENCE ON UBIQUITOUS ROBOTS AND AMBIENT INTELLIGENCE (URAI).
- Sundar, D.S., Antoniraj, M.G., Kumar, C.S., Mohapatra, S.S., Houreld, N.N., Ruckmani, K., 2016. Recent Trends of Biocompatible and Biodegradable Nanoparticles in Drug Delivery: A Review. CURRENT MEDICINAL CHEMISTRY 23, 3730–3751. <https://doi.org/10.2174/0929867323666160607103854>
- Hosseini, L., Mahboobnia, K., Irani, M., 2016. Fabrication of PLA/MWCNT/Fe₃O₄ composite nanofibers for leukemia cancer cells. INTERNATIONAL JOURNAL OF POLYMERIC MATERIALS AND POLYMERIC BIOMATERIALS 65, 176–182. <https://doi.org/10.1080/00914037.2015.1074912>
- Kafa, H., Wang, J.T.W., Al-Jamal, K.T., 2016. Current Perspective of Carbon Nanotubes Application in Neurology, in: AlJamal, K.T. (Ed.), INTERNATIONAL REVIEW OF NEUROBIOLOGY, VOL 130: NANOTECHNOLOGY AND THE BRAIN. pp. 229–263. <https://doi.org/10.1016/bs.irn.2016.07.001>
- Marino, N., Petralia, S., Perez-Lloret, M., Mosinger, J., Conoci, S., Sortino, S., 2016. Graphene oxide nanohybrid that photoreleases nitric oxide. JOURNAL OF MATERIALS CHEMISTRY B 4, 5825–5830. <https://doi.org/10.1039/c6tb01599a>
- Mo, X.J., Wei, Y., Zhang, X.H., Cai, Q., Shen, Y., Dai, X.H., Meng, S., Liu, X., Liu, Y., Hu, Z.W., Deng, X.L., 2016. Enhanced Stem Cell Osteogenic Differentiation by Bioactive Glass Functionalized Graphene Oxide Substrates. JOURNAL OF NANOMATERIALS 2016. <https://doi.org/10.1155/2016/5613980>
- Park, J., Jeong, Y.G., 2015. Effects of Chain Orientation and Packing on the Photoluminescence and Photothermal Properties of Polybenzimidazole Fibers with Meta-Linkage. MACROMOLECULES 48, 8823–8830. <https://doi.org/10.1021/acs.macromol.5b02266>
- Feng, W., Chen, L., Qin, M., Zhou, X.J., Zhang, Q.Q., Miao, Y.K., Qiu, K.X., Zhang, Y.Z., He, C.L., 2015. Flower-like PEGylated MoS₂ nanoflakes for near-infrared photothermal cancer therapy. SCIENTIFIC REPORTS 5. <https://doi.org/10.1038/srep17422>
- Bijukumar, D., Girish, C.M., Sasidharan, A., Nair, S., Koyakutty, M., 2015. RETRACTED: Transferrin-Conjugated Biodegradable Graphene for Targeted Radiofrequency Ablation of Hepatocellular Carcinoma (Retracted article. See vol. 7, pg. 382, 2021). ACS BIOMATERIALS SCIENCE & ENGINEERING I, 1211–1219. <https://doi.org/10.1021/acsbiomaterials.5b00184>
- Akhtar, M.J., Alhadlaq, H.A., Kumar, S., Airokayan, S.A., Ahamed, M., 2015. Selective cancer-killing ability of metal-based nanoparticles: implications for cancer therapy. ARCHIVES OF TOXICOLOGY 89, 1895–1907. <https://doi.org/10.1007/s00204-015-1570-1>
- Zhang, W.B., Zheng, X.M., Shen, S., Wang, X.H., 2015. Doxorubicin-loaded magnetic nanoparticle clusters for chemo-photothermal treatment of the prostate cancer cell line PC3. BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS 466, 278–282. <https://doi.org/10.1016/j.bbrc.2015.09.036>
- He, S., Qu, M.Z., Feng, Y.J., 2015. Gas-Induced Reversible Dispersion/Aggregation of Graphene. CHEMNANOMAT 1, 438–444. <https://doi.org/10.1002/cnma.201500070>
- Yslas, E.I., Ibarra, L.E., Molina, M.A., Rivarola, C., Barbero, C.A., Bertuzzi, M.L., Rivarola, V.A., 2015. Polyaniline nanoparticles for near-infrared photothermal destruction of cancer cells. JOURNAL OF NANOPARTICLE RESEARCH 17. <https://doi.org/10.1007/s11051-015-3187-y>

- Nanda, S.S., Papaefthymiou, G.C., Yi, D.K., 2015. Functionalization of Graphene Oxide and its Biomedical Applications. *CRITICAL REVIEWS IN SOLID STATE AND MATERIALS SCIENCES* 40, 291–315. <https://doi.org/10.1080/10408436.2014.1002604>
- Chung, I.M., Rahuman, A.A., Marimuthu, S., Kirthi, A. V, Anbarasan, K., Rajakumar, G., 2015. An Investigation of the Cytotoxicity and Caspase-Mediated Apoptotic Effect of Green Synthesized Zinc Oxide Nanoparticles Using *Eclipta prostrata* on Human Liver Carcinoma Cells. *NANOMATERIALS* 5, 1317–1330. <https://doi.org/10.3390/nano5031317>
- Liu, L.J., Wei, Y.C., Zhai, S.D., Chen, Q., Xing, D., 2015. Dihydroartemisinin and transferrin dual-dressed nano-graphene oxide for a pH-triggered chemotherapy. *BIOMATERIALS* 62, 35–46. <https://doi.org/10.1016/j.biomaterials.2015.05.036>
- Hu, S.S., Zeng, Y.X., Yang, S.Y., Qin, H., Cai, H., Wang, J., 2015. Application of Graphene Based Nanotechnology in Stem Cells Research. *JOURNAL OF NANOSCIENCE AND NANOTECHNOLOGY* 15, 6327–6341. <https://doi.org/10.1166/jnn.2015.10844>
- Chen, D.Q., Dougherty, C.A., Zhu, K.C., Hong, H., 2015. Theranostic applications of carbon nanomaterials in cancer: Focus on imaging and cargo delivery. *JOURNAL OF CONTROLLED RELEASE* 210, 230–245. <https://doi.org/10.1016/j.jconrel.2015.04.021>
- Hu, B., Wang, N., Han, L., Chen, M.L., Wang, J.H., 2015. Magnetic Nanohybrids Loaded with Bimetal Core-Shell-Shell Nanorods for Bacteria Capture, Separation, and Near-Infrared Photothermal Treatment. *CHEMISTRY-A EUROPEAN JOURNAL* 21, 6582–6589. <https://doi.org/10.1002/chem.201405960>
- Rajendran, K., Karunagaran, V., Mahanty, B., Sen, S., 2015. Biosynthesis of hematite nanoparticles and its cytotoxic effect on HepG2 cancer cells. *INTERNATIONAL JOURNAL OF BIOLOGICAL MACROMOLECULES* 74, 376–381. <https://doi.org/10.1016/j.ijbiomac.2014.12.028>
- Chaudhuri, B., Bhadra, D., Moroni, L., Pramanik, K., 2015. Myoblast differentiation of human mesenchymal stem cells on graphene oxide and electrospun graphene oxide-polymer composite fibrous meshes: importance of graphene oxide conductivity and dielectric constant on their biocompatibility. *BIOFABRICATION* 7. <https://doi.org/10.1088/1758-5090/7/1/015009>
- Li, W.W., Rong, P.F., Yang, K., Huang, P., Sun, K., Chen, X.Y., 2015. Semimetal nanomaterials of antimony as highly efficient agent for photoacoustic imaging and photothermal therapy. *BIOMATERIALS* 45, 18–26. <https://doi.org/10.1016/j.biomaterials.2014.12.037>
- Khan, M.S., Abdelhamid, H.N., Wu, H.F., 2015. Near infrared (NIR) laser mediated surface activation of graphene oxide nanoflakes for efficient antibacterial, antifungal and wound healing treatment. *COLLOIDS AND SURFACES B-BIOINTERFACES* 127, 281–291. <https://doi.org/10.1016/j.colsurfb.2014.12.049>
- Chen, Y., Shi, J.L., 2015. Mesoporous carbon biomaterials. *SCIENCE CHINA-MATERIALS* 58, 241–257. <https://doi.org/10.1007/s40843-015-0037-2>
- Zhang, H., Wu, H.X., Wang, J., Yang, Y., Wu, D.M., Zhang, Y.J., Zhang, Y., Zhou, Z.G., Yang, S.P., 2015. Graphene oxide-BaGdF₅ nanocomposites for multi-modal imaging and photothermal therapy. *BIOMATERIALS* 42, 66–77. <https://doi.org/10.1016/j.biomaterials.2014.11.055>
- Liang, S.L., Xu, S., Zhang, D., He, J.M., Chu, M.Q., 2015. Reproductive toxicity of nanoscale graphene oxide in male mice. *NANOTOXICOLOGY* 9, 92–105. <https://doi.org/10.3109/17435390.2014.893380>
- Qin, C.C., Fei, J.B., Wang, A.H., Yang, Y., Li, J.B., 2015. Rational assembly of a biointerfaced core@shell nanocomplex towards selective and highly efficient synergistic photothermal/photodynamic therapy. *NANOSCALE* 7, 20197–20210. <https://doi.org/10.1039/c5nr06501a>

- Jang, C., Lee, J.H., Sahu, A., Tae, G., 2015. The synergistic effect of folate and RGD dual ligand of nanographene oxide on tumor targeting and photothermal therapy in vivo. *NANOSCALE* 7, 18584–18594. <https://doi.org/10.1039/c5nr05067g>
- Lee, E.R., Cho, J.W., 2015. Near infrared laser-heated electrospinning and mechanical properties of poly(ethylene terephthalate)/multi-walled carbon nanotube nanofibers. *RSC ADVANCES* 5, 78476–78482. <https://doi.org/10.1039/c5ra14276h>
- Wu, J., Yang, R., Zhang, L., Fan, Z.J., Liu, S.Y., 2015. Cytotoxicity effect of graphene oxide on human MDA-MB-231 cells. *TOXICOLOGY MECHANISMS AND METHODS* 25, 312–319. <https://doi.org/10.3109/15376516.2015.1031415>
- Rahman, M., Akhter, S., Ahmad, M.Z., Ahmad, J., Addo, R.T., Ahmad, F.J., Pichon, C., 2015. Emerging advances in cancer nanotheranostics with graphene nanocomposites: opportunities and challenges. *NANOMEDICINE* 10, 2405–2422. <https://doi.org/10.2217/NNM.15.68>
- Tonelli, F.M.P., Goulart, V.A.M., Gomes, K.N., Ladeira, M.S., Santos, A.K., Lorencon, E., Ladeira, L.O., Resende, R.R., 2015. Graphene-based nanomaterials: biological and medical applications and toxicity. *NANOMEDICINE* 10, 2423–2450. <https://doi.org/10.2217/NNM.15.65>
- Gurunathan, S., Han, J.W., Kim, E.S., Park, J.H., Kim, J.H., 2015. Reduction of graphene oxide by resveratrol: a novel and simple biological method for the synthesis of an effective anticancer nanotherapeutic molecule. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 10, 2951–2969. <https://doi.org/10.2147/IJN.S79879>
- Huang, Y.R., Wei, T., Yu, J., Hou, Y.L., Cai, K.Y., Liang, X.J., 2015. Multifunctional Metal Rattle-Type Nanocarriers for MRI-Guided Photothermal Cancer Therapy. *COLLOIDAL NANOPARTICLES FOR BIOMEDICAL APPLICATIONS* X. <https://doi.org/10.1117/12.2073332>
- Mebrouk, K., Chotard, F., Le Goff-Gaillard, C., Arlot-Bonnemains, Y., Fourmigue, M., Camerel, F., 2015. Water-soluble nickel-bis(dithiolene) complexes as photothermal agents. *CHEMICAL COMMUNICATIONS* 51, 5268–5270. <https://doi.org/10.1039/c4cc08231a>
- Chen, Y.C., Chiu, W.T., Chen, J.C., Chang, C.S., Wang, L.H.C., Lin, H.P., Chang, H.C., 2015. The photothermal effect of silica-carbon hollow sphere-concanavalin A on liver cancer cells. *JOURNAL OF MATERIALS CHEMISTRY B* 3, 2447–2454. <https://doi.org/10.1039/c5tb00056d>
- Duran, N., Martinez, D.S.T., Silveira, C.P., Duran, M., de Moraes, A.C.M., Simoes, M.B., Alves, O.L., Favaro, W.J., 2015. Graphene Oxide: A Carrier for Pharmaceuticals and a Scaffold for Cell Interactions. *CURRENT TOPICS IN MEDICINAL CHEMISTRY* 15, 309–327. <https://doi.org/10.2174/1568026615666150108144217>
- Zhang, Y.X., Li, B., Cao, Y.J., Qin, J.B., Peng, Z.Y., Xiao, Z.Y., Huang, X.J., Zou, R.J., Hu, J.Q., 2015. Na_{0.3}WO₃ nanorods: a multifunctional agent for in vivo dual-model imaging and photothermal therapy of cancer cells. *DALTON TRANSACTIONS* 44, 2771–2779. <https://doi.org/10.1039/c4dt02927e>
- Sun, H., Feng, L.Y., Chen, Y.G., 2015. Research Status and Prospect of Health Risk of Graphene. *PROGRESS IN BIOCHEMISTRY AND BIOPHYSICS* 42, 5–15. <https://doi.org/10.3724/SP.J.1206.2014.00112>
- Lee, H.J., Sanetuntikul, J., Choi, E.S., Lee, B.R., Kim, J.H., Kim, E., Shanmugam, S., 2015. Photothermal cancer therapy using graphitic carbon-coated magnetic particles prepared by one-pot synthesis. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 10, 271–282. <https://doi.org/10.2147/IJN.S73128>
- Chen, Y.J., Yu, H.F., Quan, M.H., Zhang, L.Y., Yang, H., Lu, Y.F., 2015. Photothermal effect of azopyridine compounds and their applications. *RSC ADVANCES* 5, 4675–4680. <https://doi.org/10.1039/c4ra11844h>
- Hu, Z., Li, J., Huang, Y.D., Chen, L., Li, Z.H., 2015. Functionalized graphene/C-60 nanohybrid for targeting photothermally enhanced photodynamic therapy. *RSC ADVANCES* 5, 654–664. <https://doi.org/10.1039/c4ra13427c>

- Shen, S., Wang, S., Zheng, R., Zhu, X.Y., Jiang, X.G., Fu, D.L., Yang, W.L., 2015. Magnetic nanoparticle clusters for photothermal therapy with near-infrared irradiation. *BIOMATERIALS* 39, 67–74. <https://doi.org/10.1016/j.biomaterials.2014.10.064>
- Chen, Y., Tan, C.L., Zhang, H., Wang, L.Z., 2015. Two-dimensional graphene analogues for biomedical applications. *CHEMICAL SOCIETY REVIEWS* 44, 2681–2701. <https://doi.org/10.1039/c4cs00300d>
- Tang, L.H., Wang, Y., Li, J.H., 2015. The graphene/nucleic acid nanobiointerface. *CHEMICAL SOCIETY REVIEWS* 44, 6954–6980. <https://doi.org/10.1039/c4cs00519h>
- Ferrari, A.C., Bonaccorso, F., Fal'ko, V., Novoselov, K.S., Roche, S., Boggild, P., Borini, S., Koppens, F.H.L., Palermo, V., Pugno, N., Garrido, J.A., Sordan, R., Bianco, A., Ballerini, L., Prato, M., Lidorikis, E., Kivioja, J., Marinelli, C., Ryhanen, T., Morpurgo, A., Coleman, J.N., Nicolosi, V., Colombo, L., Fert, A., Garcia-Hernandez, M., Bachtold, A., Schneider, G.F., Guinea, F., Dekker, C., Barbone, M., Sun, Z.P., Galiotis, C., Grigorenko, A.N., Konstantatos, G., Kis, A., Katsnelson, M., Vandersypen, L., Loiseau, A., Morandi, V., Neumaier, D., Treossi, E., Pellegrini, V., Polini, M., Tredicucci, A., Williams, G.M., Hong, B.H., Ahn, J.H., Kim, J.M., Zirath, H., van Wees, B.J., van der Zant, H., Occhipinti, L., Di Matteo, A., Kinloch, I.A., Seyller, T., Quesnel, E., Feng, X.L., Teo, K., Rupasinghe, N., Hakonen, P., Neil, S.R.T., Tannock, Q., Lofwander, T., Kinaret, J., 2015. Science and technology roadmap for graphene, related two-dimensional crystals, and hybrid systems. *NANOSCALE* 7, 4598–4810. <https://doi.org/10.1039/c4nr01600a>
- Wang, H., Cao, G.X., Gai, Z., Hong, K.L., Banerjee, P., Zhou, S.Q., 2015. Magnetic/NIR-responsive drug carrier, multicolor cell imaging, and enhanced photothermal therapy of gold capped magnetite-fluorescent carbon hybrid nanoparticles. *NANOSCALE* 7, 7885–7895. <https://doi.org/10.1039/c4nr07335e>
- Nurunnabi, M., Parvez, K., Nafiujjaman, M., Revuri, V., Khan, H.A., Feng, X.L., Lee, Y.K., 2015. Bioapplication of graphene oxide derivatives: drug/gene delivery, imaging, polymeric modification, toxicology, therapeutics and challenges. *RSC ADVANCES* 5, 42141–42161. <https://doi.org/10.1039/c5ra04756k>
- Zhang, L., Li, Y.C., Jin, Z.X., Chan, K.M., Yu, J.C., 2015. Mesoporous carbon/CuS nanocomposites for pH-dependent drug delivery and near-infrared chemo-photothermal therapy. *RSC ADVANCES* 5, 93226–93233. <https://doi.org/10.1039/c5ra19458j>
- Liu, R.F., Jing, L.J., Peng, D., Li, Y., Tian, J., Dai, Z.F., 2015. Manganese (II) Chelate Functionalized Copper Sulfide Nanoparticles for Efficient Magnetic Resonance/Photoacoustic Dual-Modal Imaging Guided Photothermal Therapy. *THERANOSTICS* 5, 1144–1153. <https://doi.org/10.7150/thno.11754>
- Cheng, L., Wang, C., Feng, L.Z., Yang, K., Liu, Z., 2014. Functional Nanomaterials for Phototherapies of Cancer. *CHEMICAL REVIEWS* 114, 10869–10939. <https://doi.org/10.1021/cr400532z>
- Caputo, F., De Nicola, M., Ghibelli, L., 2014. Pharmacological potential of bioactive engineered nanomaterials. *BIOCHEMICAL PHARMACOLOGY* 92, 112–130. <https://doi.org/10.1016/j.bcp.2014.08.015>
- Nezakati, T., Cousins, B.G., Seifalian, A.M., 2014. Toxicology of chemically modified graphene-based materials for medical application. *ARCHIVES OF TOXICOLOGY* 88, 1987–2012. <https://doi.org/10.1007/s00204-014-1361-0>
- Zeng, Y.Z., Lu, J.Q., 2014. Optothermally Responsive Nanocomposite Generating Mechanical Forces for Cells Enabled by Few-Walled Carbon Nanotubes. *ACS NANO* 8, 11695–11706. <https://doi.org/10.1021/nn505042b>
- Wang, S., Zhang, Q., Luo, X.F., Li, J., He, H., Yang, F., Di, Y., Jin, C., Jiang, X.G., Shen, S., Fu, D.L., 2014. Magnetic graphene-based nanotheranostic agent for dual-modality mapping guided photothermal therapy in regional lymph nodal metastasis of pancreatic cancer. *BIOMATERIALS* 35, 9473–9483. <https://doi.org/10.1016/j.biomaterials.2014.07.064>

- Zhu, X.L., Huang, S.N., Huang, H.Q., Zhang, Y.J., Xie, Y.X., Hou, L., Zhang, H.J., Shi, J.J., Zhang, Z.Z., 2014. In vitro and in vivo anti-cancer effects of targeting and photothermal sensitive solid lipid nanoparticles. *JOURNAL OF DRUG TARGETING* 22, 822–828. <https://doi.org/10.3109/1061186X.2014.931405>
- Goncalves, G., Vila, M., Bdkin, I., de Andres, A., Emami, N., Ferreira, R.A.S., Carlos, L.D., Gracio, J., Marques, P., 2014. Breakdown into nanoscale of graphene oxide: Confined hot spot atomic reduction and fragmentation. *SCIENTIFIC REPORTS* 4. <https://doi.org/10.1038/srep06735>
- Piao, J.G., Wang, L.M., Gao, F., You, Y.Z., Xiong, Y.J., Yang, L.H., 2014. Erythrocyte Membrane Is an Alternative Coating to Polyethylene Glycol for Prolonging the Circulation Lifetime of Gold Nanocages for Photothermal Therapy. *ACS NANO* 8, 10414–10425. <https://doi.org/10.1021/nn503779d>
- Huang, Y.R., Wei, T., Yu, J., Hou, Y.L., Cai, K.Y., Liang, X.J., 2014. Multifunctional Metal Rattle-Type Nanocarriers for MRI-Guided Photothermal Cancer Therapy. *MOLECULAR PHARMACEUTICS* 11, 3386–3394. <https://doi.org/10.1021/mp500006z>
- Chen, G.Y., Chen, C.L., Tuan, H.Y., Yuan, P.X., Li, K.C., Yang, H.J., Hu, Y.C., 2014. Graphene Oxide Triggers Toll-Like Receptors/Autophagy Responses In Vitro and Inhibits Tumor Growth In Vivo. *ADVANCED HEALTHCARE MATERIALS* 3, 1486–1495. <https://doi.org/10.1002/adhm.201300591>
- Yoo, J., Kim, J., Baek, S., Park, Y., Im, H., Kim, J., 2014. Cell reprogramming into the pluripotent state using graphene based substrates. *BIOMATERIALS* 35, 8321–8329. <https://doi.org/10.1016/j.biomaterials.2014.05.096>
- Sawdon, A., Weydemeyer, E., Peng, C.A., 2014. Antitumor Therapy Using Nanomaterial-Mediated Thermolysis. *JOURNAL OF BIOMEDICAL NANOTECHNOLOGY* 10, 1894–1917. <https://doi.org/10.1166/jbn.2014.1917>
- Pekkanen, A.M., DeWitt, M.R., Rylander, M.N., 2014. Nanoparticle Enhanced Optical Imaging and Phototherapy of Cancer. *JOURNAL OF BIOMEDICAL NANOTECHNOLOGY* 10, 1677–1712. <https://doi.org/10.1166/jbn.2014.1988>
- Dong, H.Q., Dong, C.Y., Ren, T.B., Li, Y.Y., Shi, D.L., 2014. Surface-Engineered Graphene-Based Nanomaterials for Drug Delivery. *JOURNAL OF BIOMEDICAL NANOTECHNOLOGY* 10, 2086–2106. <https://doi.org/10.1166/jbn.2014.1989>
- Zhang, W.J., Liu, J.L., Zhang, Q., Li, X.D., Yu, S.H., Yang, X.G., Kong, J., Pan, W.S., 2014. Enhanced cellular uptake and anti-proliferating effect of chitosan hydrochlorides modified genistein loaded NLC on human lens epithelial cells. *INTERNATIONAL JOURNAL OF PHARMACEUTICS* 471, 118–126. <https://doi.org/10.1016/j.ijpharm.2014.05.030>
- Komatsu, H., Daimon, Y., Kawakami, K., Kanai, M., Hill, J.P., Ariga, K., 2014. Reaction mediated artificial cell termination: control of vesicle viability using Rh(I)-catalyzed hydrogenation. *PHYSICAL CHEMISTRY CHEMICAL PHYSICS* 16, 16454–16457. <https://doi.org/10.1039/c4cp02255f>
- Xing, X.B., Zheng, J.P., Li, F.J., Sun, C., Cai, X., Zhu, D.B., Lei, L., Wu, T., Zhou, B., Evans, J.L., Chen, Z.Y., 2014. Dynamic behaviors of approximately ellipsoidal microbubbles photothermally generated by a graphene oxide-microheater. *SCIENTIFIC REPORTS* 4. <https://doi.org/10.1038/srep06086>
- Major, R., Sanak, M., Mzyk, A., Lipinska, L., Kot, M., Lacki, P., Bruckert, F., Major, B., 2014. Graphene based porous coatings with antibacterial and antithrombogenic function-Materials and design. *ARCHIVES OF CIVIL AND MECHANICAL ENGINEERING* 14, 540–549. <https://doi.org/10.1016/j.acme.2014.04.012>
- Topete, A., Alatorre-Meda, M., Villar-Alvarez, E.M., Carregal-Romero, S., Barbosa, S., Parak, W.J., Taboada, P., Mosquera, V., 2014. Polymeric-Gold Nanohybrids for Combined Imaging and Cancer Therapy. *ADVANCED HEALTHCARE MATERIALS* 3, 1309–1325. <https://doi.org/10.1002/adhm.201400023>

- Jia, K., Marks, R.S., Ionescu, R.E., 2014. Influence of carbon-based nanomaterials on lux-bioreporter *Escherichia coli*. *TALANTA* 126, 208–213. <https://doi.org/10.1016/j.talanta.2014.03.024>
- Yoon, O.J., Kim, I., Sohn, I.Y., Kieu, T.T., Lee, N.E., 2014. Toxicity of graphene nanoflakes evaluated by cell-based electrochemical impedance biosensing. *JOURNAL OF BIOMEDICAL MATERIALS RESEARCH PART A* 102, 2288–2294. <https://doi.org/10.1002/jbm.a.34886>
- Li, J.L., Hou, X.L., Bao, H.C., Sun, L., Tang, B., Wang, J.F., Wang, X.G., Gu, M., 2014. Graphene oxide nanoparticles for enhanced photothermal cancer cell therapy under the irradiation of a femtosecond laser beam. *JOURNAL OF BIOMEDICAL MATERIALS RESEARCH PART A* 102, 2181–2188. <https://doi.org/10.1002/jbm.a.34871>
- Hong, T., Cao, Y.H., Ying, D., Xu, Y.Q., 2014. Thermal and optical properties of freestanding flat and stacked single-layer graphene in aqueous media. *APPLIED PHYSICS LETTERS* 104. <https://doi.org/10.1063/1.4881136>
- Yuan, Y., Jiang, W., Wang, Y.J., Shen, P., Li, F.S., Li, P.Y., Zhao, F., Gao, H.X., 2014. Hydrothermal preparation of Fe₂O₃/graphene nanocomposite and its enhanced catalytic activity on the thermal decomposition of ammonium perchlorate. *APPLIED SURFACE SCIENCE* 303, 354–359. <https://doi.org/10.1016/j.apsusc.2014.03.005>
- Kalbacova, M.H., Verdanova, M., Broz, A., Vetushka, A., Fejfar, A., Kalbac, M., 2014. Modulated surface of single-layer graphene controls cell behavior. *CARBON* 72, 207–214. <https://doi.org/10.1016/j.carbon.2014.02.004>
- Adeli, M., Madani, F., Sasanpour, P., 2014. Synthesis of graphene/gold hybrid nanomaterials by poly(ethylene glycol) linkers. *SCIENTIA IRANICA* 21, 1163–1173.
- Yeh, C.C., Chen, D.H., 2014. Ni/reduced graphene oxide nanocomposite as a magnetically recoverable catalyst with near infrared photothermally enhanced activity. *APPLIED CATALYSIS B-ENVIRONMENTAL* 150, 298–304. <https://doi.org/10.1016/j.apcatb.2013.12.040>
- Mattei, T.A., Rehman, A.A., 2014. Technological Developments and Future Perspectives on Graphene-Based Metamaterials: A Primer for Neurosurgeons. *NEUROSURGERY* 74, 499–516. <https://doi.org/10.1227/NEU.0000000000000302>
- Liu, Z., Cheng, L., Zhang, L., Yang, Z.B., Liu, Z., Fang, J.X., 2014. Sub-100 nm hollow Au-Ag alloy urchin-shaped nanostructure with ultrahigh density of nanotips for photothermal cancer therapy. *BIOMATERIALS* 35, 4099–4107. <https://doi.org/10.1016/j.biomaterials.2014.01.053>
- Miao, W., Shim, G., Lee, S., Oh, Y.K., 2014. Structure-dependent photothermal anticancer effects of carbon-based photoresponsive nanomaterials. *BIOMATERIALS* 35, 4058–4065. <https://doi.org/10.1016/j.biomaterials.2014.01.043>
- Zhou, X.F., Liang, F., 2014. Application of Graphene/Graphene Oxide in Biomedicine and Biotechnology. *CURRENT MEDICINAL CHEMISTRY* 21, 855–869. <https://doi.org/10.2174/0929867320666131119124325>
- Topete, A., Alatorre-Meda, M., Iglesias, P., Villar-Alvarez, E.M., Barbosa, S., Costoya, J.A., Taboada, P., Mosquera, V., 2014. Fluorescent Drug-Loaded, Polymeric-Based, Branched Gold Nanoshells for Localized Multimodal Therapy and Imaging of Tumoral Cells. *ACS NANO* 8, 2725–2738. <https://doi.org/10.1021/nn406425h>
- Zhang, X.D., Chen, J., Min, Y., Park, G.B., Shen, X., Song, S.S., Sun, Y.M., Wang, H., Long, W., Xie, J.P., Gao, K., Zhang, L.F., Fan, S.J., Fan, F.Y., Jeong, U., 2014. Metabolizable Bi₂Se₃ Nanoplates: Biodistribution, Toxicity, and Uses for Cancer Radiation Therapy and Imaging. *ADVANCED FUNCTIONAL MATERIALS* 24, 1718–1729. <https://doi.org/10.1002/adfm.201302312>

- Zhang, X., Li, M., Wang, Y.B., Cheng, Y., Zheng, Y.F., Xi, T.F., Wei, S.C., 2014. Cell response of nanographene platelets to human osteoblast-like MG63 cells. *JOURNAL OF BIOMEDICAL MATERIALS RESEARCH PART A* 102, 732–742. <https://doi.org/10.1002/jbm.a.34751>
- Li, Y.Y., Liu, J.Q., Dong, H.Q., Liu, G.Z., Hu, H.Q., 2014. Engineering of a Pluronic F127 functionalized magnetite/graphene nanohybrid for chemophototherapy. *NANOTECHNOLOGY* 25. <https://doi.org/10.1088/0957-4484/25/6/065602>
- Vila, M., Matesanz, M.C., Goncalves, G., Feito, M.J., Linares, J., Marques, P., Portoles, M.T., Vallet-Regi, M., 2014. Triggering cell death by nanographene oxide mediated hyperthermia. *NANOTECHNOLOGY* 25. <https://doi.org/10.1088/0957-4484/25/3/035101>
- Liu, F., Zhang, Y., Yu, J.H., Wang, S.W., Ge, S.G., Song, X.R., 2014. Application of ZnO/graphene and S6 aptamers for sensitive photoelectrochemical detection of SK-BR-3 breast cancer cells based on a disposable indium tin oxide device. *BIOSENSORS & BIOELECTRONICS* 51, 413–420. <https://doi.org/10.1016/j.bios.2013.07.066>
- Jiang, B.P., Hu, L.F., Wang, D.J., Ji, S.C., Shen, X.C., Liang, H., 2014. Graphene loading water-soluble phthalocyanine for dual-modality photothermal/photodynamic therapy via a one-step method. *JOURNAL OF MATERIALS CHEMISTRY B* 2, 7141–7148. <https://doi.org/10.1039/c4tb01038h>
- Matteini, P., Tatini, F., Cavigli, L., Ottaviano, S., Ghini, G., Pini, R., 2014. Graphene as a photothermal switch for controlled drug release. *NANOSCALE* 6, 7947–7953. <https://doi.org/10.1039/c4nr01622j>
- Wu, S.L., Zhao, X.D., Cui, Z.G., Zhao, C.T., Wang, Y.Z., Du, L., Li, Y.H., 2014. Cytotoxicity of graphene oxide and graphene oxide loaded with doxorubicin on human multiple myeloma cells. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 9, 1413–1421. <https://doi.org/10.2147/IJN.S57946>
- Jung, H.S., Kong, W.H., Sung, D.K., Lee, M.Y., Beack, S.E., Keum, D.H., Kim, K.S., Yun, S.H., Hahn, S.K., 2014. Nanographene Oxide-Hyaluronic Acid Conjugate for Photothermal Ablation Therapy of Skin Cancer. *ACS NANO* 8, 260–268. <https://doi.org/10.1021/nn405383a>
- Lim, D.J., Sim, M., Oh, L., Lim, K., Park, H., 2014. Carbon-based drug delivery carriers for cancer therapy. *ARCHIVES OF PHARMACAL RESEARCH* 37, 43–52. <https://doi.org/10.1007/s12272-013-0277-1>
- Kliscic, D., Radovanovic, I., Bezanic, N., Stamenic, L., Popovic, I., IEEE, 2014. Voltage Controlled Power Characteristics of a Custom Made Laser Diode Assembly. 2014 22ND TELECOMMUNICATIONS FORUM TELFOR (TELFOR).
- Verma, J., Lal, S., Van Noorden, C.J.F., 2014. Nanoparticles for hyperthermic therapy: synthesis strategies and applications in glioblastoma. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 9, 2863–2877. <https://doi.org/10.2147/IJN.S57501>
- Zhou, Z.G., Wang, J., Liu, W., Yu, C., Kong, B., Sun, Y.A., Yang, H., Yang, S.P., Wang, W., 2014. PEGylated nickel carbide nanocrystals as efficient near-infrared laser induced photothermal therapy for treatment of cancer cells in vivo. *NANOSCALE* 6, 12591–12600. <https://doi.org/10.1039/c4nr03727h>
- Xing, X.B., Zheng, J.P., Sun, C., Li, F.J., Zhu, D.B., Lei, L., Cai, X., Wu, T., 2013. Graphene oxide-deposited microfiber: a new photothermal device for various microbubble generation. *OPTICS EXPRESS* 21, 31862–31871. <https://doi.org/10.1364/OE.21.031862>
- Liu, J.Q., Cui, L., Losic, D., 2013. Graphene and graphene oxide as new nanocarriers for drug delivery applications. *ACTA BIOMATERIALIA* 9, 9243–9257. <https://doi.org/10.1016/j.actbio.2013.08.016>
- Singh, R., Torti, S. V., 2013. Carbon nanotubes in hyperthermia therapy. *ADVANCED DRUG DELIVERY REVIEWS* 65, 2045–2060. <https://doi.org/10.1016/j.addr.2013.08.001>

- Li, J.L., Tang, B., Yuan, B., Sun, L., Wang, X.G., 2013. A review of optical imaging and therapy using nanosized graphene and graphene oxide. *BIOMATERIALS* 34, 9519–9534. <https://doi.org/10.1016/j.biomaterials.2013.08.066>
- Akhavan, O., Ghaderi, E., 2013. Graphene Nanomesh Promises Extremely Efficient In Vivo Photothermal Therapy. *SMALL* 9, 3593–3601. <https://doi.org/10.1002/smll.201203106>
- Kleut, D.N., Markovi, Z.M., Babic, B.M., Antunovic, I.D.H., Milosavljevic, M.S., Dramicanin, M.D., Markovic, B.M.T., 2013. Raman spectroscopy study of carbon-doped resorcinol-formaldehyde thin films. *PHYSICA SCRIPTA* T157. <https://doi.org/10.1088/0031-8949/2013/T157/014039>
- Yoon, O.J., Kim, C.H., Sohn, I.Y., Lee, N.E., 2013. Toxicity analysis of graphene nanoflakes by cell-based electrochemical sensing using an electrode modified with nanocomposite of graphene and Nafion. *SENSORS AND ACTUATORS B-CHEMICAL* 188, 454–461. <https://doi.org/10.1016/j.snb.2013.07.039>
- Chung, C., Kim, Y.K., Shin, D., Ryoo, S.R., Hong, B.H., Min, D.H., 2013. Biomedical Applications of Graphene and Graphene Oxide. *ACCOUNTS OF CHEMICAL RESEARCH* 46, 2211–2224. <https://doi.org/10.1021/ar300159f>
- Wang, C.Y., Mallela, J., Garapati, U.S., Ravi, S., Chinnasamy, V., Girard, Y., Howell, M., Mohapatra, S., 2013. A chitosan-modified graphene nanogel for noninvasive controlled drug release. *NANOMEDICINE-NANOTECHNOLOGY BIOLOGY AND MEDICINE* 9, 903–911. <https://doi.org/10.1016/j.nano.2013.01.003>
- Zhu, X., Sun, L., Chen, Y.Y., Ye, Z.H., Shen, Z.M., Li, G.X., 2013. Combination of cascade chemical reactions with graphene-DNA interaction to develop new strategy for biosensor fabrication. *BIOSENSORS & BIOELECTRONICS* 47, 32–37. <https://doi.org/10.1016/j.bios.2013.02.039>
- Chen, Z.G., Kuang, X.Y., Song, L.L., Tian, Q.W., Hu, J.Q., 2013. Research Progress in NIR-Light-Driven Nanomaterials and Nanodevices. *CHINESE JOURNAL OF INORGANIC CHEMISTRY* 29, 1574–1590. <https://doi.org/10.3969/j.issn.1001-4861.2013.00.305>
- Goncalves, G., Vila, M., Portoles, M.T., Vallet-Regi, M., Gracio, J., Marques, P., 2013. Nano-Graphene Oxide: A Potential Multifunctional Platform for Cancer Therapy. *ADVANCED HEALTHCARE MATERIALS* 2, 1072–1090. <https://doi.org/10.1002/adhm.201300023>
- Sahu, A., Choi, W.I., Lee, J.H., Tae, G., 2013. Graphene oxide mediated delivery of methylene blue for combined photodynamic and photothermal therapy. *BIOMATERIALS* 34, 6239–6248. <https://doi.org/10.1016/j.biomaterials.2013.04.066>
- Chen, Y.C., Huang, X.C., Luo, Y.L., Chang, Y.C., Hsieh, Y.Z., Hsu, H.Y., 2013. Non-metallic nanomaterials in cancer theranostics: a review of silica- and carbon-based drug delivery systems. *SCIENCE AND TECHNOLOGY OF ADVANCED MATERIALS* 14. <https://doi.org/10.1088/1468-6996/14/4/044407>
- Lu, J.Y., He, Y.S., Cheng, C., Wang, Y., Qiu, L., Li, D., Zou, D.R., 2013. Self-Supporting Graphene Hydrogel Film as an Experimental Platform to Evaluate the Potential of Graphene for Bone Regeneration. *ADVANCED FUNCTIONAL MATERIALS* 23, 3494–3502. <https://doi.org/10.1002/adfm.201203637>
- Lai, B.H., Chen, D.H., 2013. LaB6 nanoparticles with carbon-doped silica coating for fluorescence imaging and near-IR photothermal therapy of cancer cells. *ACTA BIOMATERIALIA* 9, 7556–7563. <https://doi.org/10.1016/j.actbio.2013.03.034>
- Lai, B.H., Chen, D.H., 2013. Vancomycin-modified LaB6@SiO2/Fe3O4 composite nanoparticles for near-infrared photothermal ablation of bacteria. *ACTA BIOMATERIALIA* 9, 7573–7579. <https://doi.org/10.1016/j.actbio.2013.03.023>

- Lai, B.H., Lin, Y.R., Chen, D.H., 2013. Fabrication of LaB₆@SiO₂/Au composite nanoparticles as a catalyst with near infrared photothermally enhanced activity. *CHEMICAL ENGINEERING JOURNAL* 223, 418–424. <https://doi.org/10.1016/j.cej.2013.02.109>
- Mao, H.Y., Laurent, S., Chen, W., Akhavan, O., Imani, M., Ashkarran, A.A., Mahmoudi, M., 2013. Graphene: Promises, Facts, Opportunities, and Challenges in Nanomedicine. *CHEMICAL REVIEWS* 113, 3407–3424. <https://doi.org/10.1021/cr300335p>
- Hu, X.G., Zhou, Q.X., 2013. Health and Ecosystem Risks of Graphene. *CHEMICAL REVIEWS* 113, 3815–3835. <https://doi.org/10.1021/cr300045n>
- Bitounis, D., Ali-Boucetta, H., Hong, B.H., Min, D.H., Kostarelos, K., 2013. Prospects and Challenges of Graphene in Biomedical Applications. *ADVANCED MATERIALS* 25, 2258–2268. <https://doi.org/10.1002/adma.201203700>
- Chen, Z.G., Wang, Q., Wang, H.L., Zhang, L.S., Song, G.S., Song, L.L., Hu, J.Q., Wang, H.Z., Liu, J.S., Zhu, M.F., Zhao, D.Y., 2013. Ultrathin PEGylated W18O49 Nanowires as a New 980 nm-Laser-Driven Photothermal Agent for Efficient Ablation of Cancer Cells In Vivo. *ADVANCED MATERIALS* 25, 2095–2100. <https://doi.org/10.1002/adma.201204616>
- Miao, W., Shim, G., Lee, S., Lee, S., Choe, Y.S., Oh, Y.K., 2013. Safety and tumor tissue accumulation of pegylated graphene oxide nanosheets for co-delivery of anticancer drug and photosensitizer. *BIOMATERIALS* 34, 3402–3410. <https://doi.org/10.1016/j.biomaterials.2013.01.010>
- Akhavan, O., Ghaderi, E., Emamy, H., Akhavan, F., 2013. Genotoxicity of graphene nanoribbons in human mesenchymal stem cells. *CARBON* 54, 419–431. <https://doi.org/10.1016/j.carbon.2012.11.058>
- Heister, E., Brunner, E.W., Dieckmann, G.R., Jurewicz, I., Dalton, A.B., 2013. Are Carbon Nanotubes a Natural Solution? Applications in Biology and Medicine. *ACS APPLIED MATERIALS & INTERFACES* 5, 1870–1891. <https://doi.org/10.1021/am302902d>
- Wang, Y., Wang, K.Y., Zhao, J.F., Liu, X.G., Bu, J., Yan, X.Y., Huang, R.Q., 2013. Multifunctional Mesoporous Silica-Coated Graphene Nanosheet Used for Chemo-Photothermal Synergistic Targeted Therapy of Glioma. *JOURNAL OF THE AMERICAN CHEMICAL SOCIETY* 135, 4799–4804. <https://doi.org/10.1021/ja312221g>
- Das, T.K., Prusty, S., 2013. Graphene-Based Polymer Composites and Their Applications. *POLYMER-PLASTICS TECHNOLOGY AND ENGINEERING* 52, 319–331. <https://doi.org/10.1080/03602559.2012.751410>
- Qin, X.C., Guo, Z.Y., Liu, Z.M., Zhang, W., Wan, M.M., Yang, B.W., 2013. Folic acid-conjugated graphene oxide for cancer targeted chemo-photothermal therapy. *JOURNAL OF PHOTOCHEMISTRY AND PHOTOBIOLOGY B-BIOLOGY* 120, 156–162. <https://doi.org/10.1016/j.jphotobiol.2012.12.005>
- Yu, C., Wo, F.J., Shao, Y.X., Dai, X.Y., Chu, M.Q., 2013. Bovine Serum Albumin Nanospheres Synchronously Encapsulating “Gold Selenium/Gold” Nanoparticles and Photosensitizer for High-Efficiency Cancer Phototherapy. *APPLIED BIOCHEMISTRY AND BIOTECHNOLOGY* 169, 1566–1578. <https://doi.org/10.1007/s12010-012-0078-x>
- Tan, X.F., Feng, L.Z., Zhang, J., Yang, K., Zhang, S., Liu, Z., Peng, R., 2013. Functionalization of Graphene Oxide Generates a Unique Interface for Selective Serum Protein Interactions. *ACS APPLIED MATERIALS & INTERFACES* 5, 1370–1377. <https://doi.org/10.1021/am302706g>
- Chen, C.J., Chen, D.H., 2013. Preparation and near-infrared photothermal conversion property of cesium tungsten oxide nanoparticles. *NANOSCALE RESEARCH LETTERS* 8. <https://doi.org/10.1186/1556-276X-8-57>

- Chu, M.Q., Peng, J.L., Zhao, J.J., Liang, S.L., Shao, Y.X., Wu, Q., 2013. Laser light triggered-activated carbon nanosystem for cancer therapy. *BIOMATERIALS* 34, 1820–1832. <https://doi.org/10.1016/j.biomaterials.2012.11.027>
- Wu, M.C., Deokar, A.R., Liao, J.H., Shih, P.Y., Ling, Y.C., 2013. Graphene-Based Photothermal Agent for Rapid and Effective Killing of Bacteria. *ACS NANO* 7, 1281–1290. <https://doi.org/10.1021/nn304782d>
- Jeyaraj, M., Rajesh, M., Arun, R., MubarakAli, D., Sathishkumar, G., Sivanandhan, G., Dev, G.K., Manickavasagam, M., Premkumar, K., Thajuddin, N., Ganapathi, A., 2013. An investigation on the cytotoxicity and caspase-mediated apoptotic effect of biologically synthesized silver nanoparticles using *Podophyllum hexandrum* on human cervical carcinoma cells. *COLLOIDS AND SURFACES B-BIOINTERFACES* 102, 708–717. <https://doi.org/10.1016/j.colsurfb.2012.09.042>
- Feng, L.Y., Wu, L., Qu, X.G., 2013. New Horizons for Diagnostics and Therapeutic Applications of Graphene and Graphene Oxide. *ADVANCED MATERIALS* 25, 168–186. <https://doi.org/10.1002/adma.201203229>
- Hu, Z., Li, J., Li, C.Y., Zhao, S.J., Li, N., Wang, Y.F., Wei, F., Chen, L., Huang, Y.D., 2013. Folic acid-conjugated graphene-ZnO nanohybrid for targeting photodynamic therapy under visible light irradiation. *JOURNAL OF MATERIALS CHEMISTRY B* 1, 5003–5013. <https://doi.org/10.1039/c3tb20849d>
- Jaworski, S., Sawosz, E., Grodzik, M., Winnicka, A., Prasek, M., Wierzbicki, M., Chwalibog, A., 2013. In vitro evaluation of the effects of graphene platelets on glioblastoma multiforme cells. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 8, 413–420. <https://doi.org/10.2147/IJN.S39456>
- Yue, Z.G., Lv, P.P., Yue, H., Gao, Y.J., Ma, D., Wei, W., Ma, G.H., 2013. Inducible graphene oxide probe for high-specific tumor diagnosis. *CHEMICAL COMMUNICATIONS* 49, 3902–3904. <https://doi.org/10.1039/c3cc40499d>
- Shen, S., Ren, J.F., Zhu, X.Y., Pang, Z.Q., Lu, X.H., Deng, C.H., Zhang, R., Jiang, X.G., 2013. Monodisperse magnetites anchored onto carbon nanotubes: a platform for cell imaging, magnetic manipulation and enhanced photothermal treatment of tumors. *JOURNAL OF MATERIALS CHEMISTRY B* 1, 1939–1946. <https://doi.org/10.1039/c3tb00543g>
- Zhang, G., Chang, H.C., Amatore, C., Chen, Y., Jiang, H., Wang, X.M., 2013. Apoptosis induction and inhibition of drug resistant tumor growth in vivo involving daunorubicin-loaded graphene-gold composites. *JOURNAL OF MATERIALS CHEMISTRY B* 1, 493–499. <https://doi.org/10.1039/c2tb00378c>
- Liu, Z.M., Li, S.X., Hu, C.F., Zhang, W., Zhong, H.Q., Guo, Z.Y., 2013. pH-dependent surface-enhanced Raman scattering of aromatic molecules on graphene oxide. *JOURNAL OF RAMAN SPECTROSCOPY* 44, 75–80. <https://doi.org/10.1002/jrs.4142>
- Hou, C.Y., Quan, H.C., Duan, Y.R., Zhang, Q.H., Wang, H.Z., Li, Y.G., 2013. Facile synthesis of water-dispersible Cu₂O nanocrystal-reduced graphene oxide hybrid as a promising cancer therapeutic agent. *NANOSCALE* 5, 1227–1232. <https://doi.org/10.1039/c2nr32938g>
- Yang, K., Feng, L.Z., Shi, X.Z., Liu, Z., 2013. Nano-graphene in biomedicine: theranostic applications. *CHEMICAL SOCIETY REVIEWS* 42, 530–547. <https://doi.org/10.1039/c2cs35342c>
- Hu, C.F., Rong, J.H., Cui, J.H., Yang, Y.H., Yang, L.F., Wang, Y.L., Liu, Y.L., 2013. Fabrication of a graphene oxide-gold nanorod hybrid material by electrostatic self-assembly for surface-enhanced Raman scattering. *CARBON* 51, 255–264. <https://doi.org/10.1016/j.carbon.2012.08.051>
- Peretz, S., Regev, O., 2012. Carbon nanotubes as nanocarriers in medicine. *CURRENT OPINION IN COLLOID & INTERFACE SCIENCE* 17, 360–368. <https://doi.org/10.1016/j.cocis.2012.09.001>
- Jastrzebska, A.M., Kurtycz, P., Olszyna, A.R., 2012. Recent advances in graphene family materials toxicity investigations. *JOURNAL OF NANOPARTICLE RESEARCH* 14. <https://doi.org/10.1007/s11051-012-1320-8>

- Vila, M., Portoles, M.T., Marques, P., Feito, M.J., Matesanz, M.C., Ramirez-Santillan, C., Goncalves, G., Cruz, S.M.A., Nieto, A., Vallet-Regi, M., 2012. Cell uptake survey of pegylated nanographene oxide. *NANOTECHNOLOGY* 23. <https://doi.org/10.1088/0957-4484/23/46/465103>
- Akhavan, O., Ghaderi, E., Rahimi, K., 2012. Adverse effects of graphene incorporated in TiO₂ photocatalyst on minuscule animals under solar light irradiation. *JOURNAL OF MATERIALS CHEMISTRY* 22, 23260–23266. <https://doi.org/10.1039/c2jm35228a>
- Kalbacova, M., Broz, A., Kalbac, M., 2012. Influence of the fetal bovine serum proteins on the growth of human osteoblast cells on graphene. *JOURNAL OF BIOMEDICAL MATERIALS RESEARCH PART A* 100A, 3001–3007. <https://doi.org/10.1002/jbm.a.34231>
- Kleut, D., Jovanovic, S., Markovic, Z., Kepic, D., Tosic, D., Romcevic, N., Marinovic-Cincovic, M., Dramicanin, M., Holclajtner-Antunovic, I., Pavlovic, V., Drazic, G., Milosavljevic, M., Markovic, B.T., 2012. Comparison of structural properties of pristine and gamma irradiated single-wall carbon nanotubes: Effects of medium and irradiation dose. *MATERIALS CHARACTERIZATION* 72, 37–45. <https://doi.org/10.1016/j.matchar.2012.07.002>
- Zhou, H.J., Zhao, K., Li, W., Yang, N., Liu, Y., Chen, C.Y., Wei, T.T., 2012. The interactions between pristine graphene and macrophages and the production of cytokines/chemokines via TLR- and NF-kappa B-related signaling pathways. *BIOMATERIALS* 33, 6933–6942. <https://doi.org/10.1016/j.biomaterials.2012.06.064>
- Kesharwani, P., Ghanghoria, R., Jain, N.K., 2012. Carbon nanotube exploration in cancer cell lines. *DRUG DISCOVERY TODAY* 17, 1023–1030. <https://doi.org/10.1016/j.drudis.2012.05.003>
- Wei, W.L., Qu, X.G., 2012. Extraordinary Physical Properties of Functionalized Graphene. *SMALL* 8, 2138–2151. <https://doi.org/10.1002/sml.201200104>
- Yu, J.G., Jiao, F.P., Chen, X.Q., Jiang, X.Y., Peng, Z.G., Zeng, D.M., Huang, D.S., 2012. Irradiation-mediated carbon nanotubes use in cancer therapy. *JOURNAL OF CANCER RESEARCH AND THERAPEUTICS* 8, 348–354. <https://doi.org/10.4103/0973-1482.103511>
- Yang, P., Xu, Q.Z., Jin, S.Y., Lu, Y., Zhao, Y., Yu, S.H., 2012. Synthesis of Multifunctional Ag@Au@Phenol Formaldehyde Resin Particles Loaded with Folic Acids for Photothermal Therapy. *CHEMISTRY-A EUROPEAN JOURNAL* 18, 9294–9299. <https://doi.org/10.1002/chem.201201173>
- Jin, L.L., Yang, K., Yao, K., Zhang, S., Tao, H.Q., Lee, S.T., Liu, Z., Peng, R., 2012. Functionalized Graphene Oxide in Enzyme Engineering: A Selective Modulator for Enzyme Activity and Thermostability. *ACS NANO* 6, 4864–4875. <https://doi.org/10.1021/nn300217z>
- Zanni, E., De Bellis, G., Bracciale, M.P., Broggi, A., Santarelli, M.L., Sarto, M.S., Palleschi, C., Uccelletti, D., 2012. Graphite Nanoplatelets and *Caenorhabditis elegans*: Insights from an in Vivo Model. *NANO LETTERS* 12, 2740–2744. <https://doi.org/10.1021/nl204388p>
- Yang, K., Hu, L.L., Ma, X.X., Ye, S.Q., Cheng, L., Shi, X.Z., Li, C.H., Li, Y.G., Liu, Z., 2012. Multimodal Imaging Guided Photothermal Therapy using Functionalized Graphene Nanosheets Anchored with Magnetic Nanoparticles. *ADVANCED MATERIALS* 24, 1868–1872. <https://doi.org/10.1002/adma.201104964>
- Li, M., Yang, X.J., Ren, J.S., Qu, K.G., Qu, X.G., 2012. Using Graphene Oxide High Near-Infrared Absorbance for Photothermal Treatment of Alzheimer's Disease. *ADVANCED MATERIALS* 24, 1722–1728. <https://doi.org/10.1002/adma.201104864>
- Li, Q.N., Guo, D.D., Zhang, R.Y., Wang, X.M., 2012. Increasing Anticancer Drug Internalization Induced by New Au-MWCNTs Nanocomposite. *JOURNAL OF NANOSCIENCE AND NANOTECHNOLOGY* 12, 2192–2198. <https://doi.org/10.1166/jnn.2012.5733>

- Yang, K., Wan, J.M., Zhang, S., Tian, B., Zhang, Y.J., Liu, Z., 2012. The influence of surface chemistry and size of nanoscale graphene oxide on photothermal therapy of cancer using ultra-low laser power. *BIOMATERIALS* 33, 2206–2214. <https://doi.org/10.1016/j.biomaterials.2011.11.064>
- Gollavelli, G., Ling, Y.C., 2012. Multi-functional graphene as an in vitro and in vivo imaging probe. *BIOMATERIALS* 33, 2532–2545. <https://doi.org/10.1016/j.biomaterials.2011.12.010>
- Hu, Z., Huang, Y.D., Sun, S.F., Guan, W.C., Yao, Y.H., Tang, P.Y., Li, C.Y., 2012. Visible light driven photodynamic anticancer activity of graphene oxide/TiO₂ hybrid. *CARBON* 50, 994–1004. <https://doi.org/10.1016/j.carbon.2011.10.002>
- Ma, X.X., Tao, H.Q., Yang, K., Feng, L.Z., Cheng, L., Shi, X.Z., Li, Y.G., Guo, L., Liu, Z., 2012. A functionalized graphene oxide-iron oxide nanocomposite for magnetically targeted drug delivery, photothermal therapy, and magnetic resonance imaging. *NANO RESEARCH* 5, 199–212. <https://doi.org/10.1007/s12274-012-0200-y>
- Li, N., Cao, M.H., Hu, C.W., 2012. Review on the latest design of graphene-based inorganic materials. *NANOSCALE* 4, 6205–6218. <https://doi.org/10.1039/c2nr31750h>
- Choudhury, D., Jaque, D., Rodenas, A., Ramsay, W.T., Paterson, L., Kar, A.K., 2012. Quantum dot enabled thermal imaging of optofluidic devices. *LAB ON A CHIP* 12, 2414–2420. <https://doi.org/10.1039/c2lc40181a>
- Franchi, L.P., Santos, R.A., Matsubara, E.Y., de Lima, J.C., Rosolen, J.M., Takahashi, C.S., 2012. CYTOTOXICITY AND GENOTOXICITY OF CARBON NANOTUBES. *QUIMICA NOVA* 35, 571–580. <https://doi.org/10.1590/S0100-40422012000300025>
- Chen, G.Y., Pang, D.W.P., Hwang, S.M., Tuan, H.Y., Hu, Y.C., 2012. A graphene-based platform for induced pluripotent stem cells culture and differentiation. *BIOMATERIALS* 33, 418–427. <https://doi.org/10.1016/j.biomaterials.2011.09.071>
- Akhavan, O., Ghaderi, E., Aghayee, S., Fereydooni, Y., Talebi, A., 2012. The use of a glucose-reduced graphene oxide suspension for photothermal cancer therapy. *JOURNAL OF MATERIALS CHEMISTRY* 22, 13773–13781. <https://doi.org/10.1039/c2jm31396k>
- Yao, J., Sun, Y., Yang, M., Duan, Y.X., 2012. Chemistry, physics and biology of graphene-based nanomaterials: new horizons for sensing, imaging and medicine. *JOURNAL OF MATERIALS CHEMISTRY* 22, 14313–14329. <https://doi.org/10.1039/c2jm31632c>
- Hou, C.Y., Duan, Y.R., Zhang, Q.H., Wang, H.Z., Li, Y.G., 2012. Bio-applicable and electroactive near-infrared laser-triggered self-healing hydrogels based on graphene networks. *JOURNAL OF MATERIALS CHEMISTRY* 22, 14991–14996. <https://doi.org/10.1039/c2jm32255b>
- Akhavan, O., Ghaderi, E., Emamy, H., 2012. Nontoxic concentrations of PEGylated graphene nanoribbons for selective cancer cell imaging and photothermal therapy. *JOURNAL OF MATERIALS CHEMISTRY* 22, 20626–20633. <https://doi.org/10.1039/c2jm34330d>
- Goncalves, G., Cruz, S.M.A., Ramalho, A., Gracio, J., Marques, P., 2012. Graphene oxide versus functionalized carbon nanotubes as a reinforcing agent in a PMMA/HA bone cement. *NANOSCALE* 4, 2937–2945. <https://doi.org/10.1039/c2nr30303e>
- Wawrzynczyk, D., Bednarkiewicz, A., Nyk, M., Strek, W., Samoc, M., 2012. Neodymium(III) doped fluoride nanoparticles as non-contact optical temperature sensors. *NANOSCALE* 4, 6959–6961. <https://doi.org/10.1039/c2nr32203j>
- Shen, H., Zhang, L.M., Liu, M., Zhang, Z.J., 2012. Biomedical Applications of Graphene. *THERANOSTICS* 2, 283–294. <https://doi.org/10.7150/thno.3642>

Kim, H., Namgung, R., Singha, K., Oh, I.K., Kim, W.J., 2011. Graphene Oxide-Polyethylenimine Nanoconstruct as a Gene Delivery Vector and Bioimaging Tool. *BIOCONJUGATE CHEMISTRY* 22, 2558–2567. <https://doi.org/10.1021/bc200397j>

Zhang, W., Guo, Z.Y., Huang, D.Q., Liu, Z.M., Guo, X., Zhong, H.Q., 2011. Synergistic effect of chemophotothermal therapy using PEGylated graphene oxide. *BIOMATERIALS* 32, 8555–8561. <https://doi.org/10.1016/j.biomaterials.2011.07.071>

Tian, B., Wang, C., Zhang, S., Feng, L.Z., Liu, Z., 2011. Photothermally Enhanced Photodynamic Therapy Delivered by Nano-Graphene Oxide. *ACS NANO* 5, 7000–7009. <https://doi.org/10.1021/nn201560b>

Vallet-Regí, M., Ruiz-Hernandez, E., Gonzalez, B., Baeza, A., 2011. Design of Smart Nanomaterials for Drug and Gene Delivery. *JOURNAL OF BIOMATERIALS AND TISSUE ENGINEERING* 1, 6–29. <https://doi.org/10.1166/jbt.2011.1006>

Feng, L.Z., Liu, Z.A., 2011. Graphene in biomedicine: opportunities and challenges. *NANOMEDICINE* 6, 317–324. <https://doi.org/10.2217/NNM.10.158>

Iancu, C., Mocan, L., 2011. Advances in cancer therapy through the use of carbon nanotube-mediated targeted hyperthermia. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 6, 1675–1684. <https://doi.org/10.2147/IJN.S23588>

Sasidharan, A., Panchakarla, L.S., Chandran, P., Menon, D., Nair, S., Rao, C.N.R., Koyakutty, M., 2011. Differential nano-bio interactions and toxicity effects of pristine versus functionalized graphene. *NANOSCALE* 3, 2461–2464. <https://doi.org/10.1039/c1nr10172b>

Рад у врхунском међународном часопису (M21):

1. **Pantovic A, Bosnjak M, Arsikin K, Kostic M, Mandic M, Ristic B, Tosic J, Suzin-Zivkovic V, Grujicic D, Isakovic A, Trajkovic V, Harhaji-Trajkovic Lj.** In vitro antiglioma action of indomethacin is mediated through AMPK/mTOR complex 1 signaling pathway. *Int J Biochem Cell Biol.* 2017;83:84-96. IF 3.905 (8 бодова)

Хемороцитата 8:

Chang, C.Y., Li, J.R., Wu, C.C., Wang, J. Der, Yang, C.P., Chen, W.Y., Wang, W.Y., Chen, C.J., 2018. Indomethacin induced glioma apoptosis involving ceramide signals. *Experimental Cell Research* 365, 66–77. <https://doi.org/10.1016/j.yexcr.2018.02.019>

Mazurek, M., Litak, J., Kamieniak, P., Kulesza, B., Jonak, K., Baj, J., Grochowski, C., 2020. Metformin as Potential Therapy for High-Grade Glioma. *CANCERS* 12. <https://doi.org/10.3390/cancers12010210>

Chang, C.Y., Li, J.R., Wu, C.C., Wang, J.D., Liao, S.L., Chen, W.Y., Wang, W.Y., Chen, C.J., 2020. Endoplasmic Reticulum Stress Contributes to Indomethacin-Induced Glioma Apoptosis. *INTERNATIONAL JOURNAL OF MOLECULAR SCIENCES* 21. <https://doi.org/10.3390/ijms21020557>

Chen, H.T., Xu, C., Zhang, F., Liu, Y., Guo, Y., Yao, Q.H., 2020. The gut microbiota attenuates muscle wasting by regulating energy metabolism in chemotherapy-induced malnutrition rats. *CANCER CHEMOTHERAPY AND PHARMACOLOGY* 85, 1049–1062. <https://doi.org/10.1007/s00280-020-04060-w>

Flores, R., Reyes-Garcia, L.I., Rodriguez-Laguna, N., Gomez-Balderas, R., 2018. Stability constants of Cu(II)/indomethacin mononuclear complexes in solution. *THEORETICAL CHEMISTRY ACCOUNTS* 137. <https://doi.org/10.1007/s00214-018-2315-z>

Buzharevski, A., Paskas, S., Laube, M., Losnecke, P., Neumann, W., Murganic, B., Mijatovic, S., Maksimovic-Ivanic, D., Pietzsch, J., Hey-Hawkins, E., 2019. Carboranyl Analogues of Ketoprofen with Cytostatic Activity against Human Melanoma and Colon Cancer Cell Lines. ACS OMEGA 4, 8824–8833. <https://doi.org/10.1021/acsomega.9b00412>

Xiong, Z.S., Gong, S.F., Si, W., Jiang, T.P., Li, Q.L., Wang, T.J., Wang, W.J., Wu, R.Y., Jiang, K., 2019. Effect of metformin on cell proliferation, apoptosis, migration and invasion in A172 glioma cells and its mechanisms. MOLECULAR MEDICINE REPORTS 20, 887–894. <https://doi.org/10.3892/mmr.2019.10369>

Kawaguchi-Ihara, N., Zhao, Y., Nakamura, S., Suzuki, K., Zhang, Y., Tohda, S., Murohashi, I., 2019. Chloroquine Inhibits Self-Renewal of Blast Progenitors Synergistically With Phytochemicals or Nonsteroidal Anti-inflammatory Drugs in Hematological Malignant Cell Lines. ANTICANCER RESEARCH 39, 87–98. <https://doi.org/10.21873/anticancer.13083>

2. **Pantovic A, Krstic A, Janjetovic K, Kocic J, Harhaji-Trajkovic L, Bugarski D, Trajkovic V.** Coordinated time-dependent modulation of AMPK/Akt/mTOR signaling and autophagy controls osteogenic differentiation of human mesenchymal stem cells. Bone. 2013;52(1):524-31. IF 4.461 (8 бодова)

Xemepouqumama 159:

Lorzadeh, S., Kohan, L., Ghavami, S., Azarpira, N., 2021. Autophagy and the Wnt signaling pathway: A focus on Wnt/beta-catenin signaling. BIOCHIMICA ET BIOPHYSICA ACTA-MOLECULAR CELL RESEARCH 1868. <https://doi.org/10.1016/j.bbamcr.2020.118926>

Karadeniz, F., Oh, J.H., Lee, J.I., Kim, H., Seo, Y., Kong, C.S., 2020. 6-Acetyl-2,2-Dimethylchroman-4-One Isolated from Artemisia princeps Suppresses Adipogenic Differentiation of Human Bone Marrow-Derived Mesenchymal Stromal Cells via Activation of AMPK. JOURNAL OF MEDICINAL FOOD 23, 250–257. <https://doi.org/10.1089/jmf.2019.4653>

Wu, L.X., Xiang, S.Q., Hu, X.H., Mo, M., Zhao, C., Cai, Y., Tong, S.Y., Jiang, H.C., Chen, L.X., Wang, Z., Xiong, W., Ou, Z.Y., 2020. Prostate-specific antigen modulates the osteogenic differentiation of MSCs via the cadherin 11-Akt axis. CLINICAL AND TRANSLATIONAL MEDICINE 10, 363–373. <https://doi.org/10.1002/ctm2.27>

Zhang, Y., Zhang, Z.N., Li, N., Zhao, L.J., Xue, Y., Wu, H.J., Hou, J.M., 2020. Nbr1-regulated autophagy in Lactoferrin-induced osteoblastic differentiation. BIOSCIENCE BIOTECHNOLOGY AND BIOCHEMISTRY 84, 1191–1200. <https://doi.org/10.1080/09168451.2020.1737505>

Wang, T.T., Liu, X.N., He, C.Q., 2020. Glucocorticoid-induced autophagy and apoptosis in bone. APOPTOSIS 25, 157–168. <https://doi.org/10.1007/s10495-020-01599-0>

Lei, S., Liu, X.M., Liu, Y., Bi, J., Zhu, S., Chen, X., 2020. Lipopolysaccharide Downregulates the Osteo-/Odontogenic Differentiation of Stem Cells From Apical Papilla by Inducing Autophagy. JOURNAL OF ENDODONTICS 46, 502–508. <https://doi.org/10.1016/j.joen.2020.01.009>

di Giacomo, V., Cataldi, A., Sancilio, S., 2020. Biological Factors, Metals, and Biomaterials Regulating Osteogenesis through Autophagy. INTERNATIONAL JOURNAL OF MOLECULAR SCIENCES 21. <https://doi.org/10.3390/ijms21082789>

Park, Y.H., Lee, Y.S., Seo, Y.M., Seo, H., Park, J.S., Bae, H.S., Park, J.C., 2020. Midkine Promotes Odontoblast-like Differentiation and Tertiary Dentin Formation. JOURNAL OF DENTAL RESEARCH 99, 1082–1091. <https://doi.org/10.1177/0022034520925427>

- Tong, X.S., Ganta, R.R., Liu, Z.P., 2020. AMP-activated protein kinase (AMPK) regulates autophagy, inflammation and immunity and contributes to osteoclast differentiation and function. *BIOLOGY OF THE CELL* 112, 251–264. <https://doi.org/10.1111/boc.202000008>
- Yang, Y.H., Lin, Z.W., Cheng, J.W., Ding, S., Mao, W.W., Shi, S., Liang, B.R., Jiang, L.S., 2020. The roles of autophagy in osteogenic differentiation in rat ligamentum fibroblasts: Evidence and possible implications. *FASEB JOURNAL* 34, 8876–8886. <https://doi.org/10.1096/fj.201903216RR>
- Jin, L.Y., Gao, F.F., Zhang, L.L., Wang, C., Hu, L., Fan, Z.P., Xia, D.S., 2021. Pleiotropin enhances the osteo/dentinogenic differentiation potential of dental pulp stem cells. *CONNECTIVE TISSUE RESEARCH* 62, 495–507. <https://doi.org/10.1080/03008207.2020.1779238>
- Cheng, Q., Zeng, K.R., Kang, Q.Y., Qian, W.T., Zhang, W.Y., Gan, Q.L., Xia, W.W., 2020. The Antimicrobial Peptide LL-37 Promotes Migration and Odonto/Osteogenic Differentiation of Stem Cells from the Apical Papilla through the Akt/NF- κ B/ β -catenin Signaling Pathway. *JOURNAL OF ENDODONTICS* 46, 964–972. <https://doi.org/10.1016/j.joen.2020.03.013>
- Sylakowski, K., Bradshaw, A., Wells, A., 2020. Mesenchymal Stem Cell/Multipotent Stromal Cell Augmentation of Wound Healing Lessons from the Physiology of Matrix and Hypoxia Support. *AMERICAN JOURNAL OF PATHOLOGY* 190, 1370–1381. <https://doi.org/10.1016/j.ajpath.2020.03.017>
- Wang, C.L., Wang, S.Y., Liu, S.C., Cheng, Y.N., Geng, H.W., Yang, R., Feng, T.Q., Lu, G.H., Sun, X.T., Song, J., Hao, L.N., 2020. Synonymous Mutations of Porcine Igf1r Extracellular Domain Affect Differentiation and Mineralization in MC3T3-E1 Cells. *FRONTIERS IN CELL AND DEVELOPMENTAL BIOLOGY* 8. <https://doi.org/10.3389/fcell.2020.00623>
- Wang, X.C., Chu, X.Y., Cao, L., Zhao, J., Zhu, L., Rahman, S.U., Hu, Z., Zhang, Y.F., Feng, S.B., Li, Y., Wu, J.J., 2020. The role and regulatory mechanism of autophagy in hippocampal nerve cells of piglet damaged by deoxynivalenol. *TOXICOLOGY IN VITRO* 66. <https://doi.org/10.1016/j.tiv.2020.104837>
- Yan, J.F., Qin, W.P., Xiao, B.C., Wan, Q.Q., Tay, F.R., Niu, L.N., Jiao, K., 2020. Pathological calcification in osteoarthritis: an outcome or a disease initiator? *BIOLOGICAL REVIEWS* 95, 960–985. <https://doi.org/10.1111/brv.12595>
- Yuan, J.M., Dong, X.D., Yap, J.J., Hu, J.C., 2020. The MAPK and AMPK signalings: interplay and implication in targeted cancer therapy. *JOURNAL OF HEMATOLOGY & ONCOLOGY* 13. <https://doi.org/10.1186/s13045-020-00949-4>
- Chen, X.D., Tan, J.L., Feng, Y., Huang, L.J., Zhang, M., Cheng, B., 2020. Autophagy in fate determination of mesenchymal stem cells and bone remodeling. *WORLD JOURNAL OF STEM CELLS* 12, 776–786. <https://doi.org/10.4252/wjsc.v12.i8.776>
- Li, X., Xu, J.K., Dai, B.Y., Wang, X.L., Guo, Q.Y., Qin, L., 2020. Targeting autophagy in osteoporosis: From pathophysiology to potential therapy. *AGEING RESEARCH REVIEWS* 62. <https://doi.org/10.1016/j.arr.2020.101098>
- Liu, W.J., Luo, H.Y., Wang, R.L., Kang, Y.Y., Liao, W.T., Sun, Y.P., Chen, G.D., Shao, L.Q., 2020. Rapamycin-Induced Autophagy Promotes the Chondrogenic Differentiation of Synovium-Derived Mesenchymal Stem Cells in the Temporomandibular Joint in Response to IL-1 β . *BIOMED RESEARCH INTERNATIONAL* 2020. <https://doi.org/10.1155/2020/4035306>
- Zhao, X.D., Pathak, J.L., Huang, W.Y., Zhu, C.D., Li, Y.Y., Guan, H.B., Zeng, S.J., Ge, L.H., Shu, Y., 2020. Metformin enhances osteogenic differentiation of stem cells from human exfoliated deciduous teeth through AMPK pathway. *JOURNAL OF TISSUE ENGINEERING AND REGENERATIVE MEDICINE* 14, 1869–1879. <https://doi.org/10.1002/term.3142>

- Liu, Z.Z., Hong, C.G., Hu, W.B., Chen, M.L., Duan, R., Li, H.M., Yue, T., Cao, J., Wang, Z.X., Chen, C.Y., Hu, X.K., Wu, B., Liu, H.M., Tan, Y.J., Liu, J.H., Luo, Z.W., Zhang, Y., Rao, S.S., Luo, M.J., Yin, H., Wang, Y.Y., Xia, K., Xu, L., Tang, S.Y., Hu, R.G., Xie, H., 2021. Autophagy receptor OPTN (optineurin) regulates mesenchymal stem cell fate and bone-fat balance during aging by clearing FABP3. *AUTOPHAGY* 17, 2766–2782. <https://doi.org/10.1080/15548627.2020.1839286>
- Deng, J.Q., Zhong, L.J., Zhou, Z.H., Gu, C.W., Huang, X.Y., Shen, L.H., Cao, S.Z., Ren, Z.H., Zuo, Z.C., Deng, J.L., Yu, S.M., 2021. Autophagy: a promising therapeutic target for improving mesenchymal stem cell biological functions. *MOLECULAR AND CELLULAR BIOCHEMISTRY* 476, 1135–1149. <https://doi.org/10.1007/s11010-020-03978-2>
- Garcia-Bernal, D., Garcia-Arranz, M., Garcia-Guillen, A.I., Garcia-Hernandez, A.M., Blanquer, M., Garcia-Olmo, D., Sackstein, R., Moraleda, J.M., Zapata, A.G., 2020. Exofucosylation of Adipose Mesenchymal Stromal Cells Alters Their Secretome Profile. *FRONTIERS IN CELL AND DEVELOPMENTAL BIOLOGY* 8. <https://doi.org/10.3389/fcell.2020.584074>
- Yang, Y.H., Huang, Y.P., Li, W.R., 2021. Autophagy and its significance in periodontal disease. *JOURNAL OF PERIODONTAL RESEARCH* 56, 18–26. <https://doi.org/10.1111/jre.12810>
- Zhao, Y.J., Guo, W.Y., Gu, X.L., Chang, C., Wu, J.N., 2020. Repression of deoxynivalenol-triggered cytotoxicity and apoptosis by mannan/beta-glucans from yeast cell wall: Involvement of autophagy and PI3K-AKT-mTOR signaling pathway. *INTERNATIONAL JOURNAL OF BIOLOGICAL MACROMOLECULES* 164, 1413–1421. <https://doi.org/10.1016/j.ijbiomac.2020.07.217>
- Gonzalez-Gonzalez, A., Garcia-Sanchez, D., Dotta, M., Rodriguez-Rey, J.C., Perez-Campo, F.M., 2020. Mesenchymal stem cells secretome: The cornerstone of cell-free regenerative medicine. *WORLD JOURNAL OF STEM CELLS* 12, 1529–1552. <https://doi.org/10.4252/wjsc.v12.i12.1529>
- Kim, Y.J., Kim, W.J., Bae, S.W., Yang, S.M., Park, S.Y., Kim, S.M., Jung, J.Y., 2021. Mineral trioxide aggregate-induced AMPK activation stimulates odontoblastic differentiation of human dental pulp cells. *INTERNATIONAL ENDODONTIC JOURNAL* 54, 753–767. <https://doi.org/10.1111/iej.13460>
- Bi, D.C., Yao, L.J., Lin, Z.J., Chi, L.L., Li, H., Xu, H., Du, X.B., Liu, Q., Hu, Z.L., Lu, J., Xu, X., 2021. Unsaturated mannuronate oligosaccharide ameliorates beta-amyloid pathology through autophagy in Alzheimer's disease cell models. *CARBOHYDRATE POLYMERS* 251. <https://doi.org/10.1016/j.carbpol.2020.117124>
- Gao, Y., Zhang, T., 2021. The Application of Nanomaterials in Cell Autophagy. *CURRENT STEM CELL RESEARCH & THERAPY* 16, 23–35. <https://doi.org/10.2174/1574888X15666200502000807>
- Zhang, H.T., Zeng, Q.Z., Wu, B.M., Lu, J.L., Tong, K.L., Lin, J.B., Liu, Q.Y., Xu, L.P., Yang, J., Liu, X.H., Liu, W.T., Zhang, Y.F., Lian, Q.H., Liu, L.X., Gao, X.J., 2021. TRIM21-regulated Annexin A2 plasma membrane trafficking facilitates osteosarcoma cell differentiation through the TFEB-mediated autophagy. *CELL DEATH & DISEASE* 12. <https://doi.org/10.1038/s41419-020-03364-2>
- Kajiura, K., Umemura, N., Ohkoshi, E., Ohta, T., Kondoh, N., Kawano, S., 2021. Shikonin induces odontoblastic differentiation of dental pulp stem cells via AKT-mTOR signaling in the presence of CD44. *CONNECTIVE TISSUE RESEARCH* 62, 689–697. <https://doi.org/10.1080/03008207.2020.1865937>
- Pieles, O., Hartmann, M., Morscheck, C., 2021. AMP-activated protein kinase and the down-stream activated process of autophagy regulate the osteogenic differentiation of human dental follicle cells. *ARCHIVES OF ORAL BIOLOGY* 122. <https://doi.org/10.1016/j.archoralbio.2020.104951>
- Yan, W.H., Diao, S., Fan, Z.P., 2021. The role and mechanism of mitochondrial functions and energy metabolism in the function regulation of the mesenchymal stem cells. *STEM CELL RESEARCH & THERAPY* 12. <https://doi.org/10.1186/s13287-021-02194-z>

- Patel, D., Potter, M., Anaya, J.M., McGee-Lawrence, M.E., Hamrick, M.W., Hill, W.D., Isales, C.M., Fulzele, S., 2021. Kynurenine induces an age-related phenotype in bone marrow stromal cells. *MECHANISMS OF AGEING AND DEVELOPMENT* 195. <https://doi.org/10.1016/j.mad.2021.111464>
- Zhang, X.R., Cui, J.J., Cheng, L.M., Lin, K.L., 2021. Enhancement of osteoporotic bone regeneration by strontium-substituted 45S5 bioglass via time-dependent modulation of autophagy and the Akt/mTOR signaling pathway. *JOURNAL OF MATERIALS CHEMISTRY B* 9, 3489–3501. <https://doi.org/10.1039/d0tb02991b>
- Yang, S.Y., Fan, W.G., Li, Y.Y., Liu, Q., He, H.W., Huang, F., 2021. Autophagy in tooth: Physiology, disease and therapeutic implication. *CELL BIOCHEMISTRY AND FUNCTION* 39, 702–712. <https://doi.org/10.1002/cbf.3636>
- Lin, Y.J., Zhang, M., Zhou, L., Chen, X.X., Chen, J., Wu, D., 2021. Promoting effect of rapamycin on osteogenic differentiation of maxillary sinus membrane stem cells. *PEERJ* 9. <https://doi.org/10.7717/peerj.11513>
- Nie, D.B., Zhou, Y.Q., Wang, W., Zhang, J.Y., Wang, J.H.C., 2021. Mechanical Overloading Induced-Activation of mTOR Signaling in Tendon Stem/Progenitor Cells Contributes to Tendinopathy Development. *FRONTIERS IN CELL AND DEVELOPMENTAL BIOLOGY* 9. <https://doi.org/10.3389/fcell.2021.687856>
- Zhou, X., Xu, S.N., Yuan, S.T., Lei, X.J., Sun, X.Y., Xing, L., Li, H.J., He, C.X., Qin, W., Zhao, D., Li, P.Q., Moharomd, E., Xu, X.H., Cao, H.L., 2021. Multiple functions of autophagy in vascular calcification. *CELL AND BIOSCIENCE* 11. <https://doi.org/10.1186/s13578-021-00639-9>
- Sotthibundhu, A., Muangchan, P., Phonchai, R., Promjantuek, W., Chaicharoenaudomrung, N., Kunhorm, P., Noisa, P., 2021. Autophagy Promoted Neural Differentiation of Human Placenta-derived Mesenchymal Stem Cells. *IN VIVO* 35, 2609–2620. <https://doi.org/10.21873/invivo.12543>
- Othman, A., Winogradzki, M., Lee, L., Tandon, M., Blank, A., Pratap, J., 2021. Bone Metastatic Breast Cancer: Advances in Cell Signaling and Autophagy Related Mechanisms. *CANCERS* 13. <https://doi.org/10.3390/cancers13174310>
- Nimiritsky, P., Novoseletskaia, E., Eremichev, R., Alexandrushkina, N., Karagyaury, M., Vetrovoy, O., Basalova, N., Khrustaleva, A., Tyakht, A., Efimenko, A., Tkachuk, V., Makarevich, P., 2021. Self-Organization Provides Cell Fate Commitment in MSC Sheet Condensed Areas via ROCK-Dependent Mechanism. *BIOMEDICINES* 9. <https://doi.org/10.3390/biomedicines9091192>
- Xu, H.X., Xu, F., Zhao, J.J., Zhou, C.X., Liu, J.R., 2021. Platelet-Rich Plasma Induces Autophagy and Promotes Regeneration in Human Dental Pulp Cells. *FRONTIERS IN BIOENGINEERING AND BIOTECHNOLOGY* 9. <https://doi.org/10.3389/fbioe.2021.659742>
- Zhang, S., Zhou, H., Kong, N., Wang, Z.Z., Fu, H.M., Zhang, Y.H., Xiao, Y., Yang, W.R., Yan, F.H., 2021. L-cysteine-modified chiral gold nanoparticles promote periodontal tissue regeneration. *BIOACTIVE MATERIALS* 6, 3288–3299. <https://doi.org/10.1016/j.bioactmat.2021.02.035>
- Liu, X., Tao, J., Yao, Y.Y., Yang, P., Wang, J.H., Yu, M.L., Hou, J.H., Zhang, Y., Gui, L., 2021. Resveratrol induces proliferation in preosteoblast cell MC3T3-E1 via GATA-1 activating autophagy. *ACTA BIOCHIMICA ET BIOPHYSICA SINICA* 53, 1495–1504. <https://doi.org/10.1093/abbs/gmab135>
- Jadaun, P.K., Zhang, S., Koedam, M., Demmers, J., Chatterjee, S., van Leeuwen, J.P., van der Eerden, B.C., 2022. Inhibition of hypoxia-induced Mucin 1 alters the proteomic composition of human osteoblast-produced extracellular matrix, leading to reduced osteogenic and angiogenic potential. *JOURNAL OF CELLULAR PHYSIOLOGY* 237, 1440–1454. <https://doi.org/10.1002/jcp.30617>
- Park, S.Y., Cho, H.S., Chung, K.H., Lee, B.N., Kim, S.H., Kim, W.J., Jung, J.Y., 2022. Inactivation of PI3K/Akt promotes the odontoblastic differentiation and suppresses the stemness with autophagic flux in dental pulp cells. *JOURNAL OF DENTAL SCIENCES* 17, 145–154. <https://doi.org/10.1016/j.jds.2021.05.013>

- Cai, G.P., Liu, Y.L., Luo, L.P., Xiao, Y., Jiang, T.J., Yuan, J., Wang, M., 2022. Alkbh1-mediated DNA N6-methyladenine modification regulates bone marrow mesenchymal stem cell fate during skeletal aging. *CELL PROLIFERATION* 55. <https://doi.org/10.1111/cpr.13178>
- Xu, H.Y., Xia, M., Sun, L., Wang, H., Zhang, W.B., 2022. Osteocytes Enhance Osteogenesis by Autophagy-Mediated FGF23 Secretion Under Mechanical Tension. *FRONTIERS IN CELL AND DEVELOPMENTAL BIOLOGY* 9. <https://doi.org/10.3389/fcell.2021.782736>
- Park, K.H., Cho, E.H., Bae, W.J., Kim, H.S., Lim, H.C., Park, Y.D., Lee, M.O., Cho, E.S., Kim, E.C., 2017. Role of PIN1 on in vivo periodontal tissue and in-vitro cells. *JOURNAL OF PERIODONTAL RESEARCH* 52, 617–627. <https://doi.org/10.1111/jre.12430>
- Peng, Y.Q., Xiong, D., Lin, X., Cui, R.R., Xu, F., Zhong, J.Y., Zhu, T., Wu, F., Mao, M.Z., Liao, X.B., Yuan, L.Q., 2017. Oestrogen Inhibits Arterial Calcification by Promoting Autophagy. *SCIENTIFIC REPORTS* 7. <https://doi.org/10.1038/s41598-017-03801-x>
- Kaluderovic, M.R., Krajnovic, T., Maksimovic-Ivanic, D., Graf, H.L., Mijatovic, S., 2017. Ti-SLActive and TiZr-SLActive Dental Implant Surfaces Promote Fast Osteoblast Differentiation. *COATINGS* 7. <https://doi.org/10.3390/coatings7070102>
- Jo, Y.I., Kim, G., Jin, Y.M., Park, Y.J., Kim, H.S., Park, Y.S., 2017. Intracellular Remodeling and Accumulation of Aberrant Lysosomes in Differentiation of Tonsil-Derived Mesenchymal Stem Cells into Parathyroid-Like Cells. *TISSUE ENGINEERING AND REGENERATIVE MEDICINE* 14, 411–420. <https://doi.org/10.1007/s13770-017-0042-5>
- Kim, J., Lee, H.W., Rhee, D.K., Paton, J.C., Pyo, S., 2017. Pneumolysin-induced autophagy contributes to inhibition of osteoblast differentiation through downregulation of Sp1 in human osteosarcoma cells. *BIOCHIMICA ET BIOPHYSICA ACTA-GENERAL SUBJECTS* 1861, 2663–2673. <https://doi.org/10.1016/j.bbagen.2017.07.008>
- Lin, H., Ying, Y., Wang, Y.Y., Wang, G., Jiang, S.S., Huang, D., Luo, L.Y., Chen, Y.G., Gerstenfeld, L.C., Luo, Z.J., 2017. AMPK downregulates ALK2 via increasing the interaction between Smurf1 and Smad6, leading to inhibition of osteogenic differentiation. *BIOCHIMICA ET BIOPHYSICA ACTA-MOLECULAR CELL RESEARCH* 1864, 2369–2377. <https://doi.org/10.1016/j.bbamcr.2017.08.009>
- Tang, Y.H., Yue, Z.S., Li, G.S., Zeng, L.R., Xin, D.W., Hu, Z.Q., Xu, C.D., 2018. Effect of -ecdysterone on glucocorticoid-induced apoptosis and autophagy in osteoblasts. *MOLECULAR MEDICINE REPORTS* 17, 158–164. <https://doi.org/10.3892/mmr.2017.7840>
- Zhou, J.H., Gao, X.L., Huang, S.Y., Ma, L., Cui, Y.J., Wang, H.K., Qiu, J.Z., Wang, L.L., Dong, Q.J., Chen, Z.G., Wang, X.X., Zhang, D.S., 2018. Simvastatin Improves the Jaw Bone Microstructural Defect Induced by High Cholesterol Diet in Rats by Regulating Autophagic Flux. *BIOMED RESEARCH INTERNATIONAL* 2018. <https://doi.org/10.1155/2018/4147932>
- Lee, S., Le, N.H., Kang, D., 2018. Melatonin alleviates oxidative stress-inhibited osteogenesis of human bone marrow-derived mesenchymal stem cells through AMPK activation. *INTERNATIONAL JOURNAL OF MEDICAL SCIENCES* 15, 1083–1091. <https://doi.org/10.7150/ijms.26314>
- An, Y., Liu, W.J., Xue, P., Ma, Y., Zhang, L.Q., Zhu, B., Qi, M., Li, L.Y., Zhang, Y.J., Wang, Q.T., Jin, Y., 2018. Autophagy promotes MSC-mediated vascularization in cutaneous wound healing via regulation of VEGF secretion. *CELL DEATH & DISEASE* 9. <https://doi.org/10.1038/s41419-017-0082-8>
- Li, X., Deng, S.C., Liu, M.L., Jin, Y., Zhu, S., Deng, S.J., Chen, J.Y., He, C., Qin, Q., Wang, C.Y., Zhao, G., 2018. The responsively decreased PKM2 facilitates the survival of pancreatic cancer cells in hypoglycose. *CELL DEATH & DISEASE* 9. <https://doi.org/10.1038/s41419-017-0158-5>

- Chen, M.H., Li, X.J., Fan, R.F., Yang, J., Jin, X., Hamid, S., Xu, S.W., 2018. Cadmium induces BNIP3-dependent autophagy in chicken spleen by modulating miR-33-AMPK axis. *CHEMOSPHERE* 194, 396–402. <https://doi.org/10.1016/j.chemosphere.2017.12.026>
- Singh, L., Tyagi, S., Myers, D., Duque, G., 2018. Good, Bad, or Ugly: the Biological Roles of Bone Marrow Fat. *CURRENT OSTEOPOROSIS REPORTS* 16, 130–137. <https://doi.org/10.1007/s11914-018-0427-y>
- Wang, Q., Huang, Y.X., Qian, Z.Y., 2018. Nanostructured Surface Modification to Bone Implants for Bone Regeneration. *JOURNAL OF BIOMEDICAL NANOTECHNOLOGY* 14, 628–648. <https://doi.org/10.1166/jbn.2018.2516>
- Qiu, S.S., Wang, J., Huang, S.Q., Sun, S.Q., Zhang, Z., Bao, N., 2018. Overactive autophagy is a pathological mechanism underlying premature suture ossification in nonsyndromic craniosynostosis. *SCIENTIFIC REPORTS* 8. <https://doi.org/10.1038/s41598-018-24885-z>
- Shen, G.Y., Ren, H., Qiu, T., Zhang, Z.D., Zhao, W.H., Yu, X., Huang, J.J., Tang, J.J., Liang, D., Yao, Z.S., Yang, Z.D., Jiang, X.B., 2018. Mammalian target of rapamycin as a therapeutic target in osteoporosis. *JOURNAL OF CELLULAR PHYSIOLOGY* 233, 3929–3944. <https://doi.org/10.1002/jcp.26161>
- Li, M., Yan, J.K., Chen, X., Tam, W., Zhou, L., Liu, T., Pan, G.Q., Lin, J., Yang, H.L., Pei, M., He, F., 2018. Spontaneous up-regulation of SIRT1 during osteogenesis contributes to stem cells' resistance to oxidative stress. *JOURNAL OF CELLULAR BIOCHEMISTRY* 119, 4928–4944. <https://doi.org/10.1002/jcb.26730>
- Lu, W., Guo, L., Jiang, T.L., Ni, S.H., Zhao, Y.Y., 2018. The effect of Mg-2Zn-0.5Nd alloy on the mTOR signalling pathway in L6 cells. *MOLECULAR MEDICINE REPORTS* 18, 1885–1891. <https://doi.org/10.3892/mmr.2018.9103>
- Fu, C., Liu, L., Li, F., 2018. Acetate alters the process of lipid metabolism in rabbits. *ANIMAL* 12, 1895–1902. <https://doi.org/10.1017/S1751731117003275>
- Pei, D.D., Sun, J.L., Zhu, C.H., Tian, F.C., Jiao, K., Anderson, M.R., Yiu, C., Huang, C., Jin, C.X., Bergeron, B.E., Chen, J.H., Tay, F.R., Niu, L.N., 2018. Contribution of Mitophagy to Cell-Mediated Mineralization: Revisiting a 50-Year-Old Conundrum. *ADVANCED SCIENCE* 5. <https://doi.org/10.1002/advs.201800873>
- Yang, M., Wen, T., Chen, H.X., Deng, J.Y., Yang, C., Zhang, Z., 2018. Knockdown of insulin-like growth factor 1 exerts a protective effect on hypoxic injury of aged BM-MSCs: role of autophagy. *STEM CELL RESEARCH & THERAPY* 9. <https://doi.org/10.1186/s13287-018-1028-5>
- Castiglioni, S., Romeo, V., Locatelli, L., Cazzaniga, A., Maier, J.A.M., 2018. TRPM7 and MagT1 in the osteogenic differentiation of human mesenchymal stem cells in vitro. *SCIENTIFIC REPORTS* 8. <https://doi.org/10.1038/s41598-018-34324-8>
- Hassanpour, M., Rezabakhsh, A., Pezeshkian, M., Rahbarghazi, R., Nouri, M., 2018. Distinct role of autophagy on angiogenesis: highlights on the effect of autophagy in endothelial lineage and progenitor cells. *STEM CELL RESEARCH & THERAPY* 9. <https://doi.org/10.1186/s13287-018-1060-5>
- Tanaka, Y., Sonoda, S., Yamaza, H., Murata, S., Nishida, K., Hama, S., Kyumoto-Nakamura, Y., Uehara, N., Nonaka, K., Kukita, T., Yamaza, T., 2018. Suppression of AKT-mTOR signal pathway enhances osteogenic/dentinogenic capacity of stem cells from apical papilla. *STEM CELL RESEARCH & THERAPY* 9. <https://doi.org/10.1186/s13287-018-1077-9>
- Jiang, Y.Y., Zhou, Z.F., Zhu, Y.J., Chen, F.F., Lu, B.Q., Cao, W.T., Zhang, Y.G., Cai, Z.D., Chen, F., 2018. Enzymatic Reaction Generates Biomimic Nanominerals with Superior Bioactivity. *SMALL* 14. <https://doi.org/10.1002/smll.201804321>

- Smieszek, A., Tomaszewski, K.A., Kornicka, K., Marycz, K., 2018. Metformin Promotes Osteogenic Differentiation of Adipose-Derived Stromal Cells and Exerts Pro-Osteogenic Effect Stimulating Bone Regeneration. *JOURNAL OF CLINICAL MEDICINE* 7. <https://doi.org/10.3390/jcm7120482>
- Yasunaga, M., Kajiya, H., Toshimitsu, T., Nakashima, H., Tamaoki, S., Ishikawa, H., Maeda, H., Ohno, J., 2019. The Early Autophagic Pathway Contributes to Osteogenic Differentiation of Human Periodontal Ligament Stem Cells. *JOURNAL OF HARD TISSUE BIOLOGY* 28, 63–70. <https://doi.org/10.2485/jhtb.28.63>
- Wang, T.T., He, H.C., Liu, S.X., Jia, C.S., Fan, Z.Y., Zhong, C., Yu, J.D., Liu, H.H., He, C.Q., 2019. Autophagy: A Promising Target for Age-related Osteoporosis. *CURRENT DRUG TARGETS* 20, 354–365. <https://doi.org/10.2174/1389450119666180626120852>
- Feng, L., Xia, B., Tian, B.F., Lu, G.B., 2019. MiR-152 influences osteoporosis through regulation of osteoblast differentiation by targeting RICTOR. *PHARMACEUTICAL BIOLOGY* 57, 586–594. <https://doi.org/10.1080/13880209.2019.1657153>
- Morszeck, C., 2019. Cellular senescence in dental pulp stem cells. *ARCHIVES OF ORAL BIOLOGY* 99, 150–155. <https://doi.org/10.1016/j.archoralbio.2019.01.012>
- Wang, F.F., Li, Y., Liu, H.C., 2019. Preparation of icariin/TiO₂ nanotube composite coating on pure titanium surface and analysis of early drug release. *EUROPEAN REVIEW FOR MEDICAL AND PHARMACOLOGICAL SCIENCES* 23, 1864–1873.
- Chen, M.W., Hu, Y., Hou, Y.H., Li, M.H., Chen, M.H., Mu, C.Y., Tao, B.L., Zhu, W., Luo, Z., Cai, K.Y., 2019. Differentiation regulation of mesenchymal stem cells via autophagy induced by structurally-different silica based nanobiomaterials. *JOURNAL OF MATERIALS CHEMISTRY B* 7, 2657–2666. <https://doi.org/10.1039/c9tb00040b>
- Zhang, K., Liu, F.W., Jin, D., Guo, T., Hou, R., Zhang, J.R., Lu, B., Hou, Y., Zhao, X., Li, Y.P., 2019. Autophagy preserves the osteogenic ability of periodontal ligament stem cells under high glucose conditions in rats. *ARCHIVES OF ORAL BIOLOGY* 101, 172–179. <https://doi.org/10.1016/j.archoralbio.2019.03.020>
- Castiglioni, S., Romeo, V., Locatelli, L., Zocchi, M., Zecchini, S., Maier, J.A.M., 2019. The simultaneous downregulation of TRPM7 and MagT1 in human mesenchymal stem cells in vitro: Effects on growth and osteogenic differentiation. *BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS* 513, 159–165. <https://doi.org/10.1016/j.bbrc.2019.03.178>
- Nguyen, T.M.D., Klett, D., Filliatreau, L., Combarnous, Y., 2019. Inhibition by fluoxetine of LH-stimulated cyclic AMP synthesis in tumor Leydig cells partly involves AMPK activation. *PLOS ONE* 14. <https://doi.org/10.1371/journal.pone.0217519>
- Ceccariglia, S., Cargnoni, A., Silini, A.R., Parolini, O., 2020. Autophagy: a potential key contributor to the therapeutic action of mesenchymal stem cells. *AUTOPHAGY* 16, 28–37. <https://doi.org/10.1080/15548627.2019.1630223>
- Shi, L.G., Li, B.C., Zhang, B.N., Zhen, C.Y., Zhou, J.D., Tang, S.J., 2019. Mouse embryonic palatal mesenchymal cells maintain stemness through the PTEN-Akt-mTOR autophagic pathway. *STEM CELL RESEARCH & THERAPY* 10. <https://doi.org/10.1186/s13287-019-1340-8>
- Jaber, F.A., Khan, N.M., Ansari, M.Y., Al-Adlaan, A.A., Hussein, N.J., Safadi, F.F., 2019. Autophagy plays an essential role in bone homeostasis. *JOURNAL OF CELLULAR PHYSIOLOGY* 234, 12105–12115. <https://doi.org/10.1002/jcp.27071>
- Cheng, Y., Huang, L.H., Wang, Y.C., Huo, Q.Y., Shao, Y.H., Bao, H.J., Li, Z.Y., Liu, Y.D., Li, X., 2019. Strontium promotes osteogenic differentiation by activating autophagy via the AMPK/mTOR signaling pathway in MC3T3-E1 cells. *INTERNATIONAL JOURNAL OF MOLECULAR MEDICINE* 44, 652–660. <https://doi.org/10.3892/ijmm.2019.4216>

- Lu, W.Y., Xu, E.N., Li, J.J., Chen, Y., Pan, Y.H., Wu, B.L., 2019. Effects of vascular endothelial growth factor and insulin growth factor-1 on proliferation, migration, osteogenesis and vascularization of human carious dental pulp stem cells. *MOLECULAR MEDICINE REPORTS* 20, 3924–3932. <https://doi.org/10.3892/mmr.2019.10606>
- Yin, X., Zhou, C.C., Li, J.T., Liu, R.K., Shi, B., Yuan, Q., Zou, S.J., 2019. Autophagy in bone homeostasis and the onset of osteoporosis. *BONE RESEARCH* 7. <https://doi.org/10.1038/s41413-019-0058-7>
- Li, X.C., Hu, L., Ma, L.S., Chang, S.M., Wang, W.L., Feng, Y.Y., Xu, Y.P., Hu, J.C., Zhang, C.M., Wang, S.L., 2019. Severe periodontitis may influence cementum and dental pulp through inflammation, oxidative stress, and apoptosis. *JOURNAL OF PERIODONTOLOGY* 90, 1297–1306. <https://doi.org/10.1002/JPER.18-0604>
- Xi, G., D'Costa, S., Wai, C., Xia, S.K., Cox, Z.C., Clemmons, D.R., 2019. IGFBP-2 stimulates calcium/calmodulin-dependent protein kinase kinase 2 activation leading to AMP-activated protein kinase induction which is required for osteoblast differentiation. *JOURNAL OF CELLULAR PHYSIOLOGY* 234, 23232–23242. <https://doi.org/10.1002/jcp.28890>
- Li, Z., Liu, X.N., Zhu, Y., Du, Y.G., Liu, X.J., Lv, L.W., Zhang, X., Liu, Y.S., Zhang, P., Zhou, Y.S., 2019. Mitochondrial Phosphoenolpyruvate Carboxykinase Regulates Osteogenic Differentiation by Modulating AMPK/ULK1-Dependent Autophagy. *STEM CELLS* 37, 1542–1555. <https://doi.org/10.1002/stem.3091>
- Hegedus, C., Boros, G., Fidrus, E., Kis, G.N., Antal, M., Juhasz, T., Janka, E.A., Janko, L., Paragh, G., Emri, G., Bai, P., Remenyik, E., 2020. PARP1 Inhibition Augments UVB-Mediated Mitochondrial Changes-Implications for UV-Induced DNA Repair and Photocarcinogenesis. *CANCERS* 12. <https://doi.org/10.3390/cancers12010005>
- Chen, W.T., Zhang, F., Zhao, X.Q., Yu, B., Wang, B.W., 2020. Galectin-3 and TRIM16 coregulate osteogenic differentiation of human bone marrow-derived mesenchymal stem cells at least partly via enhancing autophagy. *BONE* 131. <https://doi.org/10.1016/j.bone.2019.115059>
- He, W., Zheng, Y.D., Feng, Q.L., Elkhooly, T.A., Liu, X.J., Yang, X., Wang, Y.S., Xie, Y.J., 2020. Silver nanoparticles stimulate osteogenesis of human mesenchymal stem cells through activation of autophagy. *NANOMEDICINE* 15, 337–353. <https://doi.org/10.2217/nmm-2019-0026>
- Chen, S.J., van Tok, M.N., Knaup, V.L., Kraal, L., Pots, D., Bartels, L., Gravallesse, E.M., Taurog, J.D., van de Sande, M., van Duivenvoorde, L.M., Baeten, D.L., 2020. mTOR Blockade by Rapamycin in Spondyloarthritis: Impact on Inflammation and New Bone Formation in vitro and in vivo. *FRONTIERS IN IMMUNOLOGY* 10. <https://doi.org/10.3389/fimmu.2019.02344>
- Wang, Y.Q., Wang, N.X., Luo, Y., Yu, C.Y., Xiao, J.H., 2020. Ganoderma A effectively induces osteogenic differentiation of human amniotic mesenchymal stem cells via cross-talk between Wnt/beta-catenin and BMP/SMAD signaling pathways. *BIOMEDICINE & PHARMACOTHERAPY* 123. <https://doi.org/10.1016/j.biopha.2019.109807>
- Gao, Y., You, X.R., Liu, Y.B., Gao, F., Zhang, Y., Yang, J.R., Yang, C., 2020. Induction of autophagy protects human dental pulp cells from lipopolysaccharide-induced pyroptotic cell death. *EXPERIMENTAL AND THERAPEUTIC MEDICINE* 19, 2202–2210. <https://doi.org/10.3892/etm.2020.8475>
- Zhan, J.K., Wang, Y.J., Wang, Y., Wang, S., Tan, P., Huang, W., Liu, Y.S., 2014. The Mammalian Target of Rapamycin Signalling Pathway Is Involved in Osteoblastic Differentiation of Vascular Smooth Muscle Cells. *CANADIAN JOURNAL OF CARDIOLOGY* 30, 568–575. <https://doi.org/10.1016/j.cjca.2013.11.005>
- Chen, M.L., Qiao, H.Z., Su, Z.G., Li, H.P., Ping, Q.N., Zong, L., 2014. Emerging therapeutic targets for osteoporosis treatment. *EXPERT OPINION ON THERAPEUTIC TARGETS* 18, 817–831. <https://doi.org/10.1517/14728222.2014.912632>
- McMillan, E.M., Quadrilatero, J., 2014. Autophagy is required and protects against apoptosis during myoblast differentiation. *BIOCHEMICAL JOURNAL* 462, 267–277. <https://doi.org/10.1042/BJ20140312>

- Jakhar, R., Paul, S., Chauhan, A.K., Kang, S.C., 2014. Morin hydrate augments phagocytosis mechanism and inhibits LPS induced autophagic signaling in murine macrophage. *INTERNATIONAL IMMUNOPHARMACOLOGY* 22, 356–365. <https://doi.org/10.1016/j.intimp.2014.07.020>
- Nollet, M., Santucci-Darmanin, S., Breuil, V., Al-Sahlane, R., Cros, C., Topi, M., Momier, D., Samson, M., Pagnotta, S., Caillateau, L., Battaglia, S., Farlay, D., Dacquin, R., Barois, N., Jurdic, P., Boivin, G., Heymann, D., Lafont, F., Lu, S.S., Dempster, D.W., Carle, G.F., Pierrefite-Carle, V., 2014. Autophagy in osteoblasts is involved in mineralization and bone homeostasis. *AUTOPHAGY* 10, 1965–1977. <https://doi.org/10.4161/auto.36182>
- Lee, S., Cho, H.Y., Bui, H.T.T., Kang, D., 2014. The osteogenic or adipogenic lineage commitment of human mesenchymal stem cells is determined by protein kinase C delta. *BMC CELL BIOLOGY* 15. <https://doi.org/10.1186/s12860-014-0042-4>
- Nuschke, A., Rodrigues, M., Stolz, D.B., Chu, C.T., Griffith, L., Wells, A., 2014. Human mesenchymal stem cells/multipotent stromal cells consume accumulated autophagosomes early in differentiation. *STEM CELL RESEARCH & THERAPY* 5. <https://doi.org/10.1186/scrt530>
- Pei, F., Lin, H., Liu, H., Li, L., Zhang, L., Chen, Z., 2015. Dual Role of Autophagy in Lipopolysaccharide-induced Preodontoblastic Cells. *JOURNAL OF DENTAL RESEARCH* 94, 175–182. <https://doi.org/10.1177/0022034514553815>
- Ali, J.L., Lagasse, B.J., Minuk, A.J., Love, A.J., Moraya, A.I., Lam, L., Arthur, G., Gibson, S.B., Morrison, L.C., Werbowetski-Ogilvie, T.E., Fu, Y.X., Nachtigal, M.W., 2015. Differential cellular responses induced by dorsomorphin and LDN-193189 in chemotherapy-sensitive and chemotherapy-resistant human epithelial ovarian cancer cells. *INTERNATIONAL JOURNAL OF CANCER* 136, E455–E469. <https://doi.org/10.1002/ijc.29220>
- Yang, J.W., Zhang, Y.F., Wan, C.Y., Sun, Z.Y., Nie, S., Jian, S.J., Zhang, L., Song, G.T., Chen, Z., 2015. Autophagy in SDF-1 alpha-mediated DPSC migration and pulp regeneration. *BIOMATERIALS* 44, 11–23. <https://doi.org/10.1016/j.biomaterials.2014.12.006>
- Wang, S., Xia, P.Y., Rehm, M., Fan, Z., 2015. Autophagy and cell reprogramming. *CELLULAR AND MOLECULAR LIFE SCIENCES* 72, 1699–1713. <https://doi.org/10.1007/s00018-014-1829-3>
- Bakker, A.D., Jaspers, R.T., 2015. IL-6 and IGF-1 Signaling Within and Between Muscle and Bone: How Important is the mTOR Pathway for Bone Metabolism? *CURRENT OSTEOPOROSIS REPORTS* 13, 131–139. <https://doi.org/10.1007/s11914-015-0264-1>
- Viale-Bouroncle, S., Klingelhoffer, C., Ettl, T., Morsezeck, C., 2015. The AKT signaling pathway sustains the osteogenic differentiation in human dental follicle cells. *MOLECULAR AND CELLULAR BIOCHEMISTRY* 406, 199–204. <https://doi.org/10.1007/s11010-015-2437-8>
- Bionaz, M., Monaco, E., Wheeler, M.B., 2015. Transcription Adaptation during In Vitro Adipogenesis and Osteogenesis of Porcine Mesenchymal Stem Cells: Dynamics of Pathways, Biological Processes, Up-Stream Regulators, and Gene Networks. *PLOS ONE* 10. <https://doi.org/10.1371/journal.pone.0137644>
- Kim, K.W., Moon, S.J., Park, M.J., Kim, B.M., Kim, E.K., Lee, S.H., Lee, E.J., Chung, B.H., Yang, C.W., Cho, M.L., 2015. Optimization of adipose tissue-derived mesenchymal stem cells by rapamycin in a murine model of acute graft-versus-host disease. *STEM CELL RESEARCH & THERAPY* 6. <https://doi.org/10.1186/s13287-015-0197-8>
- Pierrefite-Carle, V., Santucci-Darmanin, S., Breuil, V., Camuzard, O., Carle, G.F., 2015. Autophagy in bone: Self-eating to stay in balance. *AGEING RESEARCH REVIEWS* 24, 206–217. <https://doi.org/10.1016/j.arr.2015.08.004>

- Xi, G., Rosen, C.J., Clemmons, D.R., 2016. IGF-I and IGFBP-2 Stimulate AMPK Activation and Autophagy, Which Are Required for Osteoblast Differentiation. *ENDOCRINOLOGY* 157, 268–281. <https://doi.org/10.1210/en.2015-1690>
- Lim, H.C., Nam, O.H., Kim, M.J., El-Fiqi, A., Yun, H.M., Lee, Y.M., Jin, G.Z., Lee, H.H., Kim, H.W., Kim, E.C., 2016. Delivery of dexamethasone from bioactive nanofiber matrices stimulates odontogenesis of human dental pulp cells through integrin/BMP/mTOR signaling pathways. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 11, 2557–2567. <https://doi.org/10.2147/IJN.S97846>
- Yokomoto-Umakoshi, M., Kanazawa, I., Takeno, A., Tanaka, K., Notsu, M., Sugimoto, T., 2016. Activation of AMP-activated protein kinase decreases receptor activator of NF-kappa B ligand expression and increases sclerostin expression by inhibiting the mevalonate pathway in osteocytic MLO-Y4 cells. *BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS* 469, 791–796. <https://doi.org/10.1016/j.bbrc.2015.12.072>
- Li, B., Duan, P., Li, C.F., Jing, Y., Han, X.F., Yan, W.H., Xing, Y., 2016. Role of autophagy on bone marrow mesenchymal stem-cell proliferation and differentiation into neurons. *MOLECULAR MEDICINE REPORTS* 13, 1413–1419. <https://doi.org/10.3892/mmr.2015.4673>
- Dasgupta, B., Chhipa, R.R., 2016. Evolving Lessons on the Complex Role of AMPK in Normal Physiology and Cancer. *TRENDS IN PHARMACOLOGICAL SCIENCES* 37, 192–206. <https://doi.org/10.1016/j.tips.2015.11.007>
- Khallaghi, B., Safarian, F., Nasoohi, S., Ahmadiani, A., Dargahi, L., 2016. Metformin-induced protection against oxidative stress is associated with AKT/mTOR restoration in PC12 cells. *LIFE SCIENCES* 148, 286–292. <https://doi.org/10.1016/j.lfs.2016.02.024>
- Pei, F., Wang, H.S., Chen, Z., Zhang, L., 2016. Autophagy regulates odontoblast differentiation by suppressing NF-kappa B activation in an inflammatory environment. *CELL DEATH & DISEASE* 7. <https://doi.org/10.1038/cddis.2015.397>
- Hsu, Y.C., Wu, Y.T., Yu, T.H., Wei, Y.H., 2016. Mitochondria in mesenchymal stem cell biology and cell therapy: From cellular differentiation to mitochondrial transfer. *SEMINARS IN CELL & DEVELOPMENTAL BIOLOGY* 52, 119–131. <https://doi.org/10.1016/j.semedb.2016.02.011>
- Zhou, B., Li, H., Liu, J., Xu, L., Guo, Q., Zang, W., Sun, H., Wu, S., 2016. Autophagic dysfunction is improved by intermittent administration of osteocalcin in obese mice. *INTERNATIONAL JOURNAL OF OBESITY* 40, 833–843. <https://doi.org/10.1038/ijo.2016.1>
- Li, Y., Guo, T., Zhang, Z., Yao, Y., Chang, S., Nor, J.E., Clarkson, B.H., Ni, L., Liu, J., 2016. Autophagy Modulates Cell Mineralization on Fluorapatite-Modified Scaffolds. *JOURNAL OF DENTAL RESEARCH* 95, 650–656. <https://doi.org/10.1177/0022034516636852>
- Lim, E.G., Kim, G.T., Lee, S.H., Kim, S.Y., Kim, Y.M., 2016. Apoptotic effects of extract from *Cnidium monnieri* (L.) Cusson by adenosine monophosphate-activated protein kinase-independent pathway in HCT116 colon cancer cells. *MOLECULAR MEDICINE REPORTS* 13, 4681–4688. <https://doi.org/10.3892/mmr.2016.5115>
- Prasadam, I., Batra, J., Perry, S., Gu, W.Y., Crawford, R., Xiao, Y., 2016. Systematic Identification, Characterization and Target Gene Analysis of microRNAs Involved in Osteoarthritis Subchondral Bone Pathogenesis. *CALCIFIED TISSUE INTERNATIONAL* 99, 43–55. <https://doi.org/10.1007/s00223-016-0125-7>
- Song, W., Shi, M.Q., Chang, B., Dong, M.D., Zhang, Y.M., 2016. The influence of nanotopography on organelle organization and communication. *NANO RESEARCH* 9, 2433–2444. <https://doi.org/10.1007/s12274-016-1129-3>

- Saldivia, M., Ceballos-Perez, G., Bart, J.M., Navarro, M., 2016. The AMPK alpha 1 Pathway Positively Regulates the Developmental Transition from Proliferation to Quiescence in *Trypanosoma brucei*. *CELL REPORTS* 17, 660–670. <https://doi.org/10.1016/j.celrep.2016.09.041>
- Brenner, A.K., Tvedt, T.H.A., Bruserud, O., 2016. The Complexity of Targeting PI3K-Akt-mTOR Signalling in Human Acute Myeloid Leukaemia: The Importance of Leukemic Cell Heterogeneity, Neighbouring Mesenchymal Stem Cells and Immunocompetent Cells. *MOLECULES* 21. <https://doi.org/10.3390/molecules21111512>
- Park, M.J., Lee, S.H., Moon, S.J., Lee, J.A., Lee, E.J., Kim, E.K., Park, J.S., Lee, J., Min, J.K., Kim, S.J., Park, S.H., Cho, M.L., 2016. Overexpression of soluble RAGE in mesenchymal stem cells enhances their immunoregulatory potential for cellular therapy in autoimmune arthritis. *SCIENTIFIC REPORTS* 6. <https://doi.org/10.1038/srep35933>
- Sbrana, F. V, Cortini, M., Avnet, S., Perut, F., Columbaro, M., De Mito, A., Baldini, N., 2016. The Role of Autophagy in the Maintenance of Stemness and Differentiation of Mesenchymal Stem Cells. *STEM CELL REVIEWS AND REPORTS* 12, 621–633. <https://doi.org/10.1007/s12015-016-9690-4>
- Jin, J., Gu, H., Anders, N.M., Ren, T.H., Jiang, M.L., Tao, M., Peng, Q., Rudek, M.A., Duan, W.Z., 2016. Metformin Protects Cells from Mutant Huntingtin Toxicity Through Activation of AMPK and Modulation of Mitochondrial Dynamics. *NEUROMOLECULAR MEDICINE* 18, 581–592. <https://doi.org/10.1007/s12017-016-8412-z>
- Bae, W.J., Auh, Q.S., Kim, G.T., Moon, J.H., Kim, E.C., 2016. Effects of sodium tri- and hexameta-phosphate in vitro osteoblastic differentiation in Periodontal Ligament and Osteoblasts, and in vivo bone regeneration. *DIFFERENTIATION* 92, 257–269. <https://doi.org/10.1016/j.diff.2016.04.004>
- Nuschke, A., Rodrigues, M., Wells, A.W., Sylakowski, K., Wells, A., 2016. Mesenchymal stem cells/multipotent stromal cells (MSCs) are glycolytic and thus glucose is a limiting factor of in vitro models of MSC starvation. *STEM CELL RESEARCH & THERAPY* 7. <https://doi.org/10.1186/s13287-016-0436-7>
- Song, W., Shi, M.Q., Dong, M.D., Zhang, Y.M., 2016. Inducing Temporal and Reversible Autophagy by Nanotopography for Potential Control of Cell Differentiation. *ACS APPLIED MATERIALS & INTERFACES* 8, 33475–33483. <https://doi.org/10.1021/acsami.6b11699>
- Sylvester, F.A., 2017. Inflammatory Bowel Disease: Effects on Bone and Mechanisms, in: McCabe, L.R., Parameswaran, N. (Eds.), *UNDERSTANDING THE GUT-BONE SIGNALING AXIS: MECHANISMS AND THERAPEUTIC IMPLICATIONS*. pp. 133–150. https://doi.org/10.1007/978-3-319-66653-2_7
- Arianna, C., Eliana, C., Flavio, A., Marco, R., Giacomo, D., Manuel, S., Elena, B., Alessandra, G., 2017. Rapid Rapamycin-Only Induced Osteogenic Differentiation of Blood-Derived Stem Cells and Their Adhesion to Natural and Artificial Scaffolds. *STEM CELLS INTERNATIONAL* 2017. <https://doi.org/10.1155/2017/2976541>
- Tan, Y.Q., Zhang, J., Zhou, G., 2017. Autophagy and its implication in human oral diseases. *AUTOPHAGY* 13, 225–236. <https://doi.org/10.1080/15548627.2016.1234563>
- Zhang, Y.H., Kong, N., Zhang, Y.C., Yang, W.R., Yan, F.H., 2017. Size-dependent Effects of Gold Nanoparticles on Osteogenic Differentiation of Human Periodontal Ligament Progenitor Cells. *THERANOSTICS* 7, 1214–1224. <https://doi.org/10.7150/thno.17252>
- Chen, Q., Wang, Z.Y., Chen, L.Y., Hu, H.Y., 2017. Roles of High Mobility Group Box 1 in Cardiovascular Calcification. *CELLULAR PHYSIOLOGY AND BIOCHEMISTRY* 42, 427–440. <https://doi.org/10.1159/000477591>
- Ka, M., Smith, A.L., Kim, W.Y., 2017. MTOR controls genesis and autophagy of GABAergic interneurons during brain development. *AUTOPHAGY* 13, 1348–1363. <https://doi.org/10.1080/15548627.2017.1327927>

- Deng, X.Y., Wu, W., Liang, H., Huang, D.H., Jing, D.D., Zheng, D., Shao, Z.W., 2017. Icarin Prevents IL-1 beta-Induced Apoptosis in Human Nucleus Pulposus via the PI3K/AKT Pathway. EVIDENCE-BASED COMPLEMENTARY AND ALTERNATIVE MEDICINE 2017. <https://doi.org/10.1155/2017/2198323>
- Russo, M., Moccia, S., Bilotto, S., Spagnuolo, C., Durante, M., Lenucci, M.S., Mita, G., Volpe, M.G., Aquino, R.P., Russo, G.L., 2017. A Carotenoid Extract from a Southern Italian Cultivar of Pumpkin Triggers Nonprotective Autophagy in Malignant Cells. OXIDATIVE MEDICINE AND CELLULAR LONGEVITY 2017. <https://doi.org/10.1155/2017/7468538>
- Qi, M., Zhang, L.Q., Ma, Y., Shuai, Y., Li, L.Y., Luo, K.F., Liu, W.J., Jin, Y., 2017. Autophagy Maintains the Function of Bone Marrow Mesenchymal Stem Cells to Prevent Estrogen Deficiency-Induced Osteoporosis. THERANOSTICS 7, 4498–4516. <https://doi.org/10.7150/thno.17949>
- Chen, S.C., Brooks, R., Houskeeper, J., Bremner, S.K., Dunlop, J., Viollet, B., Logan, P.J., Salt, I.P., Ahmed, S.F., Yarwood, S.J., 2017. Metformin suppresses adipogenesis through both AMP-activated protein kinase (AMPK)-dependent and AMPK-independent mechanisms. MOLECULAR AND CELLULAR ENDOCRINOLOGY 440, 57–68. <https://doi.org/10.1016/j.mce.2016.11.011>
- Veldhuis-Vlug, A.G., Rosen, C.J., 2017. Mechanisms of marrow adiposity and its implications for skeletal health. METABOLISM-CLINICAL AND EXPERIMENTAL 67, 106–114. <https://doi.org/10.1016/j.metabol.2016.11.013>
- Husak, Z., Dworzak, M.N., 2017. Chronic stress induces CD99, suppresses autophagy, and affects spontaneous adipogenesis in human bone marrow stromal cells. STEM CELL RESEARCH & THERAPY 8. <https://doi.org/10.1186/s13287-017-0532-3>
- Yang, G.J., Liu, H.M., Hu, X.X., Chen, Z.T., Friis, T.E., Wang, J.L., Xiao, Y., Zhang, S.M., 2017. Bio-inspired hybrid nanoparticles promote vascularized bone regeneration in a morphology-dependent manner. NANOSCALE 9, 5794–5805. <https://doi.org/10.1039/c7nr00347a>
- Bartolome, A., Lopez-Herradon, A., Portal-Nunez, S., Garcia-Aguilar, A., Esbrit, P., Benito, M., Guillen, C., 2013. Autophagy impairment aggravates the inhibitory effects of high glucose on osteoblast viability and function. BIOCHEMICAL JOURNAL 455, 329–337. <https://doi.org/10.1042/BJ20130562>
- Bonini, M.G., Gantner, B.N., 2013. The Multifaceted Activities of AMPK in Tumor Progression-Why the “One Size Fits All” Definition Does Not Fit at All? IUBMB LIFE 65, 889–896. <https://doi.org/10.1002/iub.1213>
- Li, F., Song, N., Tombran-Tink, J., Niyibizi, C., 2013. Pigment Epithelium-Derived Factor Enhances Differentiation and Mineral Deposition of Human Mesenchymal Stem Cells. STEM CELLS 31, 2714–2723. <https://doi.org/10.1002/stem.1505>
- Wei, Q., Zhang, Y., Sun, L., Jia, X.Q., Huai, W.W., Yu, C.C., Wan, Z.K., Han, L.H., 2013. High dose of extracellular ATP switched autophagy to apoptosis in anchorage-dependent and anchorage-independent hepatoma cells. PURINERGIC SIGNALLING 9, 585–598. <https://doi.org/10.1007/s11302-013-9369-0>
- Liu, D.M., Cui, W., Liu, B., Hu, H.J., Liu, J., Xie, R.Q., Yang, X.H., Gu, G.Q., Zhang, J.D., Zheng, H.M., 2014. Atorvastatin Protects Vascular Smooth Muscle Cells From TGF-beta 1-Stimulated Calcification by Inducing Autophagy via Suppression of the beta-Catenin Pathway. CELLULAR PHYSIOLOGY AND BIOCHEMISTRY 33, 129–141. <https://doi.org/10.1159/000356656>
- Chen, H.Q., Liu, X.B., Chen, H., Cao, J., Zhang, L., Hu, X.Y., Wang, J.A., 2014. Role of SIRT1 and AMPK in mesenchymal stem cells differentiation. AGEING RESEARCH REVIEWS 13, 55–64. <https://doi.org/10.1016/j.arr.2013.12.002>
- Kaluderovic, M.R., Mojic, M., Schreckenbach, J.P., Maksimovic-Ivanic, D., Graf, H.L., Mijatovic, S., 2014. A Key Role of Autophagy in Osteoblast Differentiation on Titanium-Based Dental Implants. CELLS TISSUES ORGANS 200, 265–277. <https://doi.org/10.1159/000434625>

Maes, H., Martin, S., Verfaillie, T., Agostinis, P., 2014. Dynamic interplay between autophagic flux and Akt during melanoma progression in vitro. *EXPERIMENTAL DERMATOLOGY* 23, 101–106. <https://doi.org/10.1111/exd.12298>

Montezano, A.C., Touyz, R.M., 2014. Mammalian Target of Rapamycin: A Novel Pathway in Vascular Calcification. *CANADIAN JOURNAL OF CARDIOLOGY* 30, 482–484. <https://doi.org/10.1016/j.cjca.2014.03.001>

3. Wasik AM, Grabarek J, Pantovic A, Cieřlar-Pobuda A, Asgari HR, Bundgaard-Nielsen C, Rafat M, Dixon IM, Ghavami S, Los MJ. Reprogramming and carcinogenesis-parallels and distinctions. *Int Rev Cell Mol Biol.* 2014;308:167-203. Review. IF 4.973 (8 бодова)

Xemepouumama 17:

Li, C., Li, Y.F., Sui, L.X., Wang, J., Li, F., 2019. Phenyllactic acid promotes cell migration and invasion in cervical cancer via IKK/NF-kappa B-mediated MMP-9 activation. *CANCER CELL INTERNATIONAL* 19. <https://doi.org/10.1186/s12935-019-0965-0>

Liu, L.Y., Yan, Z.F., Wang, Y.Y., Meng, J.H., Chen, G., 2020. Suppression of autophagy facilitates hydrogen gas-mediated lung cancer cell apoptosis. *ONCOLOGY LETTERS* 20. <https://doi.org/10.3892/ol.2020.11973>

Long, J., Kim, H., Kim, D., Lee, J.B., Kim, D.H., 2017. A biomaterial approach to cell reprogramming and differentiation. *JOURNAL OF MATERIALS CHEMISTRY B* 5, 2375–2389. <https://doi.org/10.1039/c6tb03130g>

Zhang, B., Liu, L.T., 2021. Autophagy is a double-edged sword in the therapy of colorectal cancer. *ONCOLOGY LETTERS* 21. <https://doi.org/10.3892/ol.2021.12639>

Liu, J.J., Chen, X.S., Ward, T., Mao, Y., Bockhorn, J., Liu, X.F., Wang, G., Pegram, M., Shen, K.W., 2016. Niclosamide inhibits epithelial-mesenchymal transition and tumor growth in lapatinib-resistant human epidermal growth factor receptor 2-positive breast cancer. *INTERNATIONAL JOURNAL OF BIOCHEMISTRY & CELL BIOLOGY* 71, 12–23. <https://doi.org/10.1016/j.biocel.2015.11.014>

Ramakrishna, R.R., Abd Hamid, Z., Zaki, W., Huddin, A.B., Mathialagan, R., 2020. Stem cell imaging through convolutional neural networks: current issues and future directions in artificial intelligence technology. *PEERJ* 8. <https://doi.org/10.7717/peerj.10346>

Qiu, L., Wu, J., Pan, C., Tan, X., Lin, J., Liu, R., Chen, S., Geng, R., Huang, W., 2016. Downregulation of CDC27 inhibits the proliferation of colorectal cancer cells via the accumulation of p21(Cip1/Waf1). *CELL DEATH & DISEASE* 7. <https://doi.org/10.1038/cddis.2015.402>

Checa-Rodriguez, C., Cepeda-Garcia, C., Ramon, J., Lopez-Saavedra, A., Balestra, F.R., Dominguez-Sanchez, M.S., Gomez-Cabello, D., Huertas, P., 2020. Methylation of the central transcriptional regulator KLF4 by PRMT5 is required for DNA end resection and recombination. *DNA REPAIR* 94. <https://doi.org/10.1016/j.dnarep.2020.102902>

Gomes, K.M.S., Costa, I.C., dos Santos, J.F., Dourado, P.M.M., Forni, M.F., Ferreira, J.C.B., 2017. Induced pluripotent stem cells reprogramming: Epigenetics and applications in the regenerative medicine. *REVISTA DA ASSOCIACAO MEDICA BRASILEIRA* 63, 180–189. <https://doi.org/10.1590/1806-9282.63.02.180>

Wang, Q., Liu, Z.L., Du, K.P., Liang, M., Zhu, X.J., Yu, Z.J., Chen, R., Qin, L.Y., Li, Y., Zheng, Y.F., 2019. Babaodan inhibits cell growth by inducing autophagy through the PI3K/AKT/mTOR pathway and enhances

antitumor effects of cisplatin in NSCLC cells. AMERICAN JOURNAL OF TRANSLATIONAL RESEARCH 11, 5272–5283.

Ou, M.L., Zhao, M., Li, C.H., Tang, D.G., Xu, Y., Dai, W.E., Sui, W.G., Zhang, Y., Xiang, Z., Mo, C.N., Lin, H., Dai, Y., 2021. Single-cell sequencing reveals the potential oncogenic expression atlas of human iPSC-derived cardiomyocytes. BIOLOGY OPEN 10. <https://doi.org/10.1242/bio.053348>

Wu, R., Duan, L., Cui, F., Cao, J., Xiang, Y., Tang, Y.S., Zhou, L., 2015. S100A9 promotes human hepatocellular carcinoma cell growth and invasion through RAGE-mediated ERK1/2 and p38 MAPK pathways. EXPERIMENTAL CELL RESEARCH 334, 228–238. <https://doi.org/10.1016/j.yexcr.2015.04.008>

Bing, W.D., Pang, X.Y., Qu, Q.X., Bai, X., Yang, W.W., Bi, Y.W., Bi, X.L., 2016. Simvastatin improves the homing of BMSCs via the PI3K/AKT/miR-9 pathway. JOURNAL OF CELLULAR AND MOLECULAR MEDICINE 20, 949–961. <https://doi.org/10.1111/jcmm.12795>

Vallee, A., Lecarpentier, Y., Vallee, J.N., 2021. The Key Role of the WNT/beta-Catenin Pathway in Metabolic Reprogramming in Cancers under Normoxic Conditions. CANCERS 13. <https://doi.org/10.3390/cancers13215557>

Kamada, R., Toguchi, Y., Nomura, T., Imagawa, T., Sakaguchi, K., 2016. Tetramer Formation of Tumor Suppressor Protein p53: Structure, Function, and Applications. BIOPOLYMERS 106, 598–612. <https://doi.org/10.1002/bip.22772>

Lee, J.H., Kwon, H.K., Shin, H.J., Nam, G.H., Kim, J.H., Choi, S., 2018. Quasi-Stem Cells Derived from Human Somatic Cells by Chemically Modified Carbon Nanotubes. ACS APPLIED MATERIALS & INTERFACES 10, 8417–8425. <https://doi.org/10.1021/acsami.7b12914>

Stella, G.M., Gentile, A., Baderacchi, A., Meloni, F., Milan, M., Benvenuti, S., 2016. Ockham's razor for the MET-driven invasive growth linking idiopathic pulmonary fibrosis and cancer. JOURNAL OF TRANSLATIONAL MEDICINE 14. <https://doi.org/10.1186/s12967-016-1008-4>

4. Harhaji-Trajkovic L, Arsikin K, Kravic-Stevovic T, Petricevic S, Tovilovic G, Pantovic A, Zogovic N, Ristic B, Janjetovic K, Bumbasirevic V, Trajkovic VS. Chloroquine-mediated lysosomal dysfunction enhances the anticancer effect of nutrient deprivation. Pharmaceutical Research. 2012;29(8):2249-63. IF 4.742 (8 bodova)

Xemepouumama 45:

Catalani, E., Giovarelli, M., Zecchini, S., Perrotta, C., Cervia, D., 2021. Oxidative Stress and Autophagy as Key Targets in Melanoma Cell Fate. CANCERS 13. <https://doi.org/10.3390/cancers13225791>

Mou, Q., Yang, Y.W., Chen, L., Fang, T., Yao, Y.C., Du, Z.Q., Yang, C.X., 2022. Melatonin mitigates Chloroquine-induced defects in porcine immature Sertoli cells. THERIOGENOLOGY 177, 1–10. <https://doi.org/10.1016/j.theriogenology.2021.10.005>

Choi, M.S., Chae, Y.J., Choi, J.W., Chang, J.E., 2021. Potential Therapeutic Approaches through Modulating the Autophagy Process for Skin Barrier Dysfunction. INTERNATIONAL JOURNAL OF MOLECULAR SCIENCES 22. <https://doi.org/10.3390/ijms22157869>

Gunay, B., Goncu, E., 2021. Role of autophagy in midgut stem cells of silkworm Bombyx mori, during larval-pupal metamorphosis. ARCHIVES OF INSECT BIOCHEMISTRY AND PHYSIOLOGY 108. <https://doi.org/10.1002/arch.21832>

- Wu, H.L., Li, W.B., Wang, T.T., Rong, Y., He, Z.Y., Huang, S.T., Zhang, L.F., Wu, Z.X., Liu, C.Z., 2021. alpha-Tomatine, a novel early-stage autophagy inhibitor, inhibits autophagy to enhance apoptosis via Beclin-1 in Skov3 cells. *FITOTERAPIA* 152. <https://doi.org/10.1016/j.fitote.2021.104911>
- Zhang, Q., Feng, Z., Lu, J., Lu, J.N., Guan, S., Chen, Y., 2021. Aflatoxin B1 inhibited autophagy flux by inducing lysosomal alkalization in HepG2 cells. *TOXICOLOGY MECHANISMS AND METHODS* 31, 450–456. <https://doi.org/10.1080/15376516.2021.1909196>
- Li, Y., Liu, F., Zhang, J.J., Liu, X.Y., Xiao, P.H., Bai, H.T., Chen, S., Wang, D., Sung, S.H.P., Kwok, R.T.K., Shen, J.Z., Zhu, K., Tang, B.Z., 2021. Efficient Killing of Multidrug-Resistant Internalized Bacteria by AIEgens In Vivo. *ADVANCED SCIENCE* 8. <https://doi.org/10.1002/advs.202001750>
- Wen, X., Klionsky, D.J., 2020. At a glance: A history of autophagy and cancer. *SEMINARS IN CANCER BIOLOGY* 66, 3–11. <https://doi.org/10.1016/j.semcancer.2019.11.005>
- Monti, E., Marras, E., Prini, P., Gariboldi, M.B., 2020. Luteolin impairs hypoxia adaptation and progression in human breast and colon cancer cells. *EUROPEAN JOURNAL OF PHARMACOLOGY* 881. <https://doi.org/10.1016/j.ejphar.2020.173210>
- Gray, A., Dang, B.N., Moore, T.B., Clemens, R., Pressman, P., 2020. A review of nutrition and dietary interventions in oncology. *SAGE OPEN MEDICINE* 8. <https://doi.org/10.1177/2050312120926877>
- Zheng, X.G., Jin, X.D., Liu, X.X., Liu, B.T., Li, P., Ye, F., Zhao, T., Chen, W.Q., Li, Q., 2020. Inhibition of endoplasmic reticulum stress-induced autophagy promotes the killing effect of X-rays on sarcoma in mice. *BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS* 522, 612–617. <https://doi.org/10.1016/j.bbrc.2019.11.160>
- Joshi, V., Upadhyay, A., Prajapati, V.K., Mishra, A., 2020. How autophagy can restore proteostasis defects in multiple diseases? *MEDICINAL RESEARCH REVIEWS* 40, 1385–1439. <https://doi.org/10.1002/med.21662>
- Miao, J.K., Liu, Y.H., Liu, S., Liu, X.M., Wang, P.C., Du, Z.Q., Yang, C.X., 2019. Lysosomal dysfunction disturbs porcine oocyte maturation and developmental capacity by disorganizing chromosome/cytoskeleton and activating autophagy/apoptosis. *THERIOGENOLOGY* 140, 44–51. <https://doi.org/10.1016/j.theriogenology.2019.08.019>
- Nunez-Olvera, S.I., Gallardo-Rincon, D., Puente-Rivera, J., Salinas-Vera, Y.M., Marchat, L.A., Morales-Villegas, R., Lopez-Camarillo, C., 2019. Autophagy Machinery as a Promising Therapeutic Target in Endometrial Cancer. *FRONTIERS IN ONCOLOGY* 9. <https://doi.org/10.3389/fonc.2019.01326>
- Khandia, R., Dadar, M., Munjal, A., Dhama, K., Karthik, K., Tiwari, R., Yattoo, M.I., Iqbal, H.M.N., Singh, K.P., Joshi, S.K., Chaicumpa, W., 2019. A Comprehensive Review of Autophagy and Its Various Roles in Infectious, Non-Infectious, and Lifestyle Diseases: Current Knowledge and Prospects for Disease Prevention, Novel Drug Design, and Therapy. *CELLS* 8. <https://doi.org/10.3390/cells8070674>
- Florczak, A., Mackiewicz, A., Dams-Kozłowska, H., 2019. Cellular uptake, intracellular distribution and degradation of Her2-targeting silk nanospheres. *INTERNATIONAL JOURNAL OF NANOMEDICINE* 14, 6855–6865. <https://doi.org/10.2147/IJN.S217854>
- Maheswari, U., Ghosh, K., Sadras, S.R., 2018. Licarin A induces cell death by activation of autophagy and apoptosis in non-small cell lung cancer cells. *APOPTOSIS* 23, 210–225. <https://doi.org/10.1007/s10495-018-1449-8>
- Chen, F.F., Song, Q.L., Yu, Q., 2018. Axl inhibitor R428 induces apoptosis of cancer cells by blocking lysosomal acidification and recycling independent of Axl inhibition. *AMERICAN JOURNAL OF CANCER RESEARCH* 8, 1466–+.

- Yin, C.J., Li, P.B., 2018. Growth suppression of glioma cells using HDAC6 inhibitor, tubacin. *OPEN MEDICINE* 13, 221–226. <https://doi.org/10.1515/med-2018-0034>
- Verbaanderd, C., Maes, H., Schaaf, M.B., Sukhatme, V.P., Pantziarka, P., Sukhatme, V., Agostinis, P., Bouche, G., 2017. Repurposing Drugs in Oncology (ReDO)-chloroquine and hydroxychloroquine as anti-cancer agents. *ECANCER MEDICAL SCIENCE* 11. <https://doi.org/10.3332/ecancer.2017.781>
- Schroll, M.M., LaBonia, G.J., Ludwig, K.R., Hummon, A.B., 2017. Glucose Restriction Combined with Autophagy Inhibition and Chemotherapy in HCT 116 Spheroids Decreases Cell Clonogenicity and Viability Regulated by Tumor Suppressor Genes. *JOURNAL OF PROTEOME RESEARCH* 16, 3009–3018. <https://doi.org/10.1021/acs.jproteome.7b00293>
- Ojha, C.R., Lapiere, J., Rodriguez, M., Dever, S.M., Zadeh, M.A., DeMarino, C., Pleet, M.L., Kashanchi, F., El-Hage, N., 2017. Interplay between Autophagy, Exosomes and HIV-1 Associated Neurological Disorders: New Insights for Diagnosis and Therapeutic Applications. *VIRUSES-BASEL* 9. <https://doi.org/10.3390/v9070176>
- Zheng, X.G., Jin, X.D., Li, F.F., Liu, X.X., Liu, Y., Ye, F., Li, P., Zhao, T., Li, Q., 2017. Inhibiting autophagy with chloroquine enhances the anti-tumor effect of high-LET carbon ions via ER stress-related apoptosis. *MEDICAL ONCOLOGY* 34. <https://doi.org/10.1007/s12032-017-0883-8>
- Huang, M., Garcia, J.S., Thomas, D., Zhu, L., Nguyen, L.X.T., Chan, S.M., Majeti, R., Medeiros, B.C., Mitchell, B.S., 2016. Autophagy mediates proteolysis of NPM1 and HEXIM1 and sensitivity to BET inhibition in AML cells. *ONCOTARGET* 7, 74917–74930. <https://doi.org/10.18632/oncotarget.12493>
- Weikel, K.A., Cacicedo, J.M., Ruderman, N.B., Ido, Y., 2016. Knockdown of GSK3 beta increases basal autophagy and AMPK signalling in nutrient-laden human aortic endothelial cells. *BIOSCIENCE REPORTS* 36. <https://doi.org/10.1042/BSR20160174>
- Liang, D.H., Choi, D.S., Ensor, J.E., Kaiparettu, B.A., Bass, B.L., Chang, J.C., 2016. The autophagy inhibitor chloroquine targets cancer stem cells in triple negative breast cancer by inducing mitochondrial damage and impairing DNA break repair. *CANCER LETTERS* 376, 249–258. <https://doi.org/10.1016/j.canlet.2016.04.002>
- Weikel, K.A., Ruderman, N.B., Cacicedo, J.M., 2016. Unraveling the actions of AMP-activated protein kinase in metabolic diseases: Systemic to molecular insights. *METABOLISM-CLINICAL AND EXPERIMENTAL* 65, 634–645. <https://doi.org/10.1016/j.metabol.2016.01.005>
- Pasquier, B., 2016. Autophagy inhibitors. *CELLULAR AND MOLECULAR LIFE SCIENCES* 73, 985–1001. <https://doi.org/10.1007/s00018-015-2104-y>
- Ban, G.Y., Pham, D.L., Trinh, T.H.K., Lee, S.I., Suh, D.H., Yang, E.M., Ye, Y.M., Shin, Y.S., Chwae, Y.J., Park, H.S., 2016. Autophagy mechanisms in sputum and peripheral blood cells of patients with severe asthma: a new therapeutic target. *CLINICAL AND EXPERIMENTAL ALLERGY* 46, 48–59. <https://doi.org/10.1111/cea.12585>
- Dong, X.Q., Wang, Y., Zhou, Y.C., Wen, J.F., Wang, S.L., Shen, L.Z., 2016. Aquaporin 3 facilitates chemoresistance in gastric cancer cells to cisplatin via autophagy. *CELL DEATH DISCOVERY* 2. <https://doi.org/10.1038/cddiscovery.2016.87>
- Pestana, C.R., Oishi, J.C., Salistre-Araujo, H.S., Rodrigues, G.J., 2015. Inhibition of Autophagy by Chloroquine Stimulates Nitric Oxide Production and Protects Endothelial Function during Serum Deprivation. *CELLULAR PHYSIOLOGY AND BIOCHEMISTRY* 37, 1168–1177. <https://doi.org/10.1159/000430240>
- Mei, L.F., Chen, Y.C., Wang, Z.M., Wang, J., Wan, J.L., Yu, C.R., Liu, X., Li, W.H., 2015. Synergistic anti-tumour effects of tetrandrine and chloroquine combination therapy in human cancer: a potential antagonistic role for p21. *BRITISH JOURNAL OF PHARMACOLOGY* 172, 2232–2245. <https://doi.org/10.1111/bph.13045>

- Li, X., Lao, Y.Z., Zhang, H., Wang, X.Y., Tan, H.S., Lin, Z.X., Xu, H.X., 2015. The natural compound Gutiferone F sensitizes prostate cancer to starvation induced apoptosis via calcium and JNK elevation. *BMC CANCER* 15. <https://doi.org/10.1186/s12885-015-1292-z>
- Junco, J.J., Mancha-Ramirez, A., Malik, G., Wei, S.J., Kim, D.J., Liang, H.Y., Slaga, T.J., 2015. Ursolic acid and resveratrol synergize with chloroquine to reduce melanoma cell viability. *MELANOMA RESEARCH* 25, 103–112. <https://doi.org/10.1097/CMR.0000000000000137>
- Johnson, C.E., Hunt, D.K., Wiltshire, M., Herbert, T.P., Sampson, J.R., Errington, R.J., Davies, D.M., Tee, A.R., 2015. Endoplasmic reticulum stress and cell death in mTORC1-overactive cells is induced by nelfinavir and enhanced by chloroquine. *MOLECULAR ONCOLOGY* 9, 675–688. <https://doi.org/10.1016/j.molonc.2014.11.005>
- Lao, Y.Z., Wan, G., Liu, Z.Y., Wang, X.Y., Ruan, P., Xu, W., Xu, D.Q., Xie, W.D., Zhang, Y., Xu, H.X., Xu, N.H., 2014. The natural compound oblongifolin C inhibits autophagic flux and enhances antitumor efficacy of nutrient deprivation. *AUTOPHAGY* 10, 736–749. <https://doi.org/10.4161/auto.28034>
- Todorovic, Z., Medic, B., Basta-Jovanovic, G., Skodric, S.R., Stojanovic, R., Rovcanin, B., Prostran, M., 2014. Acute Pretreatment with Chloroquine Attenuates Renal I/R Injury in Rats. *PLOS ONE* 9. <https://doi.org/10.1371/journal.pone.0092673>
- Vlahopoulos, S., Critselis, E., Voutsas, I.F., Perez, S.A., Moschovi, M., Baxevas, C.N., Chrousos, G.P., 2014. New Use for Old Drugs? Prospective Targets of Chloroquinines in Cancer Therapy. *CURRENT DRUG TARGETS* 15, 843–851.
- D'Amico, A.G., Scuderi, S., Saccone, S., Castorina, A., Drago, F., D'Agata, V., 2013. Antiproliferative Effects of PACAP and VIP in Serum-Starved Glioma Cells. *JOURNAL OF MOLECULAR NEUROSCIENCE* 51, 503–513. <https://doi.org/10.1007/s12031-013-0076-7>
- Liu, H., He, Z.Y., Simon, H.U., 2013. Targeting autophagy as a potential therapeutic approach for melanoma therapy. *SEMINARS IN CANCER BIOLOGY* 23, 352–360. <https://doi.org/10.1016/j.semcancer.2013.06.008>
- Rojas-Puentes, L.L., Gonzalez-Pinedo, M., Crismatt, A., Ortega-Gomez, A., Gamboa-Vignolle, C., Nunez-Gomez, R., Dorantes-Gallareta, Y., Arce-Salinas, C., Arrieta, O., 2013. Phase II randomized, double-blind, placebo-controlled study of whole-brain irradiation with concomitant chloroquine for brain metastases. *RADIATION ONCOLOGY* 8. <https://doi.org/10.1186/1748-717X-8-209>
- Egger, M.E., Huang, J.S., Yin, W.Y., McMasters, K.M., McNally, L.R., 2013. Inhibition of autophagy with chloroquine is effective in melanoma. *JOURNAL OF SURGICAL RESEARCH* 184, 274–281. <https://doi.org/10.1016/j.jss.2013.04.055>
- Yang, Y.P., Hu, L.F., Zheng, H.F., Mao, C.J., Hu, W.D., Xiong, K.P., Wang, F., Liu, C.F., 2013. Application and interpretation of current autophagy inhibitors and activators. *ACTA PHARMACOLOGICA SINICA* 34, 625–635. <https://doi.org/10.1038/aps.2013.5>
- Ning, Y.C., Cai, G.Y., Zhuo, L., Gao, J.J., Dong, D., Cui, S.Y., Shi, S.Z., Feng, Z., Zhang, L., Sun, X.F., Chen, X.M., 2013. Beneficial Effects of Short-Term Calorie Restriction against Cisplatin-Induced Acute Renal Injury in Aged Rats. *NEPHRON EXPERIMENTAL NEPHROLOGY* 124, 19–27. <https://doi.org/10.1159/000357380>
- Wang, C.K., Yu, X.D., Li, Q., Xie, G., Teng, Y., 2013. Chloroquine and Valproic Acid Combined Treatment in Vitro has Enhanced Cytotoxicity in an Osteosarcoma Cell Line. *ASIAN PACIFIC JOURNAL OF CANCER PREVENTION* 14, 4651–4654. <https://doi.org/10.7314/APJCP.2013.14.8.4651>

5. Pantic I, Harhaji-Trajkovic L, **Pantovic A**, Milosevic NT, Trajkovic V. Changes in fractal dimension and lacunarity as early markers of UV-induced apoptosis. *J Theor Biol.* 2012;303:87-92. IF 2.351 (8 бодова)

Хетероцитата 31:

Chirila, F. V, Khan, T.K., Alkon, D.L., 2013. Spatiotemporal Complexity of Fibroblast Networks Screens for Alzheimer's Disease. *JOURNAL OF ALZHEIMERS DISEASE* 33, 165–176. <https://doi.org/10.3233/JAD-2012-120745>

Badea, A.F., Platon, M.L., Crisan, M., Cattani, C., Badea, I., Pierro, G., Sannino, G., Baciut, G., 2013. Fractal Analysis of Elastographic Images for Automatic Detection of Diffuse Diseases of Salivary Glands: Preliminary Results. *COMPUTATIONAL AND MATHEMATICAL METHODS IN MEDICINE* 2013. <https://doi.org/10.1155/2013/347238>

Cattani, C., Pierro, G., 2013. On the Fractal Geometry of DNA by the Binary Image Analysis. *BULLETIN OF MATHEMATICAL BIOLOGY* 75, 1544–1570. <https://doi.org/10.1007/s11538-013-9859-9>

Pippa, N., Dokoumetzidis, A., Demetzos, C., Macheras, P., 2013. On the ubiquitous presence of fractals and fractal concepts in pharmaceutical sciences: A review. *INTERNATIONAL JOURNAL OF PHARMACEUTICS* 456, 340–352. <https://doi.org/10.1016/j.ijpharm.2013.08.087>

Roy, A., Perfect, E., 2014. LACUNARITY ANALYSES OF MULTIFRACTAL AND NATURAL GRAYSCALE PATTERNS. *FRACTALS-COMPLEX GEOMETRY PATTERNS AND SCALING IN NATURE AND SOCIETY* 22. <https://doi.org/10.1142/S0218348X14400039>

Lennon, F.E., Cianci, G.C., Cipriani, N.A., Hensing, T.A., Zhang, H.J., Chen, C.T., Murgu, S.D., Vokes, E.E., Vannier, M.W., Salgia, R., 2015. Lung cancer-a fractal viewpoint. *NATURE REVIEWS CLINICAL ONCOLOGY* 12, 664–675. <https://doi.org/10.1038/nrclinonc.2015.108>

Kimori, Y., Hikino, K., Nishimura, M., Mano, S., 2016. Quantifying morphological features of actin cytoskeletal filaments in plant cells based on mathematical morphology. *JOURNAL OF THEORETICAL BIOLOGY* 389, 123–131. <https://doi.org/10.1016/j.jtbi.2015.10.031>

Grbatinic, I., Milosevic, N.T., 2016. Incipient UV-Induced Structural Changes in Neutrophil Granulocytes: Morphometric and Texture Analysis of Two-Dimensional Digital Images. *MICROSCOPY AND MICROANALYSIS* 22, 387–393. <https://doi.org/10.1017/S1431927616000532>

Grbatinic, I., Milosevic, N., 2016. Classification of adult human dentate nucleus border neurons: Artificial neural networks and multidimensional approach. *JOURNAL OF THEORETICAL BIOLOGY* 404, 273–284. <https://doi.org/10.1016/j.jtbi.2016.06.011>

Eom, T., Woo, K., Cho, W., Heo, J.E., Jang, D., Shin, J.I., Martin, D.C., Wie, J.J., Shim, B.S., 2017. Nanoarchitecturing of Natural Melanin Nanospheres by Layer-by-Layer Assembly: Macroscale Anti-inflammatory Conductive Coatings with Optoelectronic Tunability. *BIOMACROMOLECULES* 18, 1908–1917. <https://doi.org/10.1021/acs.biomac.7b00336>

Caudron, Q., Garnier, R., Pilkington, J.G., Watt, K.A., Hansen, C., Grenfell, B.T., Aboellail, T., Graham, A.L., 2017. Robust extraction of quantitative structural information from high-variance histological images of livers from necropsied Soay sheep. *ROYAL SOCIETY OPEN SCIENCE* 4. <https://doi.org/10.1098/rsos.170111>

Pronin, S., Koh, C.H., Hughes, M., 2017. Effects of Ultraviolet Radiation on Glioma: Systematic Review. *JOURNAL OF CELLULAR BIOCHEMISTRY* 118, 4063–4071. <https://doi.org/10.1002/jcb.26061>

- Gonzalez-Gutierrez, J., Carreon, Y.J.P., Moctezuma, R.E., 2018. Structural evolution of a granular medium during simultaneous penetration. *PHYSICA A-STATISTICAL MECHANICS AND ITS APPLICATIONS* 489, 9–17. <https://doi.org/10.1016/j.physa.2017.07.014>
- Grbatinic, I., Milosevic, N., Maric, D., 2018. Translaminar neuromorphotopological clustering and classification of dentate nucleus neurons. *JOURNAL OF INTEGRATIVE NEUROSCIENCE* 17, 105–124. <https://doi.org/10.31083/JIN-170044>
- Liu, Y., Ren, J., ASME, 2018. MODELING AND CONTROL OF DYNAMIC CELLULAR MECHANOTRANSDUCTION (I): ACTIN CYTOSKELETON QUANTIFICATION. *PROCEEDINGS OF THE ASME 11TH ANNUAL DYNAMIC SYSTEMS AND CONTROL CONFERENCE*, 2018, VOL 1.
- Grbatinic, I., Milosevic, N., Krstonosic, B., 2018. The neuromorphological caudate-putaminal clustering of neostriate interneurons: Kohonen self-organizing maps and supervised artificial neural networks with multivariate analysis. *JOURNAL OF THEORETICAL BIOLOGY* 438, 96–115. <https://doi.org/10.1016/j.jtbi.2017.11.013>
- Velazquez, J.D.H., Mejia-Rosales, S., Goicochea, A.G., 2018. Fractal properties of biophysical models of pericellular brushes can be used to differentiate between cancerous and normal cervical epithelial cells. *COLLOIDS AND SURFACES B-BIOINTERFACES* 170, 572–577. <https://doi.org/10.1016/j.colsurfb.2018.06.059>
- Khalo, I. V, Konokhova, A.I., Orlova, D.Y., Trusov, K. V, Yurkin, M.A., Bartova, E., Kozubek, S., Maltsev, V.P., Chernyshev, A. V, 2018. Nuclear apoptotic volume decrease in individual cells: Confocal microscopy imaging and kinetic modeling. *JOURNAL OF THEORETICAL BIOLOGY* 454, 60–69. <https://doi.org/10.1016/j.jtbi.2018.05.034>
- Grbatinic, I., Krstonosic, B., Maric, D., Milosevic, N., 2018. Morphological Properties of the Two Types of Caudate Interneurons: Kohonen Self-Organizing Maps and Correlation-Comparison Analysis. *MICROSCOPY AND MICROANALYSIS* 24, 684–707. <https://doi.org/10.1017/S1431927618015337>
- Liu, Y., Mollaeian, K., Ren, J., 2018. An Image Recognition-Based Approach to Actin Cytoskeleton Quantification. *ELECTRONICS* 7. <https://doi.org/10.3390/electronics7120443>
- Metze, K., Adam, R., Florindo, J.B., 2019. The fractal dimension of chromatin - a potential molecular marker for carcinogenesis, tumor progression and prognosis. *EXPERT REVIEW OF MOLECULAR DIAGNOSTICS* 19, 299–312. <https://doi.org/10.1080/14737159.2019.1597707>
- Shin, J.I., Cho, S.J., Jeon, J., Lee, K.H., Wie, J.J., 2019. Three-dimensional micropatterning of semiconducting polymers via capillary force-assisted evaporative self-assembly. *SOFT MATTER* 15, 3854–3863. <https://doi.org/10.1039/c9sm00478e>
- Todorovic, J., Dincic, M., Ostojic, J.N., Zaletel, I., Lopacic, S., Dundjerovic, D., Tatic, S., Kovacevic, S., Paunovic, I., Puskas, N., Markovic, L., 2019. Differences in Chromatin Texture and Nuclear Fractal Dimension Between Hashimoto's and Lymphocytic Thyroiditis Lymphocytes. *MICROSCOPY AND MICROANALYSIS* 25, 762–768. <https://doi.org/10.1017/S1431927619000163>
- Dincic, M., Todorovic, J., Ostojic, J.N., Kovacevic, S., Dunderovic, D., Lopacic, S., Spasic, S., Radojevic-Skodric, S., Stanisavljevic, D., Ilic, A.Z., 2020. The Fractal and GLCM Textural Parameters of Chromatin May Be Potential Biomarkers of Papillary Thyroid Carcinoma in Hashimoto's Thyroiditis Specimens. *MICROSCOPY AND MICROANALYSIS* 26, 717–730. <https://doi.org/10.1017/S1431927620001683>
- de Mattos, A.C., Florindo, J.B., Adam, R.L., Lorand-Metze, I., Metze, K., n.d. The Fractal Dimension Suggests Two Chromatin Configurations in Small Cell Neuroendocrine Lung Cancer and Is an Independent Unfavorable Prognostic Factor for Overall Survival. *MICROSCOPY AND MICROANALYSIS*. <https://doi.org/10.1017/S1431927622000113>

6. Trpkovic A, Todorovic-Markovic B, Kleut D, Misirkic M, Janjetovic K, Vucicevic L, **Pantovic A**, Jovanovic S, Dramicanin M, Markovic Z, Trajkovic V. Oxidative stress-mediated hemolytic activity of solvent exchange-prepared fullerene (C60) nanoparticles. *Nanotechnology*. 2010;21(37):375102. IF 3.652 (8 бодова)

Хемороумана 21:

Mehrizi, T.Z., 2021. Hemocompatibility and Hemolytic Effects of Functionalized Nanoparticles on Red Blood Cells: A Recent Review Study. *NANO* 16. <https://doi.org/10.1142/S1793292021300073>

Gerasimova, L. V, Charykov, N.A., Semenov, K.N., Keskinov, V.A., Kurilenko, A. V, Shaimardanov, Z.K., Shaimardanova, B.K., Kulenova, N.A., Letenko, D.G., Kanbar, A., 2021. Catalytic fullerenol action on *Chlorella* growth in the conditions of limited resource base and in the conditions of oxidation stress. *NANOSYSTEMS-PHYSICS CHEMISTRY MATHEMATICS* 12, 346–362. <https://doi.org/10.17586/2220-8054-2021-12-3-346-362>

Drvenica, I., Mojsilovic, S., Stancic, A., Markovic, D., Kovacic, M., Maslovacic, I., Rapajic, I., Vucetic, D., Ilic, V., 2021. The effects of incubation media on the assessment of the shape of human erythrocytes by flow cytometry: a contribution to mathematical data interpretation to enable wider application of the method. *EUROPEAN BIOPHYSICS JOURNAL WITH BIOPHYSICS LETTERS* 50, 829–846. <https://doi.org/10.1007/s00249-021-01527-3>

Fedel, M., 2020. Hemocompatibility of Carbon Nanostructures. *C-JOURNAL OF CARBON RESEARCH* 6. <https://doi.org/10.3390/c6010012>

Chitra, S., Bargavi, P., Balakumar, S., 2020. Effect of microwave and probe sonication processes on sol-gel-derived bioactive glass and its structural and biocompatible investigations. *JOURNAL OF BIOMEDICAL MATERIALS RESEARCH PART B-APPLIED BIOMATERIALS* 108, 143–155. <https://doi.org/10.1002/jbm.b.34373>

Sumi, N., Chitra, K.C., 2019. Cytogenotoxic effects of fullerene C-60 in the freshwater teleostean fish, *Anabas testudineus* (Bloch, 1792). *MUTATION RESEARCH-GENETIC TOXICOLOGY AND ENVIRONMENTAL MUTAGENESIS* 847. <https://doi.org/10.1016/j.mrgentox.2019.503104>

Sumi, N., Chitra, K.C., 2019. Impact of fullerene C-60 on behavioral and hematological changes in the freshwater fish, *Anabas testudineus* (Bloch, 1792). *APPLIED NANOSCIENCE* 9, 2147–2167. <https://doi.org/10.1007/s13204-019-01041-1>

Wadhwa, R., Aggarwal, T., Thapliyal, N., Kumar, A., Priya, Yadav, P., Kumari, V., Reddy, B.S.C., Chandra, P., Maurya, P.K., 2019. Red blood cells as an efficient in vitro model for evaluating the efficacy of metallic nanoparticles. *3 BIOTECH* 9. <https://doi.org/10.1007/s13205-019-1807-4>

Sakhautdinov, I.M., Mukhamet'yanova, A.F., 2018. Cycloaddition of Bromo(chloro)methyl Ketones and Allenates of Adipic Acid Monoesters to Fullerene C-60. *RUSSIAN JOURNAL OF GENERAL CHEMISTRY* 88, 2528–2533. <https://doi.org/10.1134/S1070363218120125>

Goodarzi, S., Da Ros, T., Conde, J., Sefat, F., Mozafari, M., 2017. Fullerene: biomedical engineers get to revisit an old friend. *MATERIALS TODAY* 20, 460–480. <https://doi.org/10.1016/j.mattod.2017.03.017>

Hu, X.G., Sun, A.Q., Kang, W.L., Zhou, Q.X., 2017. Strategies and knowledge gaps for improving nanomaterial biocompatibility. *ENVIRONMENT INTERNATIONAL* 102, 177–189. <https://doi.org/10.1016/j.envint.2017.03.001>

- Clark, J., Gregory, C.C., Matthews, I.P., Hoogendoorn, B., 2016. The biological effects upon the cardiovascular system consequent to exposure to particulates of less than 500 nm in size. *BIOMARKERS* 21, 1–47. <https://doi.org/10.3109/1354750X.2015.1118540>
- Vinardell, M.P., Sorde, A., Diaz, J., Baccarin, T., Mitjans, M., 2015. Comparative effects of macro-sized aluminum oxide and aluminum oxide nanoparticles on erythrocyte hemolysis: influence of cell source, temperature, and size. *JOURNAL OF NANOPARTICLE RESEARCH* 17. <https://doi.org/10.1007/s11051-015-2893-9>
- Li, H.C., Hsieh, F.J., Chen, C.P., Chang, M.Y., Hsieh, P.C.H., Chen, C.C., Hung, S.U., Wu, C.C., Chang, H.C., 2013. The hemocompatibility of oxidized diamond nanocrystals for biomedical applications. *SCIENTIFIC REPORTS* 3. <https://doi.org/10.1038/srep03044>
- Grebowski, J., Krokosz, A., Puchala, M., 2013. Fullerenol C-60(OH)(36) could associate to band 3 protein of human erythrocyte membranes. *BIOCHIMICA ET BIOPHYSICA ACTA-BIOMEMBRANES* 1828, 2007–2014. <https://doi.org/10.1016/j.bbamem.2013.05.009>
- Mitra, S., Chandra, S., Pathan, S.H., Sikdar, N., Pramanik, P., Goswami, A., 2013. Room temperature and solvothermal green synthesis of self passivated carbon quantum dots. *RSC ADVANCES* 3, 3189–3193. <https://doi.org/10.1039/c2ra23085b>
- Snow, S.D., Lee, J., Kim, J.H., 2012. Photochemical and Photophysical Properties of Sequentially Functionalized Fullerenes in the Aqueous Phase. *ENVIRONMENTAL SCIENCE & TECHNOLOGY* 46, 13227–13234. <https://doi.org/10.1021/es303237v>
- Stamopoulos, D., Grapsa, E., Manios, E., Gogola, V., Bakirtzi, N., 2012. Defected red blood cell membranes and direct correlation with the uraemic milieu: the connection with the decreased red blood cell lifespan observed in haemodialysis patients. *NANOTECHNOLOGY* 23. <https://doi.org/10.1088/0957-4484/23/48/485101>
- Lin, J.J., Lin, W.C., Dong, R.X., Hsu, S.H., 2012. The cellular responses and antibacterial activities of silver nanoparticles stabilized by different polymers. *NANOTECHNOLOGY* 23. <https://doi.org/10.1088/0957-4484/23/6/065102>
- Fitzgerald, K.T., Holladay, C.A., McCarthy, C., Power, K.A., Pandit, A., Gallagher, W.M., 2011. Standardization of Models and Methods Used to Assess Nanoparticles in Cardiovascular Applications. *SMALL* 7, 705–717. <https://doi.org/10.1002/smll.201001347>
- Chandra, S., Das, P., Bag, S., Laha, D., Pramanik, P., 2011. Synthesis, functionalization and bioimaging applications of highly fluorescent carbon nanoparticles. *NANOSCALE* 3, 1533–1540. <https://doi.org/10.1039/c0nr00735h>

Рад у истакнутом међународном часопису (M22):

1. Sumarac-Dumanovic M, Jeremic D, **Pantovic A**, Janjetovic K, Stamenkovic-Pejkovic D, Cvijovic G, Stevanovic D, Micic D, Trajkovic V. Therapeutic improvement of glucoregulation in newly diagnosed type 2 diabetes patients is associated with a reduction of IL-17 levels. *Immunobiology*. 2013;218(8):1113-8. IF 3.180 (8 бодова)

Хетероцитата 31:

- McCarty, M.F., 2014. AMPK activation-protean potential for boosting healthspan. *AGE* 36, 641–663. <https://doi.org/10.1007/s11357-013-9595-y>

Strissel, K.J., Denis, G. V, Nikolajczyk, B.S., 2014. Immune regulators of inflammation in obesity-associated type 2 diabetes and coronary artery disease. *CURRENT OPINION IN ENDOCRINOLOGY DIABETES AND OBESITY* 21, 330–338. <https://doi.org/10.1097/MED.0000000000000085>

Kumar, P., Subramaniam, G., 2015. Molecular underpinnings of Th17 immune-regulation and their implications in autoimmune diabetes. *CYTOKINE* 71, 366–376. <https://doi.org/10.1016/j.cyto.2014.10.010>

Romanowski, M., Domanski, L., Pawlik, A., Osekowska, B., Dziedziejko, V., Safranow, K., Ciechanowski, K., 2015. Interleukin-17 gene polymorphisms in patients with post-transplant diabetes mellitus. *EUROPEAN REVIEW FOR MEDICAL AND PHARMACOLOGICAL SCIENCES* 19, 3152–3156.

Cortez-Espinosa, N., Cortes-Garcia, J.D., Martinez-Leija, E., Rodriguez-Rivera, J.G., Barajas-Lopez, C., Gonzalez-Amaro, R., Portales-Perez, D.P., 2015. CD39 expression on Treg and Th17 cells is associated with metabolic factors in patients with type 2 diabetes. *HUMAN IMMUNOLOGY* 76, 622–630. <https://doi.org/10.1016/j.humimm.2015.09.007>

Olson, N.C., Doyle, M.F., de Boer, I.H., Huber, S.A., Jenny, N.S., Kronmal, R.A., Psaty, B.M., Tracy, R.P., 2015. Associations of Circulating Lymphocyte Subpopulations with Type 2 Diabetes: Cross-Sectional Results from the Multi-Ethnic Study of Atherosclerosis (MESA). *PLOS ONE* 10. <https://doi.org/10.1371/journal.pone.0139962>

Francisco, C.O., Catai, A.M., Moura-Tonello, S.C.G., Arruda, L.C.M., Lopes, S.L.B., Benze, B.G., Del Vale, A.M., Malmegrim, K.C.R., Leal, A.M.O., 2016. Cytokine profile and lymphocyte subsets in type 2 diabetes. *BRAZILIAN JOURNAL OF MEDICAL AND BIOLOGICAL RESEARCH* 49. <https://doi.org/10.1590/1414-431X20155062>

Liu, S.L., Lin, Y., Liu, X., 2016. Protective effects of SIRT1 in patients with proliferative diabetic retinopathy via the inhibition of IL-17 expression. *EXPERIMENTAL AND THERAPEUTIC MEDICINE* 11, 257–262. <https://doi.org/10.3892/etm.2015.2877>

Wang, R., Solt, L.A., 2016. Metabolism of murine T(H)17 cells: Impact on cell fate and function. *EUROPEAN JOURNAL OF IMMUNOLOGY* 46, 807–816. <https://doi.org/10.1002/eji.201545788>

Santalathi, K., Maksimow, M., Airola, A., Pahikkala, T., Hutri-Kahonen, N., Jalkanen, S., Raitakari, O.T., Salmi, M., 2016. Circulating Cytokines Predict the Development of Insulin Resistance in a Prospective Finnish Population Cohort. *JOURNAL OF CLINICAL ENDOCRINOLOGY & METABOLISM* 101, 3361–3369. <https://doi.org/10.1210/jc.2016-2081>

Greer, R.L., Dong, X.X., Moraes, A.C.F., Zielke, R.A., Fernandes, G.R., Peremyslova, E., Vasquez-Perez, S., Schoenborn, A.A., Gomes, E.P., Pereira, A.C., Ferreira, S.R.G., Yao, M., Fuss, I.J., Strober, W., Sikora, A.E., Taylor, G.A., Gulati, A.S., Morgun, A., Shulzhenko, N., 2016. Akkermansia muciniphila mediates negative effects of IFN gamma on glucose metabolism. *NATURE COMMUNICATIONS* 7. <https://doi.org/10.1038/ncomms13329>

Susilowati, R., Sulistyoningrum, D.C., Witari, N.P.D., Huriyati, E., Luglio, H.F., Julia, M., 2016. Sexual dimorphism in interleukin 17A and adipocytokines and their association with insulin resistance among obese adolescents in Yogyakarta, Indonesia. *ASIA PACIFIC JOURNAL OF CLINICAL NUTRITION* 25, S93–S101. <https://doi.org/10.6133/apjcn.122016.s13>

Owczarczyk-Saczonek, A., Placek, W., 2017. Interleukin-17 as a factor linking the pathogenesis of psoriasis with metabolic disorders. *INTERNATIONAL JOURNAL OF DERMATOLOGY* 56, 260–268. <https://doi.org/10.1111/ijd.13420>

Brahimaj, A., Ligthart, S., Ghanbari, M., Ikram, M.A., Hofman, A., Franco, O.H., Kavousi, M., Dehghan, A., 2017. Novel inflammatory markers for incident pre-diabetes and type 2 diabetes: the Rotterdam Study. *EUROPEAN JOURNAL OF EPIDEMIOLOGY* 32, 217–226. <https://doi.org/10.1007/s10654-017-0236-0>

- Takeuchi, M., Sato, T., Sakurai, Y., Taguchi, M., Harimoto, K., Karasawa, Y., Ito, M., 2017. Association between aqueous humor and vitreous fluid levels of Th17 cell-related cytokines in patients with proliferative diabetic retinopathy. *PLOS ONE* 12. <https://doi.org/10.1371/journal.pone.0178230>
- Touch, S., Clement, K., Andre, S., 2017. T Cell Populations and Functions Are Altered in Human Obesity and Type 2 Diabetes. *CURRENT DIABETES REPORTS* 17. <https://doi.org/10.1007/s11892-017-0900-5>
- Phoksawat, W., Jumnainsong, A., Leelayuwat, N., Leelayuwat, C., 2017. Aberrant NKG2D expression with IL-17 production of CD4+T subsets in patients with type 2 diabetes. *IMMUNOBIOLOGY* 222, 944–951. <https://doi.org/10.1016/j.imbio.2016.05.001>
- Amersfoort, J., Kuiper, J., 2017. T cell metabolism in metabolic disease-associated autoimmunity. *IMMUNOBIOLOGY* 222, 925–936. <https://doi.org/10.1016/j.imbio.2017.03.001>
- Azuma, M.M., Gomes, J.E., Prieto, A.K.C., Samuel, R.O., de Lima, V.M.F., Sumida, D.H., Ervolino, E., Cintra, L.T.A., 2017. Diabetes increases interleukin-17 levels in periapical, hepatic, and renal tissues in rats. *ARCHIVES OF ORAL BIOLOGY* 83, 230–235. <https://doi.org/10.1016/j.archoralbio.2017.08.001>
- Nahomi, R.B., Sampathkumar, S., Myers, A.M., Elghazi, L., Smith, D.G., Tang, J., Lee, C.A., Kern, T.S., Nagaraj, R.H., Fort, P.E., 2018. The Absence of Indoleamine 2,3-Dioxygenase Inhibits Retinal Capillary Degeneration in Diabetic Mice. *INVESTIGATIVE OPHTHALMOLOGY & VISUAL SCIENCE* 59, 2042–2053. <https://doi.org/10.1167/iovs.17-22702>
- Ursini, F., Russo, E., Pellino, G., D'Angelo, S., Chiaravalloti, A., De Sarro, G., Manfredini, R., De Giorgio, R., 2018. Metformin and Autoimmunity: A “New Deal” of an Old Drug. *FRONTIERS IN IMMUNOLOGY* 9. <https://doi.org/10.3389/fimmu.2018.01236>
- Van Raemdonck, K., Umar, S., Szekecz, Z., Zomorodi, R.K., Shahrara, S., 2018. Impact of obesity on autoimmune arthritis and its cardiovascular complications. *AUTOIMMUNITY REVIEWS* 17, 821–835. <https://doi.org/10.1016/j.autrev.2018.02.007>
- Wang, M., Chen, F.Q., Wang, J.L., Zeng, Z.X., Yang, Q., Shao, S.Y., 2018. Th17 and Treg lymphocytes in obesity and Type 2 diabetic patients. *CLINICAL IMMUNOLOGY* 197, 77–85. <https://doi.org/10.1016/j.clim.2018.09.005>
- Porta, M., Amione, C., Barutta, F., Fornengo, P., Merlo, S., Gruden, G., Albano, L., Ciccarelli, M., Ungaro, P., Durazzo, M., Beguinot, F., Berchialla, P., Cavallo, F., Trento, M., 2019. The co-activator-associated arginine methyltransferase 1 (CARM1) gene is overexpressed in type 2 diabetes. *ENDOCRINE* 63, 284–292. <https://doi.org/10.1007/s12020-018-1740-z>
- Borzouei, S., Sheikh, V., Ghasemi, M., Zamani, A., Telikani, Z., Zareighane, Z., Salehi, I., Mozayanimonfared, A., Amirzargar, M.A., Alahgholi-Hajibehzad, M., 2019. Anti-Inflammatory Effect of Combined Sitagliptin and Vitamin D3 on Cytokines Profile in Patients with Type 2 Diabetes Mellitus. *JOURNAL OF INTERFERON AND CYTOKINE RESEARCH* 39, 293–301. <https://doi.org/10.1089/jir.2018.0144>
- Jennings, L., Hambly, R., Hughes, R., Moriarty, B., Kirby, B., 2020. Metformin use in hidradenitis suppurativa. *JOURNAL OF DERMATOLOGICAL TREATMENT* 31, 261–263. <https://doi.org/10.1080/09546634.2019.1592100>
- Ahmadi, M., Mahrooz, A., Abediankenari, S., Roodbari, N.H., 2020. Association of rs11558471 in SLC30A8 Gene with Interleukin 17 Serum Levels and Insulin Resistance in Iranian Patients with Type 2 Diabetes. *IRANIAN JOURNAL OF IMMUNOLOGY* 17, 215–225. <https://doi.org/10.22034/iji.2020.85513.1715>
- Michalak-Stoma, A., Bartosinska, J., Kowal, M., Raczkiwicz, D., Krasowska, D., Chodorowska, G., 2020. IL-17A in the Psoriatic Patients' Serum and Plaque Scales as Potential Marker of the Diseases Severity and Obesity. *MEDIATORS OF INFLAMMATION* 2020. <https://doi.org/10.1155/2020/7420823>

Wang, S., Pan, L.F., Gao, L., Qin, X.Y., Wang, J.N., 2021. Randomized research on the mechanism of local oxygen therapy promoting wound healing of diabetic foot based on RNA-seq technology. ANNALS OF PALLIATIVE MEDICINE 10, 973–983. <https://doi.org/10.21037/apm-20-295>

Croce, S., Avanzini, M.A., Regalbuto, C., Cordaro, E., Vinci, F., Zuccotti, G., Calcaterra, V., 2021. Adipose Tissue Immunomodulation and Treg/Th17 Imbalance in the Impaired Glucose Metabolism of Children with Obesity. CHILDREN-BASEL 8. <https://doi.org/10.3390/children8070554>

SantaCruz-Calvo, S., Bharath, L., Pugh, G., SantaCruz-Calvo, L., Lenin, R.R., Lutshumba, J., Liu, R., Bachstetter, A.D., Zhu, B.B., Nikolajczyk, B.S., 2022. Adaptive immune cells shape obesity-associated type 2 diabetes mellitus and less prominent comorbidities. NATURE REVIEWS ENDOCRINOLOGY 18, 23–42. <https://doi.org/10.1038/s41574-021-00575-1>

7. КВАЛИТЕТ НАУЧНИХ РЕЗУЛТАТА

Др сц. мед. Александар Пантовић публиковао је значајне радове на пољу неуронауке, молекуларне медицине и клиничке неурологије. Већина радова је публикован у међународним часописима. Током научноистраживачког рада кандидат је исказао познавање научноистраживачке методологије. Учествовао је у свим фазама научноистраживачког процеса, од дизајнирања истраживања па све до публиковања рада. У неколико радова заузима прво или друго место у списку коаутора, а такође је у неколико радова и аутор за кореспонденцију.

Од почетка научног и клиничког рада, др Пантовић је објавио 29 публицистичких јединица, од тога 10 радова штампаних у целини у међународним часописима (7×M21 + 2×M22 + 1×M23). Збирни импакт фактор радова које је др Пантовић до сада публиковао је 39,583 (као први аутор збирни импакт фактор је 12,841). Просечан број коаутора у радовима др Пантовића публикованим у целиности износи осам.

Др Пантовић је од почетка научног и клиничког рада објавио 12 радова in extenso, при чему је на 6 био први аутор. На основу захтева Правилника о избору у звање збир од потребних 10 у категоријама M10+M20+M31+M32+M33+M41+M42 износи 72.5 (нормирано 54.04), односно, од потребних 6 у категоријама M11+M12+M21+M22+M23, збир износи 71 (нормирано 52,55).

Др Пантовић је до сада цитиран 901 пута (хетероцитати): Scopus 901; Web of Science 850

Табела 3. Укупне вредности М коефицијента кандидата од почетка каријере према категоријама прописаним у Правилнику за област природно-математичких и медицинских наука.

КРИТЕРИЈУМИ МИНИСТАРСТВА		РЕЗУЛТАТИ КАНДИДАТА		
			Укупно	НББ
УКУПНО	16	УКУПНО	85	66,55
M10+M20+M31+M32+M33+M41+M42	10	M10+M20+M31+M32+M33+M41+M42	72,50	54,04
M11+M12+M21+M22+M23	6	M11+M12+M21+M22+M23	71	52,55

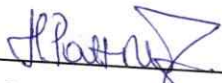
8. ЗАВРШНО МИШЉЕЊЕ И ПРЕДЛОГ КОМИСИЈЕ

Истраживачки рад др сц. мед. Александра Пантовића је везан за област молекуларне медицине и неурологије. Од почетка каријере, кандидат је објавио укупно 29 публицистичких јединица, од тога је 10 радова штампано у целини, са укупним фактором утицаја од 39,53 и са преко 900 хетероцитата.

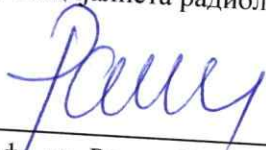
Приказани резултати научног рада указују да је др сц. мед. Александар Пантовић својим истраживањима допринео развоју научне области којом се бави. Посебно треба истаћи да је део резултата његових истраживања по први пут објашњава цитотоксичне механизме појединих нестероидних антиинфламаторних лекова на ћелије глиома и молекуларне механизме активације и улогу процеса аутофагије у диференцијацији матичних ћелија. Радови др сц. мед. Александра Пантовића су објављени и у значајним међународним часописима. Током свог истраживачког рада, др сц. мед. Александар Пантовић је био самосталан истраживач, способан да осмисли и реализује истраживање. Др сц. мед. Пантовић је објавио и неколико клиничких приказа случајева из области неурологије у домаћим и страним часописима.

Узимајући у обзир квантитет и квалитет публикованих резултата као и остале квалитативне показатеље успеха у научном раду, Комисија сматра да кандидат испуњава све законом прописане критеријуме за стицање научног звања научни сарадник, донетих од стране Министарства просвете, науке и технолошког развоја Републике Србије, те са великим задовољством предлаже Научном већу Војномедицинске академије да усвоји извештај и предлог Комисије да се др сц. мед. Александар Пантовић, специјалиста неурологије, изабере у звање научни сарадник.

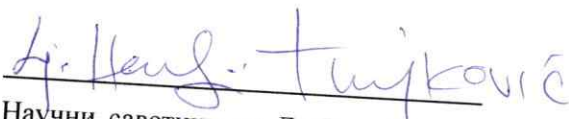
ЧЛАНОВИ КОМИСИЈЕ:



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У Београду, 27.10.2023.

РЕПУБЛИКА СРБИЈА
МИНИСТАРСТВО ОДБРАНЕ
УНИВЕРЗИТЕТ ОДБРАНЕ
Медицински факултет ВМА

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